UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL PROGRAMA DE PÓS-GRADUAÇÃO EM CIÊNCIAS MÉDICAS: ENDOCRINOLOGIA

ATIVIDADE INFLAMATÓRIA INDUZIDA PELA MORTE ENCEFÁLICA NO TECIDO PANCREÁTICO HUMANO

TESE DE DOUTORADO

TATIANA HELENA RECH

Rech, Tatiana
Atividade inflamatória induzida pela morte
encefálica no tecido pancreático humano / Tatiana
Rech. -- 2012.
93 f.

Orientadora: Cristiane Leitão. Coorientadora: Daisy Moreira.

Tese (Doutorado) -- Universidade Federal do Rio Grande do Sul, Faculdade de Medicina, Programa de Pós-Graduação em Ciências Médicas: Endocrinologia, Porto Alegre, BR-RS, 2012.

1. Transplante de ilhotas pancreáticas . 2. Morte encefálica. 3. Diabetes tipo 1 . 4. Inflamação. 5. Doação de múltiplos órgãos. I. Leitão, Cristiane, orient. II. Moreira, Daisy, coorient. III. Título.

Elaborada pelo Sistema de Geração Automática de Ficha Catalográfica da UFRGS com os dados fornecidos pelo(a) autor(a).

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Orientadores: Profa. Dra. Cristiane Bauermann Leitão

Dra. Daisy Crispim Moreira

Tese de Doutorado apresentada ao Programa de Pós-Graduação em Ciências Médicas: Endocrinologia da Universidade Federal do Rio Grande do Sul (UFRGS) como requisito parcial para a obtenção do título de Doutor em Endocrinologia.

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DEDICATÓRIA

À minha irmã Kárin.

AGRADECIMENTOS

Às minhas orientadoras, Profa. Dra. Cristiane Bauermann Leitão e Dra. Daisy Crispim Moreira, pelo enorme conhecimento científico, seriedade, entusiasmo e determinação com que desempenham suas funções. O Laboratório de Biologia das Ilhotas Pancreáticas Humanas é o resultado bem sucedido dessas qualidades. Vocês tornaram meu trabalho um prazer e ensinaram-me que é possível fazer pesquisa séria e de qualidade. Muito obrigada.

À minha grande amiga Cristiane Bauermann Leitão, de quem me orgulho ainda mais do que da pesquisadora, e que me honra com sua amizade há muitos anos.

Às famílias dos doadores de órgãos, que, em um momento de tanta dor, forneceram tão nobre material para a nossa pesquisa. Meus agradecimentos também aos pacientes controles.

À equipe da Comissão Intra-Hospitalar de Doação de Órgãos e Tecidos para Transplante (CIHDOTT) do Hospital de Clínicas de Porto Alegre (HCPA), que sempre colaborou, muito gentilmente, para a inclusão dos doadores de pâncreas nesta pesquisa.

Aos meus colegas intensivistas do Centro de Tratamento Intensivo do HCPA, pela compreensão em relação às minhas ausências durante as tardes desses últimos meses.

À funcionária Flávia, do Centro de Pesquisa Experimental do HCPA, pela disposição em ajudar e pela alegria que enche de som o laboratório.

À aluna de iniciação científica Sabrina Sigal Barkan e à biomédica Jakeline Reinheimer, pela pacienciosa ajuda na aquisição dos dados deste trabalho.

Ao meu marido Tiago, por acreditar em mim como pesquisadora.

À minha filha Marina, que antes mesmo de nascer já participava das madrugadas desta pesquisa.

E, como em todas as minhas vitórias, aos meus pais.

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LISTA DE ABREVIATURAS

ATP Adenosina trifosfato

BD Brain death

BMI Body mass index

cDNA Complementary deoxyribonucleic acid

CI *Confidence interval*

CIHDOTT Comissão Intra-Hospitalar de Doação de Órgãos e Tecidos para

Transplante

DM Diabetes melito ou *diabetes mellitus*

ELISA Enzyme-linked immunosorbent assay

ERK Cinase extracelular sinal-regulada

FIPE Fundo de Incentivo à Pesquisa e Ensino

FT Fator tecidual

GLP-1 *Glucagon-like peptide 1*

GRADE Grading of Recommendations Assessment, Development and

Evaluation

HbA1c Glycated hemoglobin

HCPA Hospital de Clínicas de Porto Alegre

HLA Human leukocyte antigen

IBMIR *Instant blood-mediated inflammatory reaction*

IFN-γ *Interferon-*γ

IL-1β Interleukin-1β

IL-10 Interleucina-10

IL-6 *Interleukin-6* e interleucina-6

IκB Inibidor κB

ME Morte encefálica

MeSH Medical Subject Headings

mRNA Messenger ribonucleic acid

NF-κB *Nuclear factor-κB* e fator nuclear-κB

PRISMA Preferred Reporting Items for Systematic Reviews and Meta-Analyses

RCT Randomized clinical trial

RNA Ribonucleic acid

RR Relative risk

RT-qPCR Reverse transcription quantitative polymerase chain reaction

SD Standard deviation

TF Tissue factor

TNF- α Tumor necrosis factor- α e fator de necrose tumoral- α

UCP-2 Proteína desacopladora 2

UFRGS Universidade Federal do Rio Grande do Sul

WMD Weighted mean difference

Esta tese de doutorado será apresentada no formato exigido pelo Programa de Pós-Graduação em Ciências Médicas: Endocrinologia. Ela será constituída de uma introdução em português e de dois artigos em inglês, estes formatados conforme as exigências das respectivas revistas médicas às quais serão submetidos para avaliação e posterior publicação. Os artigos em inglês desta tese são um artigo do tipo Revisão Sistemática e Meta-Análise e outro do tipo Artigo Original.

RESUMO

O transplante de ilhotas pancreáticas restabelece a secreção de insulina em pacientes diabéticos tipo 1 lábil. A despeito dos avanços da técnica de isolamento de ilhotas, a inabilidade de se obter um número suficiente de células de um único doador persiste como um obstáculo para o sucesso desse transplante. A identificação dos fatores relacionados com o dano das ilhotas pancreáticas durante todo o procedimento do transplante tem sido buscada na tentativa de desenvolver terapias capazes de minimizar a perda de células e otimizar a enxertia das ilhotas transplantadas, reduzindo, assim, a necessidade de múltiplos doadores para o alcance da independência de insulina. A morte encefálica (ME) está associada a uma inflamação sistêmica que produz profundas alterações fisiológicas na condição hemodinâmica do doador, antes mesmo do início do processo de retirada dos órgãos. Essa característica única do doador cadavérico influencia negativamente a função dos órgãos pós-transplante, o que torna os cuidados com o doador uma peça chave no cenário dos transplantes de órgãos. Contudo, o uso de protocolos com terapias específicas, como a reposição de hormônios tireoidiano e suprarrenal, tem demonstrado eficácia muito limitada em melhorar os desfechos de órgãos transplantados. Os resultados desta pesquisa demonstram que marcadores inflamatórios estão aumentados no estado de ME. A precipitação da ME é seguida de um aumento das concentrações de fator de necrose tumoral-α (TNF-α) e interleucina-6 (IL-6) no sangue e de TNF-α no tecido pancreático, mas não de fator tecidual (FT), cuja expressão já conhecida em ilhotas isoladas provavelmente se deva a fatores de estresse

relacionados ao isolamento das células. Esse aumento de TNF- α pode explicar, pelo menos em parte, os melhores desfechos alcançados por protocolos de transplante de ilhotas que incluem o uso de receptores solúveis do TNF- α . Em conclusão, a ME está associada a um aumento da expressão de TNF- α no sangue e no tecido pancreático humano. Portanto, o uso de terapias anti-inflamatórias dirigidas ao doador de múltiplos órgãos pode tornar-se uma estratégia promissora para melhorar os resultados dos transplantes de ilhotas.

CAPÍTULO 1

Introdução

O diabetes melito (DM) é uma síndrome caracterizada por alterações metabólicas associadas a elevada morbidade e mortalidade e que atinge milhões de pessoas em todo o mundo. O DM tipo 1, responsável por 5 a 10% dos casos de diabetes, é uma doença autoimune resultante da destruição das células β pancreáticas, o que determina a deficiência total da produção de insulina e a necessidade de administração de insulina exógena para a sobrevivência (1).

O transplante de pâncreas é, no momento, a maneira mais eficaz de se restabelecer a homeostase glicêmica em pacientes diabéticos tipo 1 com controle metabólico instável (2). O transplante de pâncreas como órgão inteiro promove controle glicêmico adequado e reduz as complicações crônicas do diabetes. Além disso, os pacientes submetidos ao transplante apresentam uma boa sobrevida a longo prazo (3). No entanto, esse procedimento está associado à morbidade de uma cirurgia de grande porte. Nesse cenário, a ideia da substituição do pâncreas endócrino deficiente por células produtoras de insulina através do transplante de ilhotas pancreáticas foi introduzida por Lacy em 1960. Seus relatos pioneiros demonstraram que ratos diabéticos ficavam normoglicêmicos após serem submetidos a transplante de ilhotas pancreáticas (4). Ao longo das últimas décadas, o aperfeiçoamento das técnicas de isolamento de ilhotas avançou muito (5), e, no ano 2000, o transplante de ilhotas se consolidou como uma opção de tratamento para pacientes com diabetes tipo 1 com

controle instável, com base nos estudos de Shapiro *et al.*, que propuseram um protocolo de imunossupressão livre de corticosteroides (6). A qualidade de vida é afetada positivamente pelo transplante (7), e a percepção da hipoglicemia é restaurada (8), promovendo a estabilização do controle glicêmico e a prevenção de hipoglicemias graves.

Em relação ao transplante de órgão inteiro, o transplante de ilhotas tem a grande vantagem de ser menos invasivo, uma vez que a injeção das células é feita através da canulação percutânea da veia porta (9). Por outro lado, um controle glicêmico adequado pós-transplante exige que um grande número de ilhotas seja transplantado. Frequentemente, são necessários transplantes sequenciais de dois ou mais pâncreas para se atingir a independência de insulina (10, 11). A escassez de órgãos para transplante acaba, então, sendo um forte limitador dessa terapia (12, 13). Em razão disso, vêm-se estudando exaustivamente maneiras de atingir o máximo aproveitamento de ilhotas por pâncreas doado (14, 15). É preciso ainda muito desenvolvimento nesse campo para que o transplante de ilhotas se torne a terapia padrão para o tratamento do diabetes tipo 1. O grande objetivo dessa terapia é o alcance da independência de insulina com apenas um doador (3).

Durante o processo de retirada e estocagem do pâncreas, as ilhotas são submetidas a múltiplos fatores deletérios celulares, entre eles a isquemia fria, as mudanças súbitas de temperatura, o estresse oxidativo, as forças de cisalhamento que agem sobre o órgão, além do processo de digestão necessário ao isolamento das células (16, 17). A esses agravos soma-se o intenso estresse inflamatório produzido pela morte encefálica (ME), resultando em lesão tecidual e na redução da função e da sobrevida dos enxertos (18).

A maior fonte de órgãos para transplante e a única fonte substancial de pâncreas é o doador cadavérico em ME (19). O doador ideal de ilhotas pancreáticas é um homem jovem, vítima de trauma, com índice de massa corporal $\geq 25~{\rm kg/m^2}$, controle glicêmico adequado, curto período de internação em unidade de tratamento intensivo e sem instabilidade hemodinâmica prolongada (20-23).

A ME é uma síndrome inflamatória com efeitos adversos graves bem definidos sobre os desfechos dos transplantes. A influência não imunológica da ME sobre os órgãos captados foi inicialmente estudada no rim. Enxertos renais de ratos receptores de rins cadavéricos apresentaram um curso mais acelerado de rejeição crônica do que os daqueles de doadores vivos (24). Mesmo rins de doadores vivos sem compatibilidade HLA (*human leukocyte antigen*) apresentaram uma sobrevida maior do que rins de doadores em ME em estudo bem desenhado com 368 transplantes entre cônjuges (25).

Órgãos provenientes de qualquer doador vivo, relacionado ou não relacionado, demonstram resultados consistentemente superiores quando comparados aos de doadores cadavéricos. Uma lesão cerebral catastrófica leva a ME, e esta desencadeia alterações hemodinâmicas, neuro-humorais e imunológicas que afetam a qualidade dos órgãos (26). A liberação aguda maciça de catecolaminas, conhecida como tempestade autonômica, é consequência da herniação cerebral e é tanto mais intensa quanto maior for a velocidade de instalação da hipertensão intracraniana. Essa liberação explosiva de catecolaminas produz um aumento na expressão de citocinas nos órgãos sólidos, além de mediar a ativação do complemento (27). O gatilho inflamatório que afeta adversamente a função dos órgãos transplantados de uma maneira antígeno-independente foi bem documentado por Kasuka *et al.*, que quantificaram a expressão de fator de necrose tumoral-α (TNF-α), interleucina-1β (IL-1β) e interleucina-6 (IL-6) em cobaias submetidas a ME ou somente ventilação mecânica. Após 5 dias, ocorreu uma

densa infiltração dessas citocinas nos túbulos e glomérulos renais dos ratos em ME (28). Contreras *et al.*, por sua vez, demonstraram que a ativação de citocinas próinflamatórias tem um impacto importante e tempo-dependente na função das células β pancreáticas. Em ratos, a precipitação da ME é seguida de um aumento imediato das concentrações de TNF-α, IL-1β e IL-6, que induzem a disfunção e morte da célula β, principalmente por apoptose (29). Há evidências também da implicação do aumento da interleucina-10 (IL-10) e do interferon-γ (INF-γ) na apoptose de ilhotas nesse cenário (30, 31).

Pâncreas originados de doadores vivos estão associados com maiores taxas de pureza, viabilidade e funcionalidade das células pós-isolamento em modelos animais (32) e humanos (33). Os efeitos inflamatórios deletérios da ME sobre o isolamento e a funcionalidade das células β são muito relevantes, na medida em que determinam uma menor recuperação de ilhotas por pâncreas doado. Por outro lado, a independência de insulina depende do transplante de um grande número de células β.

Além da perda de ilhotas secundária ao estresse inflamatório induzido pela ME e pelo estresse oxidativo da estocagem e do isolamento (34), um percentual significativo de ilhotas é destruído imediatamente após a infusão das células na circulação porta. Esse fenômeno, chamado de reação inflamatória instantânea sangue-mediada (do inglês *instant blood-mediated inflammatory reaction*, IBMIR), caracteriza-se por uma intensa atividade pró-coagulante, com eventos deletérios trombótico-inflamatórios responsáveis pela perda precoce de células transplantadas (35). Esse estado pró-coagulante induzido pelo transplante de ilhotas pode culminar, inclusive, com trombose da veia porta. A expressão do fator tecidual (FT) tem sido implicada como o principal gatilho da IBMIR no transplante de ilhotas e de hepatócitos (36, 37).

O FT é uma glicoproteína de 47 kDa responsável por desencadear a formação do coágulo, transformando o fator VII em fator VIIa. O complexo FT-fator VIIa é pró-inflamatório. A expressão de FT pelas ilhotas provoca intensa reação inflamatória no momento em que elas entram em contato direto com sangue ABO-compatível e ocorre a ligação rápida de plaquetas na sua superfície. Existe uma associação entre a intensidade da expressão do FT nas ilhotas e a magnitude com que a IBMIR afeta a função das células transplantadas (38). A perda de ilhotas desencadeada pela IBMIR foi estimada em 50 a 60% do total de células em primatas não humanos (39). Um estudo pequeno sugeriu que o bloqueio do FT através do uso de anticorpos monoclonais específicos anti-FT poderia aumentar a taxa de sucesso do transplante de ilhotas humanas (40).

A escassez de órgãos é o principal fator limitante ao desenvolvimento dos transplantes como terapia viável (41, 42). Desta forma, o manejo do doador de múltiplos órgãos concretiza-se como um capítulo muito importante na medicina intensiva (43). Uma das razões principais para potenciais doadores não se tornarem doadores de fato é o suporte inadequado das funções vitais no período que permeia a ME (44). Sabe-se que o uso sistemático de protocolos de cuidados com o doador aumenta a taxa de captação de órgãos (45-47). Porém, não se sabe, de forma consistente, quais terapias especificamente estão associadas à melhora dos resultados.

Diante do exposto, esta tese tem dois objetivos:

- Determinar quais procedimentos adotados no manejo do doador de múltiplos órgãos em ME alteram a qualidade dos órgãos a serem enxertados, através de uma revisão sistemática e meta-análise da literatura;
- Avaliar se a ME está associada a aumento de marcadores pró-inflamatórios e pró-trombóticos no tecido pancreático humano, por meio de um estudo de casos e controles.

Referências

- 1. Kobayashi N. The current status of islet transplantation and its perspectives. The review of diabetic studies: RDS. 2008;5(3):136-43. Epub 2008/12/23.
- 2. Zaman F, Abreo KD, Levine S, Maley W, Zibari GB. Pancreatic transplantation: evaluation and management. J Intensive Care Med. 2004;19(3):127-39.
- 3. Vardanyan M, Parkin E, Gruessner C, Rodriguez Rilo HL. Pancreas vs. islet transplantation: a call on the future. Current opinion in organ transplantation. 2010;15(1):124-30. Epub 2009/12/17.
- 4. Lacy PE, Walker MM, Fink CJ. Perifusion of isolated rat islets in vitro. Participation of the microtubular system in the biphasic release of insulin. Diabetes. 1972;21(10):987-98.
- Ricordi C. Islet transplantation: a brave new world. Diabetes. 2003;52(7):1595-603.
 Epub 2003/06/28.
- 6. Shapiro AM, Lakey JR, Ryan EA, Korbutt GS, Toth E, Warnock GL, et al. Islet transplantation in seven patients with type 1 diabetes mellitus using a glucocorticoid-free immunosuppressive regimen. The New England journal of medicine. 2000;343(4):230-8. Epub 2000/07/27.
- 7. Tharavanij T, Betancourt A, Messinger S, Cure P, Leitao CB, Baidal DA, et al. Improved long-term health-related quality of life after islet transplantation. Transplantation. 2008;86(9):1161-7. Epub 2008/11/14.
- 8. Leitao CB, Tharavanij T, Cure P, Pileggi A, Baidal DA, Ricordi C, et al. Restoration of hypoglycemia awareness after islet transplantation. Diabetes care. 2008;31(11):2113-5. Epub 2008/08/14.
- 9. Ryan EA, Bigam D, Shapiro AM. Current indications for pancreas or islet transplant. Diabetes, obesity & metabolism. 2006;8(1):1-7. Epub 2005/12/22.

- Robertson RP. Successful islet transplantation for patients with diabetes--fact or fantasy? The New England journal of medicine. 2000;343(4):289-90. Epub 2000/07/27.
- 11. Kim SC, Han DJ, Kang CH, We YM, Back JH, Kim YH, et al. Analysis on donor and isolation-related factors of successful isolation of human islet of Langerhans from human cadaveric donors. Transplant Proc. 2005;37(8):3402-3.
- Rech TH, Rodrigues Filho EM. Manuseio do potencial doador de múltiplos órgãos.
 Rev Bras Ter Intensiva. 2007;19(2):197-204.
- Ridgway D, Manas D, Shaw J, White S. Preservation of the donor pancreas for whole pancreas and islet transplantation. Clin Transplant. 2010;24(1):1-19. Epub 2009/12/18.
- 14. Sakuma Y, Ricordi C, Miki A, Yamamoto T, Pileggi A, Khan A, et al. Factors that affect human islet isolation. Transplant Proc. 2008;40(2):343-5.
- 15. O'Gorman D, Kin T, Murdoch T, Richer B, McGhee-Wilson D, Ryan E, et al. The standardization of pancreatic donors for islet isolation. Transplant Proc. 2005;37(2):1309-10.
- 16. Armann B, Hanson MS, Hatch E, Steffen A, Fernandez LA. Quantification of basal and stimulated ROS levels as predictors of islet potency and function. Am J Transplant. 2007;7(1):38-47.
- 17. Hering BJ, Matsumoto I, Sawada T, Nakano M, Sakai T, Kandaswamy R, et al. Impact of two-layer pancreas preservation on islet isolation and transplantation. Transplantation. 2002;74(12):1813-6.
- Bugge JF. Brain death and its implications for management of the potential organ donor. Acta anaesthesiologica Scandinavica. 2009;53(10):1239-50. Epub 2009/08/18.

- 19. Nagata H, Matsumoto S, Okitsu T, Iwanaga Y, Noguchi H, Yonekawa Y, et al. Procurement of the human pancreas for pancreatic islet transplantation from marginal cadaver donors. Transplantation. 2006;82(3):327-31. Epub 2006/08/15.
- 20. Matsumoto I, Sawada T, Nakano M, Sakai T, Liu B, Ansite JD, et al. Improvement in islet yield from obese donors for human islet transplants. Transplantation. 2004;78(6):880-5.
- 21. Briones RM, Miranda JM, Mellado-Gil JM, Castro MJ, Gonzalez-Molina M, Cuesta-Munoz AL, et al. Differential analysis of donor characteristics for pancreas and islet transplantation. Transplant Proc. 2006;38(8):2579-81.
- 22. Hesse UJ, Berrevoet F, Pattyn P, Vanholder R, de Hemptinne B. Donor parameters of pancreas grafts processed for islet or beta-cell isolation and transplantation. Transplant Proc. 1997;29(4):2259.
- 23. Goto T, Tanioka Y, Sakai T, Matsumoto I, Kakinoki K, Tanaka T, et al. Successful islet transplantation from a single pancreas harvested from a young, low-BMI, non-heart-beating cadaver. Transplant Proc. 2005;37(8):3430-2.
- Pratschke J, Wilhelm MJ, Laskowski I, Kusaka M, Paz D, Tullius SG, et al. The influence of donor brain death on long-term function of renal allotransplants in rats.
 Transplant Proc. 2001;33(1-2):693-4.
- 25. Terasaki PI, Cecka JM, Gjertson DW, Takemoto S. High survival rates of kidney transplants from spousal and living unrelated donors. The New England journal of medicine. 1995;333(6):333-6. Epub 1995/08/10.
- 26. Pratschke J, Wilhelm MJ, Kusaka M, Basker M, Cooper DK, Hancock WW, et al. Brain death and its influence on donor organ quality and outcome after transplantation. Transplantation. 1999;67(3):343-8. Epub 1999/02/25.

- 27. Tjernberg J, Ekdahl KN, Lambris JD, Korsgren O, Nilsson B. Acute antibody-mediated complement activation mediates lysis of pancreatic islets cells and may cause tissue loss in clinical islet transplantation. Transplantation. 2008;85(8):1193-9. Epub 2008/04/24.
- 28. Kusaka M, Pratschke J, Wilhelm MJ, Ziai F, Zandi-Nejad K, Mackenzie HS, et al. Activation of inflammatory mediators in rat renal isografts by donor brain death. Transplantation. 2000;69(3):405-10. Epub 2000/03/08.
- 29. Contreras JL, Eckstein C, Smyth CA, Sellers MT, Vilatoba M, Bilbao G, et al. Brain death significantly reduces isolated pancreatic islet yields and functionality in vitro and in vivo after transplantation in rats. Diabetes. 2003;52(12):2935-42. Epub 2003/11/25.
- 30. Pirot P, Cardozo AK, Eizirik DL. Mediators and mechanisms of pancreatic betacell death in type 1 diabetes. Arquivos brasileiros de endocrinologia e metabologia. 2008;52(2):156-65. Epub 2008/04/29.
- 31. Takada M, Toyama H, Tanaka T, Suzuki Y, Kuroda Y. Augmentation of interleukin-10 in pancreatic islets after brain death. Transplant Proc. 2004;36(5):1534-6.
- 32. Eckhoff DE, Eckstein C, Smyth CA, Vilatoba M, Bilbao G, Rahemtulla FG, et al. Enhanced isolated pancreatic islet recovery and functionality in rats by 17beta-estradiol treatment of brain death donors. Surgery. 2004;136(2):336-45. Epub 2004/08/10.
- 33. Jung HS, Choi SH, Kim SJ, Lee KT, Lee JK, Jang KT, et al. A better yield of islet cell mass from living pancreatic donors compared with cadaveric donors. Clinical transplantation. 2007;21(6):738-43. Epub 2007/11/09.

- 34. Mohseni Salehi Monfared SS, Larijani B, Abdollahi M. Islet transplantation and antioxidant management: a comprehensive review. World J Gastroenterol. 2009;15(10):1153-61.
- 35. Jackson A, McWilliams C, Kaizer E, Chaussabel D, Glaser C, Noguchi H, et al. Gene expression profiling of human pancreatic islets undergoing a simulated process of instant blood-mediated inflammatory reaction. Transplantation proceedings. 2008;40(2):430-2. Epub 2008/04/01.
- 36. Beuneu C, Vosters O, Movahedi B, Remmelink M, Salmon I, Pipeleers D, et al. Human pancreatic duct cells exert tissue factor-dependent procoagulant activity: relevance to islet transplantation. Diabetes. 2004;53(6):1407-11. Epub 2004/05/27.
- 37. Stephenne X, Vosters O, Najimi M, Beuneu C, Dung KN, Wijns W, et al. Tissue factor-dependent procoagulant activity of isolated human hepatocytes: relevance to liver cell transplantation. Liver transplantation: official publication of the American Association for the Study of Liver Diseases and the International Liver Transplantation Society. 2007;13(4):599-606. Epub 2007/03/31.
- 38. Peppelenbosch MP, Spek CA. Type I diabetes: a role for tissue factor in pancreatic islet transplantation? Lancet. 2002;360(9350):1999-2000. Epub 2002/12/31.
- 39. Berman DM, Cabrera O, Kenyon NM, Miller J, Tam SH, Khandekar VS, et al. Interference with tissue factor prolongs intrahepatic islet allograft survival in a nonhuman primate marginal mass model. Transplantation. 2007;84(3):308-15. Epub 2007/08/19.
- 40. Moberg L, Johansson H, Lukinius A, Berne C, Foss A, Kallen R, et al. Production of tissue factor by pancreatic islet cells as a trigger of detrimental thrombotic reactions in clinical islet transplantation. Lancet. 2002;360(9350):2039-45. Epub 2002/12/31.

- 41. Wood KE, Becker BN, McCartney JG, D'Alessandro AM, Coursin DB. Care of the potential organ donor. The New England journal of medicine. 2004;351(26):2730-9. Epub 2004/12/24.
- 42. Westphal GA, Caldeira Filho M, Vieira KD, Zaclikevis VR, Bartz MCM. Guidelines for potential multiple organ donors (adult). Part I. Mechanical ventilation, endocrine metabolic management, hematologial and infectious aspects. Rev Bras Ter Intensiva. 2011;23(3):255-68.
- 43. Macdonald PS, Aneman A, Bhonagiri D, Jones D, O'Callaghan G, Silvester W, et al. A systematic review and meta-analysis of clinical trials of thyroid hormone administration to brain dead potential organ donors. Critical care medicine. 2012;40(5):1635-44. Epub 2012/04/19.
- 44. DuBose J, Salim A. Aggressive organ donor management protocol. Journal of intensive care medicine. 2008;23(6):367-75. Epub 2008/09/26.
- 45. Rosendale JD, Kauffman HM, McBride MA, Chabalewski FL, Zaroff JG, Garrity ER, et al. Hormonal resuscitation yields more transplanted hearts, with improved early function. Transplantation. 2003;75(8):1336-41. Epub 2003/04/30.
- 46. Rosendale JD, Kauffman HM, McBride MA, Chabalewski FL, Zaroff JG, Garrity ER, et al. Aggressive pharmacologic donor management results in more transplanted organs. Transplantation. 2003;75(4):482-7. Epub 2003/02/28.
- 47. Venkateswaran RV, Patchell VB, Wilson IC, Mascaro JG, Thompson RD, Quinn DW, et al. Early donor management increases the retrieval rate of lungs for transplantation. The Annals of thoracic surgery. 2008;85(1):278-86; discussion 86. Epub 2007/12/25.

CAPÍTULO 2

Management of the brain-dead organ donor: a systematic review and metaanalysis

Short title: Management of brain-dead organ donor

Rech T.H., MD, MsC;¹ Moraes R.B., MD, MsC;¹ Crispim D., PhD;² Czepielewski M.A., PhD;² Leitão C.B., PhD²

¹ Division of Intensive Care Medicine, Hospital de Clínicas de Porto Alegre (HCPA), Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, Brazil.

² Division of Endocrinology, HCPA, UFRGS, Porto Alegre, Brazil.

Corresponding author and reprint requests:

Tatiana Helena Rech

Division of Intensive Care Medicine, Hospital de Clínicas de Porto Alegre

Rua Ramiro Barcelos 2350, 13º andar, 90035-003, Porto Alegre-RS, Brazil.

E-mail: tatiana.rech@hotmail.com

Phone: + 55 51 3359 8127 Fax: + 55 51 3359 8777

Key words: Brain death; randomized clinical trials; directed tissue donation; tissue and organ procurement.

Abstract

The shortage of organs is a limitation for transplantation, making the care of potential organ donors an important issue. The present systematic review and meta-analysis was carried out to assess the efficacy of interventions to stabilize hemodynamics in braindead donors or improve organ function and outcomes of transplantation. Medline, Embase and Cochrane databases were searched. Of 5096 articles retrieved, 39 randomized clinical trials (RCTs) were selected. Twenty were included in a qualitative synthesis, providing data on 1277 patients. The main interventions described were desmopressin use, triiodothyronine and methylprednisolone replacement, fluid management, vasopressor therapy, mechanical ventilation strategies, and surgical techniques. Three meta-analyses were conducted: the first included two studies and showed that desmopressin administered to brain-dead patients was not advantageous with respect to early organ function in kidney recipients (RR 0.97; 95%CI 0.85-1.10; I² 0%, p=0.809). The second included four studies and showed that triiodothyronine did not add hemodynamic benefits vs. standard management (weighted mean difference 0.15; 95% CI -0.13-0.42; *I*² 17.4%, p=0.304). The third meta-analysis (two studies) showed that ischemic liver preconditioning during harvesting procedures did not benefit survival (RR 1.0; 95% CI 0.93-1.08; I² 0%, p=0.459). The present results suggest limited efficacy of interventions focusing on the management of brain-dead donors.

Introduction

Organ transplantation is the treatment of choice for many end-stage organ diseases. However, it is still strongly limited by organ shortage (1), with increasing disparity between organ supply and demand (2). One promising way to overcome this problem is to optimize brain-dead organ donation.

Brain death is an inflammatory syndrome (3, 4) that causes a massive catecholamine release, with a sudden decrease in cortisol, insulin, thyroid and pituitary hormone levels (5). Hormonal replacement therapy has been reported to stabilize and improve cardiac function in brain-dead donors (6, 7). Rosendale et al. (8) described a retrospective analysis of more than 10 thousand consecutive donors, suggesting that aggressive pharmacologic therapy results in more transplanted organs. However, other studies have failed to confirm these benefits (9, 10).

In addition, hormonal alterations have a profound hemodynamic and metabolic impact on potential donors, inducing a variety of deleterious effects that can threat organ perfusion and result in cardiac arrest (11). Frequently, hemodynamic collapse precludes organ donation (12). Therefore, early donor management is associated with increased organ retrieval (13), and many transplantation centers and critical care societies have developed standardized donor management protocols that focus on hemodynamic and hormonal resuscitation (14-16).

However, the strategy of brain-dead donor management is still controversial; it is time and resource consuming and has potential deleterious effects, such as metabolic acidosis. Therefore, the present systematic review and meta-analysis was carried out to assess the impact of interventions focusing on the care of brain-dead donors on transplantation outcomes.

Methods

Search strategy and study selection

To identify randomized clinical trials (RCTs) comparing any category of intervention on management of brain-dead organ donors, in August 2012 we performed an initial electronic literature search in MEDLINE, EMBASE and Cochrane as well as the Cochrane Controlled Trials Register, without language or date restriction, using the following medical subject headings (MeSH): "Tissue and Organ Procurement" [MeSH] or "Directed Tissue Donation" [MeSH] or "Brain Death" [MeSH]. A high sensitivity strategy for the search of RCTs was used (17). Additionally, we manually searched the references of the selected studies. This systematic review is reported in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) (18).

Eligibility criteria

We included RCTs comparing interventions aimed to stabilize hemodynamics in brain-dead donors or to improve donated organ function or organ receptor outcomes after transplantation as compared to a control group. The following were excluded: 1) RCTs that did not provide information regarding the associations of the intervention with donor stability, organ function or receptor's outcomes in the experimental group, the control group, or both; 2) duplicate publications or substudies of included trials.

Data extraction

Titles and abstracts of retrieved articles were independently evaluated by two reviewers (T.H.R. and R.B.M.). Disagreements were solved by consensus or by a third reviewer (C.B.L.). The investigators were not blinded to authors, institutions or

journals. Articles whose abstracts did not provide enough information regarding the inclusion and exclusion criteria were retrieved for full text evaluation. To avoid possible double counting of patients included in more than one report by the same authors or working groups, recruitment periods were evaluated. If necessary, the corresponding author was contacted for elucidation. Two reviewers (T.H.R. and R.B.M.) independently conducted data extraction.

Assessment of risk of bias and study quality

Risk of bias was evaluated according to GRADE (The Grading of Recommendations Assessment, Development and Evaluation) recommendations (19). Study quality assessment included adequate sequence generation, allocation concealment, blinding of outcomes assessment, and intention-to-treat analysis. Quality assessment was independently performed by two reviewers (T.H.R and R.B.M) and disagreements solved by consensus or a third reviewer (C.B.L).

Statistical analysis

The clinical outcomes of interest were hemodynamic parameters before transplantation, quantification of organ retrieval, organ function after transplantation, and patient or graft survival after transplantation. Studies reporting a similar intervention and outcome were grouped, and a separate forest plot was constructed whenever possible (similar intervention/outcomes in two or more studies identified during the search). For analysis of single studies (that could not be grouped), only qualitative assessment was performed.

For continuous variable outcomes, means or differences between means and respective dispersion values were extracted, and pooled-effect estimates were obtained

by comparing the least squares mean percentage change from baseline to the end of the study for each group; these results were expressed as the weighted mean difference between groups. For categorical outcomes, the total number of patients included and the number of participants with the outcome were used to calculate the overall relative risk (RR) of the intervention to improve an outcome.

Cochran's Q test was used to evaluate heterogeneity between studies, and a threshold p value= 0.1 was considered statistically significant. The I^2 test was also conducted to evaluate the magnitude of the heterogeneity between studies. We used risk estimates obtained with a fixed-effect meta-analysis because no significant heterogeneity was found between the studies. For the triiodothyronine meta-analysis, publication bias was assessed using a contour-enhanced funnel plot of each trial's effect size against the standard error (20). Funnel plot asymmetry was evaluated by Begg and Egger tests, and a significant publication bias was considered if the p value was less than 0.1 (21, 22). All statistical analyses were performed by Stata 11.0 software (Stata, College Station, TX, USA).

Results

Literature search results and study characteristics

We identified 5094 potentially relevant citations from electronic database search and two from manual search. After duplicates were removed, 2726 studies were screened on the basis of title and abstracts, resulting in 39 RCTs for further evaluation. Twenty studies fulfilled our inclusion criteria, providing data on 1277 patient (Table 1). Of these, 12 trials could not be grouped (single intervention and/or outcome) (11, 23-

33) and were evaluated qualitatively, and 8 studies (15, 16, 34-39) were included in the meta-analysis. A flowchart of search and selection criteria is shown in Figure 1.

The trials were published from 1977 to 2010. The main interventions included: desmopressin use (n=146 patients), triiodothyronine replacement (n=264 patients), methylprednisolone replacement (n=150 patients), triiodothyronine and methilprednisolone replacement (n=60 patients), fluid management (n=33 patients), vasopressor therapy (n=264 patients), mechanical ventilation strategies (n=118 patients), and surgical techniques (n=242 patients). The outcomes assessed varied substantially between studies, from hemodynamic parameters and organ function to graft and patient survival. There were no major treatment-associated adverse events.

Table 2 shows the risk of bias in each trial. Eight studies were blinded, seven used placebo controlled groups, nine trials described adequate sequence generation and allocation concealment, 15 used the intention-to-treat principle for statistical analysis, two declared to have received grant support from the pharmaceutical industry and three were stopped early, one because of harm (impaired immediate renal function of kidney recipients), one because of benefit, and the other because of termination of funding.

Trials included in qualitative-only analysis

Twelve studies (11, 23-33) reported interventions and/or outcomes not duplicated by other authors. Because it was not possible to group them, they were not therefore included in a meta-analysis. One of them, a well-designed multicenter RCT including 118 patients, demonstrated that more lungs were eligible for transplantation when a lung protective ventilator strategy with low tidal volumes was used as compared with a conventional ventilator strategy (30). Another trial, performed in 60 European centers, allocated brain-dead patients to pretreatment with low-dose dopamine or no

treatment. The results showed that early kidney graft function was improved with the pharmacological approach, but with no impact on patient survival (33). Two underpowered studies tested the role of fluid management of donors. When low molecular weight hydroxyethyl starch was administered to liver donors, no differences in early function were found (31). However, impaired immediate renal function in kidney recipients was reported when hydroxyethyl starch was used as plasma expander in kidney donors, leading to early termination of the trial (24).

The impact of methylprednisolone replacement on brain-dead donors was investigated in three different RCTs that evaluated distinct organ outcomes. Hormone therapy was not effective to either increase lung yield (13) or improve early kidney graft function (23). Only one study demonstrated a protective effect of methylprednisolone on liver grafts, with significant downregulation of inflammation markers and decreased incidence of acute rejection after liver transplantation (29).

Two out of six studies of thyroid hormone replacement therapy described study-specific outcomes and were not included in the meta-analysis. The first one measured the impact of triiodothyronine on liver function tests during the first week post-transplantation, obtaining similar results in both groups and worse metabolic acidosis in the treatment group (32). The other study showed a trend toward less inotrope need in the treatment group (28).

Circulatory deterioration and cardiac arrest usually occur soon after brain death. However, long-term maintenance of circulation of brain-dead donors was reported in a desmopressin-treated group when compared to a control group (27).

No difference regarding graft performance was found in a trial designed to compare two preservation solutions as the initial flush in hepatic allograft procurement (25). Finally, a trial of the impact of donor harvesting technique using a modified

double (aortic and portal) perfusion technique for suboptimal liver grafts was terminated early because of benefit, with improved six-month graft and patient survival rates when compared with a single aortic perfusion technique (26).

Trials included in meta-analyses

We retrieved eight studies that could be meta-analyzed, two evaluating desmopressin (n=121 patients), four intravenous triiodothyronine (n=209 patients), and two ischemic liver preconditioning (n=151 patients).

The two studies on desmopressin use (36, 37) assessed the effects of desmopressin administration to brain-dead donors on early graft function in kidney recipients. As shown in Figure 2A, no benefits of desmopressin on early graft function of kidney transplants were observed (RR = 0.97, CI = 0.85 – 1.10, I^2 = 0% and p for heterogeneity = 0.819).

Four trials allocated brain-dead patients to receive intravenous triiodothyronine or placebo and used cardiac index as outcome (9, 10, 38, 39). No differences in cardiac index were found between groups (difference between groups: 0.15, CI = -0.13 - 0.42 L/min/m², $I^2 = 17.4\%$ and p for heterogeneity = 0.304; Figure 2B). Funnel plot analysis did not show significant publication bias for the triiodothyronine intervention.

Figure 2C depicts the meta-analysis of two RCTs (34, 35) that assessed the effects of ischemic liver preconditioning during the donor harvesting procedures. No differences were observed in patient survival at 24-25 months (RR = 1.00, CI = 0.93 – 1.08, $I^2 = 0\%$ and p for heterogeneity = 0.459).

Discussion

Aggressive donor management protocols must rely on strong pathophysiological evidence. However, the present systematic review and meta-analyses did not find consistent support for recommending such strategies, especially hormonal replacement.

It has been suggested that the hemodynamic instability associated with brain death is in part a result of diminished levels of circulating thyroxine, leading to a reduction of myocardial energy stores and a shift from aerobic to anaerobic metabolism (7). Experimental studies demonstrated improved cardiac function following thyroid hormonal replacement therapy in brain-dead baboons (40). Many retrospective analyses suggest that thyroid hormonal replacement could improve cardiac function and increase the number of organs transplanted per donor (8, 41, 42). We retrieved six RCTs designed to evaluate the effects of triiodothyronine replacement to organ donors, and all of them had consistent negative results. Moreover, the pooled analysis of four RCTs (n=209 patients) evaluating cardiac index as outcome turned out to be negative. Another recent meta-analysis has evaluated the effect of triiodothyronine replacement on donor heart function, and no benefit was found (43). These inconsistent findings suggest that depletion of triiodothyronine (and the subsequent relative hypothyroid state) is not the major determinant of myocardial dysfunction in these patients, but rather perhaps only an adaptive response to illness. Similarly, the evidence in favor of the administration of triiodothyronine in another setting of adaptive relative hypothyroidism, that is, critical care, is far from compelling, to the point that some authors advise withholding its use in critically ill patients unless there is clear evidence of previous hypothyroidism (44).

The complex hemodynamic dysfunction related to brain death is frequently associated with major complications in the potential donor, and has multiple causes (1). Diabetes insipidus may be present in up to 80% of these patients, with severe

dehydration and hypovolemia (11). The use of desmopressin was not associated with better kidney graft outcomes in the present meta-analysis (36, 37), but it is safe and useful to limit the harmful effects of profuse polyuria, decreasing the need for large volume infusions and preventing hemodynamic collapse (45). Studies have suggested the use of colloid solutions as an option to avoid the infusion of large volumes to treat hypovolemia, since fluid overload could be deleterious to lung grafts (46). The only two RCTs retrieved by us dealing with fluid replacement did not support this notion. On the contrary, one study showed no difference between treatment groups of liver donors when hydroxyethyl starch was used compared to crystalloid solutions. The other study was terminated early because of harm to immediate renal function of kidney recipients (24, 31). To this point, there is no evidence supporting the use of hydroxyethyl starch in brain-dead or other critical care patients (47). Vasodilation and hypotension are almost always present in these patients, but no high level evidence for the choice of one or another vasopressor agent is available. Donor treatment with dopamine resulted in a reduction in dialysis requirement after kidney transplantation, with no clinically significant impact on graft or patient survival (33). This fact, taken together with the results of a recent meta-analysis of septic shock patients, would advise in favor of norepinephrine, because dopamine was associated with greater mortality and higher incidence of arrhythmias when compared to norepinephrine in this study (48).

Marginal livers, which have been used to increase the donor pool, are especially susceptible to ischemia-reperfusion injury. Ischemic preconditioning during harvesting is a strategy to prevent the deleterious effects of ischemia-reperfusion on the liver graft, probably by modulating the inflammatory response (49). Compared with standard orthotopic liver transplant, this strategy is associated with better tolerance to ischemia, but with no significant difference on patient survival (34, 35).

Brain death is associated with a profound pro-inflammatory process. Under these circumstances, a beneficial role of steroids could be expected, as suggested by retrospective studies (42, 50). However, RCTs using methylprednisolone in brain-dead donors contradict this hypothesis. Methylprednisolone neither increased lung yield (13), nor improved kidney function post-transplantation (23). The only positive effect was observed for a surrogate outcome: downregulation of inflammatory and apoptotic markers in liver biopsies (29). As brain-dead donors might have a relative adrenal insufficiency (51), another potential benefit of corticosteroid use is promotion of hemodynamic stability. However, only methylprednisolone, which lacks significant mineralocorticoid activity, has been evaluated in RCTs. The use of hydrocortisone or even fludrocortisones may result in better outcomes and should be evaluated in future RCTs.

The best evidence in the management of organ donor refers to mechanical ventilation. The use of lung protective strategies with low tidal volumes increases the yield of lungs when compared to conventional ventilatory strategies (30). High tidal volumes are known to be detrimental to patients with acute lung injury (52, 53), and prevention of overdistension seems to be beneficial to potential lung donors.

This study has several limitations. First, there are few RCTs dealing with the management of brain-dead donors. Second, the general quality of these studies was considered low according to GRADE guidelines (19), raising the possibility of bias. Third, the end points of various trials differed, and many of them had evaluated only surrogate hemodynamic end points as their primary outcomes, such as hemodynamic parameters and initial organ function. Besides, studies had reported no major-treatment-associated adverse events, raising the question if they were properly evaluated. However, similar results were obtained in a systematic review and meta-analysis

focusing specifically on clinical trials of thyroid hormone administration to brain-dead potential organ donors (43), which concludes that the use of thyroid hormone in marginal donors is based on low-level evidence.

It should be noted that the interventions found in this study to be ineffective to increase patient or organ survival when used alone are nevertheless recommended by international guidelines (1, 5, 14, 16, 54). However, we believe that a multi-intervention strategy protocol conducted by a dedicated senior physician at the bedside could produce more favorable outcomes. An early combined strategy holding the best choice of fluids, vasopressor drugs, mechanical ventilation parameters, surgical techniques and combined hormonal replacement therapy should be tested in a well-designed clinical trial.

In summary, despite the implementation of aggressive donor care protocols focusing on hemodynamic and hormonal resuscitation by many transplantation centers and critical care societies, these recommendations are weakly supported, with most evidence based on surrogate outcomes and retrospective data. We recognize the great importance of brain-dead donor care to improve transplantation outcomes, but this systematic review and meta-analysis did not provide consistent evidence for recommending this strategy. Therefore, further RCTs are required to elucidate to what extent a multi-intervention management strategy of brain-dead donors is helpful for transplant recipients.

Contributors

T.H.R participated in the study conception and design, data acquisition, analysis, and interpretation of data, drafting of the manuscript and revision of the manuscript.

R.B.M participated in data acquisition, analysis, and interpretation, drafting of the

manuscript and revision of the manuscript. D.C. and M.A.C. critically reviewed the manuscript for intellectual content. C.B.L participated in the study conception and design, analysis and interpretation of data, revision of the manuscript and statistical analysis.

Acknowledgements

This study was supported by Fundo de Incentivo à Pesquisa e Eventos (FIPE) from Hospital de Clínicas de Porto Alegre (Porto Alegre, RS, Brazil).

References

- 1. DuBose J, Salim A. Aggressive organ donor management protocol. J Intensive Care Med. 2008;23(6):367-75. Epub 2008/09/26.
- Joshi NR, Margulies DR. Aggressive organ donor management: more from less?
 Current Opin Organ Transplant. 2006;11(2):141-5.
- 3. Kusaka M, Pratschke J, Wilhelm MJ, Ziai F, Zandi-Nejad K, Mackenzie HS, et al. Activation of inflammatory mediators in rat renal isografts by donor brain death. Transplantation. 2000;69(3):405-10. Epub 2000/03/08.
- 4. Nijboer WN, Schuurs TA, van der Hoeven JA, Leuvenink HG, van der Heide JJ, van Goor H, et al. Effects of brain death on stress and inflammatory response in the human donor kidney. Transplant Proc. 2005;37(1):367-9. Epub 2005/04/06.
- 5. Wood KE, Becker BN, McCartney JG, D'Alessandro AM, Coursin DB. Care of the potential organ donor. The New England journal of medicine. 2004;351(26):2730-9. Epub 2004/12/24.
- 6. Novitzky D, Cooper DK, Chaffin JS, Greer AE, DeBault LE, Zuhdi N. Improved cardiac allograft function following triiodothyronine therapy to both donor and recipient. Transplantation. 1990;49(2):311-6. Epub 1990/02/01.
- 7. Salim A, Vassiliu P, Velmahos GC, Sava J, Murray JA, Belzberg H, et al. The role of thyroid hormone administration in potential organ donors. Arch Surg. 2001;136(12):1377-80. Epub 2001/12/26.
- 8. Rosendale JD, Kauffman HM, McBride MA, Chabalewski FL, Zaroff JG, Garrity ER, et al. Aggressive pharmacologic donor management results in more transplanted organs. Transplantation. 2003;75(4):482-7. Epub 2003/02/28.

- 9. Goarin JP, Cohen S, Riou B, Jacquens Y, Guesde R, Le Bret F, et al. The effects of triiodothyronine on hemodynamic status and cardiac function in potential heart donors. Anesth Analg. 1996;83(1):41-7. Epub 1996/07/01.
- 10. Mariot J, Jacob F, Voltz C, Perrier JF, Strub P. [Value of hormonal treatment with triiodothyronine and cortisone in brain dead patients]. Ann Fr Anesth Reanim. 1991;10(4):321-8. Epub 1991/01/01. Interet de l'hormonotherapie associant triiodothyronine et cortisone chez le patient en etat de mort cerebrale.
- 11. Rech TH, Rodrigues Filho ÉM. Manuseio do potencial doador de múltiplos órgãos. Rev Bras Ter Intensiva. 2007;19(2):197-204.
- 12. Bugge JF. Brain death and its implications for management of the potential organ donor. Acta Anaesthesiol Scand. 2009;53(10):1239-50. Epub 2009/08/18.
- 13. Venkateswaran RV, Patchell VB, Wilson IC, Mascaro JG, Thompson RD, Quinn DW, et al. Early donor management increases the retrieval rate of lungs for transplantation. The Annals of thoracic surgery. 2008;85(1):278-86; discussion 86. Epub 2007/12/25.
- 14. Zaroff JG, Rosengard BR, Armstrong WF, Babcock WD, D'Alessandro A, Dec GW, et al. Consensus conference report: maximizing use of organs recovered from the cadaver donor: cardiac recommendations, March 28-29, 2001, Crystal City, Va. Circulation. 2002;106(7):836-41. Epub 2002/08/15.
- 15. Rosendale JD, Chabalewski FL, McBride MA, Garrity ER, Rosengard BR, Delmonico FL, et al. Increased transplanted organs from the use of a standardized donor management protocol. American journal of transplantation: official journal of the American Society of Transplant Surgeons. 2002;2(8):761-8. Epub 2002/09/24.

- 16. Westphal GA, Caldeira Filho M, Vieira KD, Zaclikevis VR, Bartz MCM, Wanzuita R, et al. Guidelines for potential multiple organ donors (adult). Part II. Mechanical ventilation, endocrine metabolic management, hematological and infectious aspects Rev Bras Ter Intensiva. 2001;23(3):269-82.
- 17. Robinson KA, Dickersin K. Development of a highly sensitive search strategy for the retrieval of reports of controlled trials using PubMed. International journal of epidemiology. 2002;31(1):150-3. Epub 2002/03/27.
- 18. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. Ann Intern Med. 2009;151(4):264-9, W64. Epub 2009/07/23.
- 19. Balshem H, Helfand M, Schunemann HJ, Oxman AD, Kunz R, Brozek J, et al. GRADE guidelines: 3. Rating the quality of evidence. Journal of clinical epidemiology. 2011;64(4):401-6. Epub 2011/01/07.
- 20. Peters JL, Sutton AJ, Jones DR, Abrams KR, Rushton L. Contour-enhanced metaanalysis funnel plots help distinguish publication bias from other causes of asymmetry. Journal of clinical epidemiology. 2008;61(10):991-6. Epub 2008/06/10.
- 21. Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. BMJ. 2003;327(7414):557-60. Epub 2003/09/06.
- Duval S, Tweedie R. Trim and fill: A simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. Biometrics. 2000;56(2):455-63. Epub 2000/07/06.
- 23. Chatterjee SN, Terasaki PI, Fine S, Schulman B, Smith R, Fine RN. Pretreatment of cadaver donors with methylprednisolone in human renal allografts. Surg Gynecol Obstet. 1977;145(5):729-32. Epub 1977/11/01.

- 24. Cittanova ML, Leblanc I, Legendre C, Mouquet C, Riou B, Coriat P. Effect of hydroxyethylstarch in brain-dead kidney donors on renal function in kidney-transplant recipients. Lancet. 1996;348(9042):1620-2. Epub 1996/12/14.
- 25. Cofer JB, Klintmalm GB, Morris CV, Solomon H, Watemberg IA, Husberg BS, et al. A prospective randomized trial between Euro-Collins and University of Wisconsin solutions as the initial flush in hepatic allograft procurement. Transplantation. 1992;53(5):995-8. Epub 1992/05/01.
- 26. D'Amico F, Vitale A, Gringeri E, Valmasoni M, Carraro A, Brolese A, et al. Liver transplantation using suboptimal grafts: impact of donor harvesting technique. Liver Transpl. 2007;13(10):1444-50. Epub 2007/09/29.
- 27. Iwai A, Sakano T, Uenishi M, Sugimoto H, Yoshioka T, Sugimoto T. Effects of vasopressin and catecholamines on the maintenance of circulatory stability in brain-dead patients. Transplantation. 1989;48(4):613-7. Epub 1989/10/01.
- 28. Jeevanandam V. Triiodothyronine: spectrum of use in heart transplantation. Thyroid. 1997;7(1):139-45. Epub 1997/02/01.
- 29. Kotsch K, Ulrich F, Reutzel-Selke A, Pascher A, Faber W, Warnick P, et al. Methylprednisolone therapy in deceased donors reduces inflammation in the donor liver and improves outcome after liver transplantation: a prospective randomized controlled trial. Ann Surg. 2008;248(6):1042-50. Epub 2008/12/19.
- 30. Mascia L, Pasero D, Slutsky AS, Arguis MJ, Berardino M, Grasso S, et al. Effect of a lung protective strategy for organ donors on eligibility and availability of lungs for transplantation: a randomized controlled trial. JAMA. 2010;304(23):2620-7. Epub 2010/12/16.

- 31. Randell T, Orko R, Hockerstedt K. Peroperative fluid management of the brain-dead multiorgan donor. Acta Anaesthesiol Scand. 1990;34(7):592-5. Epub 1990/10/01.
- 32. Randell TT, Hockerstedt KA. Triiodothyronine treatment in brain-dead multiorgan donors--a controlled study. Transplantation. 1992;54(4):736-8. Epub 1992/10/01.
- 33. Schnuelle P, Gottmann U, Hoeger S, Boesebeck D, Lauchart W, Weiss C, et al. Effects of donor pretreatment with dopamine on graft function after kidney transplantation: a randomized controlled trial. JAMA. 2009;302(10):1067-75. Epub 2009/09/10.
- 34. Amador A, Grande L, Marti J, Deulofeu R, Miquel R, Sola A, et al. Ischemic preconditioning in deceased donor liver transplantation: a prospective randomized clinical trial. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2007;7(9):2180-9. Epub 2007/08/19.
- 35. Azoulay D, Del Gaudio M, Andreani P, Ichai P, Sebag M, Adam R, et al. Effects of 10 minutes of ischemic preconditioning of the cadaveric liver on the graft's preservation and function: the ying and the yang. Ann Surg. 2005;242(1):133-9. Epub 2005/06/24.
- 36. Guesde R, Barrou B, Leblanc I, Ourahma S, Goarin JP, Coriat P, et al. Administration of desmopressin in brain-dead donors and renal function in kidney recipients. Lancet. 1998;352(9135):1178-81. Epub 1998/10/20.
- Pennefather SH, Bullock RE, Mantle D, Dark JH. Use of low dose arginine vasopressin to support brain-dead organ donors. Transplantation. 1995;59(1):58-62. Epub 1995/01/15.

- 38. Perez-Blanco A, Caturla-Such J, Canovas-Robles J, Sanchez-Paya J. Efficiency of triiodothyronine treatment on organ donor hemodynamic management and adenine nucleotide concentration. Intensive Care Med. 2005;31(7):943-8. Epub 2005/06/07.
- 39. Venkateswaran RV, Steeds RP, Quinn DW, Nightingale P, Wilson IC, Mascaro JG, et al. The haemodynamic effects of adjunctive hormone therapy in potential heart donors: a prospective randomized double-blind factorially designed controlled trial. Eur Heart J. 2009;30(14):1771-80. Epub 2009/03/28.
- 40. Novitzky D, Wicomb WN, Cooper DKC, Rose AG, Fraser C, Barnard CN. Electrocardiographic, hemodynamic and endocrine changes occurring during experimental brain death in the chacma baboon. J Heart Transplant. 1984;4:63-9.
- 41. Novitzky D, Cooper DK, Reichart B. Hemodynamic and metabolic responses to hormonal therapy in brain-dead potential organ donors. Transplantation. 1987;43(6):852-4. Epub 1987/06/01.
- 42. Rosendale JD, Kauffman HM, McBride MA, Chabalewski FL, Zaroff JG, Garrity ER, et al. Hormonal resuscitation yields more transplanted hearts, with improved early function. Transplantation. 2003;75(8):1336-41. Epub 2003/04/30.
- 43. Macdonald PS, Aneman A, Bhonagiri D, Jones D, O'Callaghan G, Silvester W, et al. A systematic review and meta-analysis of clinical trials of thyroid hormone administration to brain dead potential organ donors. Crit Care Med. 2012;40(5):1635-44. Epub 2012/04/19.
- 44. Stathatos N, Levetan C, Burman KD, Wartofsky L. The controversy of the treatment of critically ill patients with thyroid hormone. Best Pract Res Clin Endocrinol Metab. 2001;15(4):465-78. Epub 2002/01/22.

- 45. Dictus C, Vienenkoetter B, Esmaeilzadeh M, Unterberg A, Ahmadi R. Critical care management of potential organ donors: our current standard. Clinical transplantation. 2009;23 Suppl 21:2-9. Epub 2009/12/16.
- 46. Pennefather SH, Bullock RE, Dark JH. The effect of fluid therapy on alveolar arterial oxygen gradient in brain-dead organ donors. Transplantation. 1993;56(6):1418-22. Epub 1993/12/01.
- 47. Gattas DJ, Dan A, Myburgh J, Billot L, Lo S, Finfer S. Fluid resuscitation with 6% hydroxyethyl starch (130/0.4) in acutely ill patients: an updated systematic review and meta-analysis. Anesth Analg. 2012;114(1):159-69. Epub 2011/12/21.
- 48. De Backer D, Aldecoa C, Njimi H, Vincent JL. Dopamine versus norepinephrine in the treatment of septic shock: a meta-analysis*. Crit Care Med. 2012;40(3):725-30. Epub 2011/11/01.
- 49. Jassem W, Fuggle SV, Cerundolo L, Heaton ND, Rela M. Ischemic preconditioning of cadaver donor livers protects allografts following transplantation. Transplantation. 2006;81(2):169-74. Epub 2006/01/27.
- 50. Follette DM, Rudich SM, Babcock WD. Improved oxygenation and increased lung donor recovery with high-dose steroid administration after brain death. The Journal of heart and lung transplantation: the official publication of the International Society for Heart Transplantation. 1998;17(4):423-9. Epub 1998/05/20.
- 51. Dimopoulou I, Tsagarakis S, Anthi A, Milou E, Ilias I, Stavrakaki K, et al. High prevalence of decreased cortisol reserve in brain-dead potential organ donors. Crit Care Med. 2003;31(4):1113-7. Epub 2003/04/19.
- 52. Amato MB, Barbas CS, Medeiros DM, Magaldi RB, Schettino GP, Lorenzi-Filho G, et al. Effect of a protective-ventilation strategy on mortality in the acute

- respiratory distress syndrome. The New England journal of medicine. 1998;338(6):347-54. Epub 1998/02/05.
- 53. Brower RG, Lanken PN, MacIntyre N, Matthay MA, Morris A, Ancukiewicz M, et al. Higher versus lower positive end-expiratory pressures in patients with the acute respiratory distress syndrome. The New England journal of medicine. 2004;351(4):327-36. Epub 2004/07/23.
- 54. Powner DJ, Darby JM, Kellum JA. Proposed treatment guidelines for donor care.

 Prog Transplant. 2004;14(1):16-26; quiz 7-8. Epub 2004/04/14.

 Table 1. Summary of Randomized Controlled Trials of Interventions for Care of Brain-Dead Organ Donors

Intervention, source and	Treatment			Specific	Organ function	
number of patients	groups	Control group	Outcome	outcome	post TX	Survival
ADH						
Iwai et al, 1989 (28)	Group 1: epinephrine for	Epinephrine	Donor survival	Hours to death	No	No
- 25 patients	SBP > 100 mmHg					
	Group 2: ADH 0.1-0.4 U/hr					
	Group 3: ADH 1-2 U/hr +					
	epinephrine					
Pennefather et al,		Placebo	Hemodynamic	CI, MAP, pressor	Yes	Patient
1995 (31)*	ADH 300 μU.kg ⁻¹ min ⁻¹		parameters	dose		
- 24 patients						
Guesde et al,	ADH 1 μ g 2/2h when	No treatment	Renal function	Hemodialysis	Yes	Graft
1998 (32)*	diuresis > 300 mL/h			requirement,		
- 97 patients				creatinine D1 to D15		
Methylprednisolone						
Chatterjee et al,	Methylprednisolone 5 g	No treatment	Renal function	Graft failure at	Yes	Graft
1977 (24)	prior to harvesting			1 and 3 months		
- 50 patients						
Kotsch et al,	Methylprednisolone 250	No treatment	Liver function	ALT/AST/BB D1	Yes	Graft
2008 (25)	mg bolus + 100 mg/h until			and D10		
- 100 patients	recovery of organs					
Venkateswaran	Methylprednisolone 1 g	Placebo	Lung function	PaO ₂ /FiO ₂ ratio	No	No
et al, 2008 (9)			and suitability			
- 60 patients						

Triiodothyronine						
Mariot et al,	1/1h or 0.5/0.5h: T ₃ 2 or 4	Placebo	Hemodynamic	CI, MAP, pH pre and	No	No
1991 (14)*	μg and hydrocortisone 100		parameters;	post protocol		
- 40 patients	mg		organ retrieval			
Randell et al,	T_3 2 µg/h started	No treatment	Hemodynamic	Maximum	Yes	No
1992 (26)	immediately before surgery		parameters;	ALT/AST/BB to D7,		
- 25 patients			liver function	pressor dose		
Goarin et al,	T_3 0.2 µg/kg bolus	Placebo	Hemodynamic	CI, LVF, MAP pre	No	No
1996 (13)*			parameters;	and post protocol		
- 37 patients			heart function			
Jeevanandam	T_3 0.6 µg/kg bolus	Placebo	Hemodynamic	Creatinine, pressor	Yes	Graft
et al, 1997 (27)			parameters;	dose		
- 30 patients			kidney			
			function			
Perez-Blanco et al,	T_3 1 µg/kg bolus + 0.06	Placebo	Hemodynamic	CI, CO2 gap and	No	No
2005 (33)*	μg/kg/h		parameters	lactate pre and post		
- 52 patients				protocol		
Venkateswaran	$T_3 0.8 \mu\text{g/kg bolus} + 0.113$	Placebo	Hemodynamic	CI, LVSWI, SVR pre	No	No
et al, 2009 (34)*	μg/kg/h and/or		parameters	and post protocol		
- 80 patients	methylprednisolone 1000					
T1 '1	mg bolus					
Fluid management-						
hydroxyethyl starch	TT 1 41 1 4 1	C . 11 : 1	TT 1 '	IID MAD	3 7	NT
Randell et al,	Hydroxyethyl starch	Crystalloid	Hemodynamic	HR, MAP,	Yes	No
1990 (22)	according to hemodynamic		parameters;	electrolyte balance		
- 16 patients	data	Calada	liver function	Constinue of D10	NI.	NT.
Cittanova et al,	Hydroxyethyl starch up to	Gelatin	Kidney	Creatinine at D10,	No	No
1996 (23)	33 mL/kg		function	hemodialysis		
- 27 patients				requirement		

Vasopressor- dopamine						
Schnuelle et al,	Dopamine 4 μg/kg/m	No treatment	Renal function	Hemodialysis	Yes	Graft and
2009 (21)				requirement		Patient
- 264 patients						
Surgical technique-						
ischemic preconditioning						
Azoulay et al,	10 min of ischemic liver	Conventional	Liver function	ALT/AST D5 and	Yes	Patient
2005 (35)*	preconditioning before	technique		D10, BB D7 and		
- 91 patients	harvesting			D15, PT D3-D15		
Amador et al,	10 min of ischemic liver	Conventional	Liver function	ALT/AST D1-D10,	Yes	Graft and
2007 (36)*	preconditioning before	technique		apoptosis		patient
- 60 patients	harvesting					
Donor harvesting						
technique						
D'Amico et al,	Modified double perfusion	Conventional	Liver function	ALT/AST/BB/PT D2	Yes	Graft and
2007 (30)	technique	technique		and D7		patient
- 35 patients						
Preservation solution						
(Euro-Collins)						
Cofer et al,	Euro-Collins solution	University of	Liver function	ALT/AST/BB/PT	Yes	Patient
1992 (29)		Wisconsin				
- 56 patients		solution				
Mechanical ventilation						
Mascia et al,	Lung protective strategy	Conventional	Lung function	PaO ₂ /FiO ₂ ratio, pH,	No	Patient
2010 (20)		ventilator	and suitability	MAP		
- 118 patients		strategy				
ADU: dosmonrossin: AI	T. alanina aminatransfarasa.	ACT: asportate	minotronoforoso	DD. bilimbin, CI. cord	ing indox. UE	D. boomt roto, I VE

ADH: desmopressin; ALT: alanine aminotransferase; AST: aspartate aminotransferase; BB: bilirubin; CI: cardiac index; HR: heart rate; LVF: left ventricular function; LVSWI: left ventricular stroke work index; MAP: mean arterial pressure; PT: prothrombin time; SVR: systemic vascular resistance; TX: transplantation. * Trials included in meta-analysis.

Table 2. Risk of bias in studies

Source	Adequate sequence generation	Allocation concealment	Blinding	Complete organ function data addressed	Complete survival outcome data addressed	
ADH						
Iwai et al, 1989 (28)	Unclear	Unclear	No	NM	NM	
Pennefather et al, 1995 (31)*	Unclear	Unclear	No	No	No	
Guesde et al, 1998 (32)*	Yes	Yes	Yes	No	No	
Methylprednisolone						
Chatterjee et al, 1977 (24)	Yes	Yes	Yes	No	No	
Kotsch et al, 2008 (25)	Yes	Yes	Yes	No	Unclear	
Venkateswaran et al, 2008 (9)	Yes	Yes	Yes	NM	NM	
Triiodothyronine						
Mariot et al, 1991 (14)*	Yes	No	Yes	NM	NM	
Randell et al, 1992 (26)	Unclear	Unclear	No	No	NM	
Goarin et al, 1996 (13)*	Yes	Unclear	Yes	NM	NM	
Jeevanandam et al, 1997 (27)	Unclear	Unclear	Yes	No	No	
Perez-Blanco et al, 2005 (33)*	Unclear	Unclear	Yes	NM	NM	
Venkateswaran et al, 2009 (34)*	Yes	Yes	Yes	NM	NM	
Fluid management-						
hydroxyethyl starch						
Randell et al, 1990 (22)	Unclear	Unclear	No	No	NM	
Cittanova et al, 1996 (23)	Unclear	Unclear	Yes	NM	NM	
Vasopressor- dopamine						
Schnuelle et al, 2009 (21)	Yes	Yes	No	No	NM	

Surgical technique-ischemic					
preconditioning					
Azoulay et al, 2005 (35)*	No	No	No	No	No
Amador et al, 2007 (36)*	Yes	Yes	No	No	No
Donor harvesting technique					
D'Amico et al, 2007 (30)	Yes	Yes	No	No	No
Preservation solution (Euro-Collins)					
Cofer et al, 1992 (29)	Unclear	Unclear	No	No	No
Mechanical ventilation					
Mascia et al, 2010 (20)	Yes	Yes	No	NM	No

ADH: desmopressin; ALT: alanine aminotransferase; AST: aspartate aminotransferase; BB: bilirubin; CI: cardiac index; HR: heart rate; LVF: left ventricular function; LVSWI: left ventricular stroke work index; MAP: mean arterial pressure; NM: not measured; PT: prothrombin time; SVR: systemic vascular resistance; TX: transplantation.

^{*} Trials included in meta-analyses.

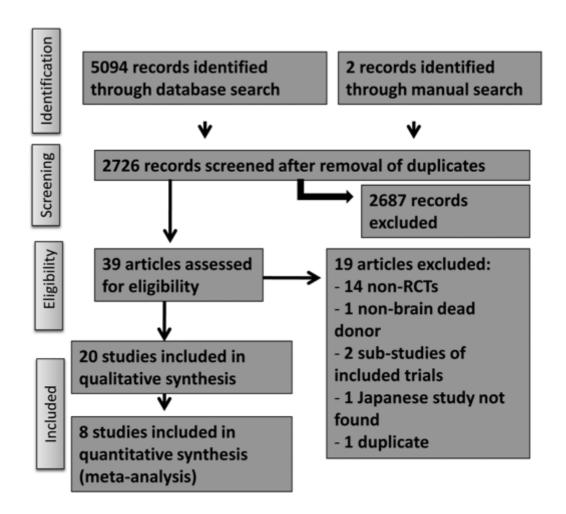


Figure 1. Flowchart summarizing search strategies and selection of trials.

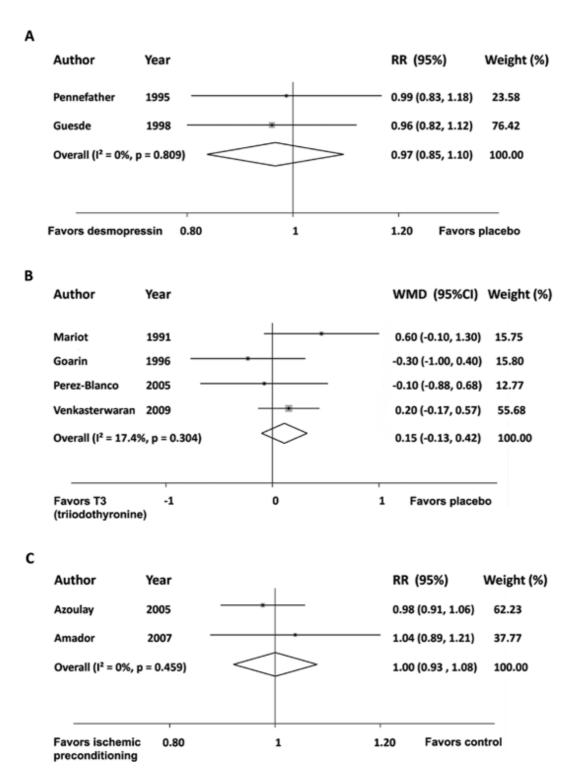


Figure 2. A. Forest plot comparing the effects of desmopressin versus placebo on early graft function in kidney recipients. B. Forest plot comparing the effects of triiodothyronine versus placebo on cardiac index in donor hearts. C. Forest plot comparing the effects of liver preconditioning during harvesting procedures with conventional technique on patient survival. WMD = weighted mean difference.

CAPÍTULO 3

Brain death-induced inflammatory and procoagulant activity in human pancreatic tissue: a case-control study

Short title: Brain death-induced human pancreatic tissue inflammation

Rech T.H., MsC^a; Crispim D., PhD^b; Rheinheimer J.^b; Sigal S.B.^b; Osvaldt A.B.^c, PhD; Grezzana Filho T. J. M.^c, PhD; Kruel C.R.P., PhD^c; Gross J.L, PhD^b; Leitão C.B., PhD^b.

^aDivision of Intensive Care Medicine, Hospital de Clínicas de Porto Alegre, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil.

^bHuman Islet Biology Laboratory, Division of Endocrinology, Hospital de Clínicas de Porto Alegre, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil.

^cDivision of Surgery, Hospital de Clínicas de Porto Alegre, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil.

Corresponding author and reprint requests:

Tatiana Helena Rech

Division of Intensive Care Medicine of Hospital de Clínicas de Porto Alegre Rua Ramiro Barcelos 2350, 13º andar, 90035-003, Porto Alegre-RS, Brazil.

E-mail: tatiana.rech@hotmail.com

Phone: + 55 51 3359 8127 Fax: + 55 51 3359 8777

Key words: brain death; cytokines; thromboplastin; islets of Langerhans

Abstract

Long-term insulin independence after islet transplantation depends on engraftment of a large number of islets. However, the yield of islet cells from brain-dead donors is negatively affected by the up-regulation of inflammatory mediators. Brain death (BD) is also believed to increase the expression of tissue factor (TF), which further contributes to a low rate of islet cell engraftment. We conducted a case-control study to assess BD-induced inflammatory and thrombotic effects in human pancreatic tissue.

Seventeen brain-dead patients and 20 control patients undergoing pancreatectomy were included in the study. Serum TNF- α , IL-6, IL-1 β , IFN- γ , and TF were measured using commercial ELISA kits. Gene expressions of these cytokines and *TF* were evaluated by RT-qPCR on pancreatic samples. Protein quantification was performed by immunohistochemistry in paraffin-embedded pancreas sections.

Brain-dead patients had higher serum concentrations of TNF- α and IL-6 in comparison to controls. The groups had similar TNF- α , IL-6, IL-1 β , and IFN- γ mRNA levels in pancreatic tissue. RT-qPCR revealed significant up-regulation of *TF* mRNA expression in control patients. Immunohistochemical analyses showed that brain-dead patients had increased TNF- α protein levels compared to controls.

BD induces profound inflammatory derangements that are evidenced by the upregulation of TNF- α in serum and pancreatic tissue. Blocking the expression of key inflammatory mediators in brain-dead donors should be evaluated as a new approach to improve the outcomes of islet transplantation.

Introduction

Type 1 diabetes mellitus is characterized by severe insulin deficiency resulting from progressive destruction of pancreatic beta-cells by the immune system (1). Islet transplantation is an effective therapy for unstable type 1 diabetes mellitus (2, 3). However, restoration of β -cell function after transplantation and the ensuing improvement in glycemic control (4) depend on a high rate of islet engraftment, which is hindered by the marked destruction of islets during the transplantation process (5) as a consequence of pancreas preservation, isolation procedures (6), and donor and recipient characteristics (7). Even if important benefits are conferred by partial function (8, 9), the ultimate goal of insulin independence after islet transplantation still requires infusion of a large number of islets, and therefore multiple donors (10).

Previous studies have reported that the source of transplants, from either brain dead or living donors, impacts graft survival rate (11, 12). Jung *et al* showed a better yield of islet cell mass from living pancreatic donors compared with brain-dead donors (13). Animal and human studies suggest that BD causes the release of potent proinflammatory cytokines (14-17) and procoagulant mediators (18, 19) into the circulation, with potential deleterious effects on transplanted organs (20). Contreras *et al* demonstrated that BD-associated up-regulation of tumor necrosis factor- α (TNF- α), interleukin-1 β (IL-1 β), and interleukin-6 (IL-6) significantly reduces isolated islet yield, viability, and function after transplantation in rats (21). Therefore, it has been suggested that cytokine blockade could enhance islet engraftment (3, 22).

Recently, expression of tissue factor (TF), a glycoprotein implicated in extrinsic and intrinsic coagulation pathways, has been reported in the islets of Langerhans (23). Infusion of TF-expressing islets into the portal vein triggered an instant blood-mediated inflammatory reaction (IBMIR) characterized by rapid binding of platelets to islet

surface as the islets come into direct contact with ABO-compatible blood (24, 25). TF up-regulation was demonstrated in a rat model of BD, suggesting that BD could stimulate TF expression (18).

To the best of our knowledge, there are no studies focusing on BD-induced inflammation in human pancreatic tissue. The aim of this case-control study was therefore to assess the inflammatory and procoagulant effects induced by BD in human pancreatic tissue.

Patients and Methods

Brain-dead patients and controls

The study protocol was approved by the research ethics committee at Hospital de Clínicas de Porto Alegre. Informed consent was obtained from control patients or from the next of kin in the case of brain-dead individuals. BD was assessed independently by two physicians, according to Brazilian law (26), and was based on the following criteria: coma with complete unresponsiveness, including absence of all brain stem reflexes, apnea test, and confirmation image with absence of cerebral blood flow. All brain-dead patients (n=17) older than 18 years and not accepted for pancreas donation were prospectively included in the study from November 2010 to December 2011. Pancreas was procured during multiorgan harvest for organ transplantation, stored in histidine-tryptophan-ketoglutarate (HTK, Custodiol®) preservation solution and immediately transferred to the islet isolation laboratory, where a 2cm² biopsy was taken. A 2cm² pancreas biopsy was also obtained from control patients (n=20) during partial or total pancreatectomy for the treatment of underlying lesions (mainly periampullary tumors). Control patients were prospectively enrolled during the same period as cases.

Clinical and biochemical data were recorded at the time of BD diagnosis or on the day before surgery for controls.

Serum TNF- α , IL-6, IL-1 β , IFN- γ and TF determinations

A 20-mL whole blood sample was collected in silicone-coated tubes (Vacutainer) at the time of surgical procedures for all patients, and was centrifuged at 2500g for 10 min at 4°C. Serum was separated and immediately stored at -20°C until analysis. Circulating levels of TNF- α , IL-6, IL-1 β , interferon- γ (IFN- γ), and TF were assessed by enzyme-linked immunosorbent assay (ELISA) using commercially available kits with primary polyclonal antibodies following the manufacturer's recommendations (Biosource Europe S.A., Nivelles, Belgium).

TNF-α, IL-6, IL-1β, IFN-γ, and TF mRNA isolation and quantification by RT-qPCR

Pancreatic tissue biopsies were excised, snap-frozen in liquid nitrogen and stored at -80°C until use. Pancreatic tissue (120mg) was homogenized in phenol-guanidine isothiocyanate (Invitrogen Life Technologies, Carlsbad, CA). RNA was extracted with chloroform and precipitated with isopropanol by centrifugation (12,000g) at 4°C. RNA pellet was washed twice with 75% ethanol and re-suspended in 10-50μl of water treated with diethylpyrocarbonate. The concentration and quality of total RNA samples were assessed using a NANODROP 2000 spectrophotometer (Thermo Scientific Inc., Newark, DE). Only RNA samples with adequate purity ratios (A260/A280=1.9-2.1) were used for subsequent analyses (27). In addition, RNA integrity and purity was also checked on agarose gel containing GelRedTM Nucleic Acid Gel Stain (Biotium Inc., Hayward, CA). The mean RNA concentration isolated was 800μg/120mg pancreatic tissue.

Real-time reverse transcription-PCR was performed in two separate reactions. First, RNA was reverse-transcribed into cDNA. cDNA was then amplified by quantitative real-time PCR (RT-qPCR). Reverse transcription of 1µg of RNA into cDNA was carried out using the SuperScriptTM III First-Strand Synthesis System for RT-PCR (Invitrogen Life Technologies) following the manufacturer's protocol for the oligo (dT) method (28-31).

RT-qPCR experiments were performed in a ViiTM 7 Fast Real-Time PCR System Thermal Cycler with ViiTM 7 Ruo Software (Life Technologies, Foster City, CA). Experiments were performed by real-time monitoring of the increase in fluorescence of SYBER® Green dye (Life Technologies) (32). Primer sequences for *TNF-a*, *IL-6*, *IL-1β*, *IFN-γ*, *TF*, and *β-actin* genes were designed using Primer Express 3.0 Software (Life Technologies) and are depicted in Table 1. PCR reactions were performed using 5µl of 2x Fast SYBER® Green Master Mix (Life Technologies), 0.5µl (1ng/µl) of forward and reverse primers for *TNF-a*, *IL-6*, *IL-1β*, *IFN-γ*, *TF*, or *β-actin* and 1µl of cDNA template (5µg/µl), in a total volume of 10µl. Each sample was assayed in triplicate and a negative control was included in each experiment. Thermocycling conditions for these genes were as follows: initial cycle of 95°C for 5s and 60°C for 1min10s. RT-qPCR specificity was determined using melting curve analyses. All primers generated amplicons that produced a single sharp peak during the analyses.

Quantification of TNF- α mRNA was performed using the relative standard curve method (Life Technologies) (27) and the β -actin gene as reference. Relative standard curves were generated for both target and reference genes by preparing serial dilutions of a pool of cDNA samples with a known relative quantity. Then, relative amounts of each TNF- α mRNA sample were obtained by normalizing their signal by those of β -actin. Results are represented as arbitrary units (AU). Relative quantification of IL- δ ,

IL-1β, IFN-γ and *TF* cDNA was performed using the comparative $\Delta\Delta$ Cq method (27, 33). Quantities were expressed relative to the reference gene (β -actin). Validation assays were done by separate amplification of the target (*IL-6, IL-1β, IFN-γ* and *TF*) and reference (β -actin) genes, using serial dilutions of a pool of cDNA samples. As a requirement of this method, both target and reference genes exhibited equal amplification efficiencies (E=95% to 105%) in all experiments. The $\Delta\Delta$ Cq method calculates changes in gene expression as relative fold difference (n-fold changes) between an experimental and external calibrator sample (27, 33).

Immunohistochemistry for TNF- α , IL-6, IL-1 β , IFN- γ , and TF proteins in human pancreatic tissue

TNF-α, IL-6, IL-1β, IFN-γ and TF protein distributions and intensities were determined by immunohistochemistry in formalin-fixed, paraffin-embedded pancreas sections. Anti-TNF- α , anti-IL-6, anti-IL-1 β , anti-IFN- γ , and anti-TF rabbit polyclonal antibodies (Santa Cruz Biotechnology, Inc.) were used to detect TNF-α, IL-6, IL-1β, IFN-γ, and TF protein expression in human pancreatic tissue, with intestine and placenta used as positive controls. Immunohistochemical analyses were performed on 4μm pancreas sections. Routine immunohistochemical techniques comprised deparaffination and rehydration, antigenic recovery, inactivation of endogenous peroxidase, and blocking of non-specific reactions. Slides were incubated with primary antibody and then with a biotinylated secondary antibody, streptavidin horseradish peroxidase conjugate (LSAB; Dako Cytomation Inc., Carpinteria, CA), and diaminobenzidine tetrahydrochloride (kit DAB; Dako Cytomation Inc., Carpinteria, CA). Quantifications of TNF-α, IL-6, IL-1β, IFN-γ, and TF proteins were performed by digital image analyses using Image Pro Plus software, version 4.5 (Media Cybernetics, Bethesda, MD). Images were visualized through a Zeiss microscope (model AXIOSKOP-40; Carl Zeiss, Oberkochen, Germany) and captured using the Cool Snap-Pro CS (Media Cybernetics, Bethesda, MD, USA) camera in a blinded fashion. Two independent blinded investigators (T.H.R and S.S.B) analyzed the intensity of brownish-colored immunostaining in pixels in 10 fields from each slide. A Pearson correlation of r^2 =0.902 (P< 0.001) was obtained for the two analyses. The mean number of pixels identified by the two investigators was used to quantify TNF- α , IL-6, IL-1 β , IFN- γ , and TF protein expression in each sample. Paraffin-embedded pancreas sections from 10 adult pancreatic necropsies performed during the study period were used as additional controls for TF protein expression, since pancreatic cancer may induce TF expression (34), which would make our controls unsuitable for this marker.

Statistical Analysis

Variables with normal distribution are presented as mean \pm SD. Variables with skewed distribution were log-transformed before analysis and are presented as median and interquartile intervals. Categorical variables are presented as percentages. Serum TNF- α , IL-6, IL-1 β , IFN- γ , and TF levels and mRNA and protein expressions were compared between the groups using Student's t test. Statistical significance of differences in TF protein expression in brain-dead individuals, controls, and necropsies was determined by one-way ANOVA with Bonferroni-Dunn post hoc test. Pearson's test was used to assess correlations between different quantitative variables. A multiple linear regression was performed in order to adjust TNF- α expression for possible confounding factors. Values were considered statistically significant if P<0.05. All statistical analyses were performed using SPSS 18.0 (Chicago, IL).

Results

The characteristics of 17 brain-dead patients and 20 control patients included in the study are summarized in Table 2. Periampullary tumors were the indication for pancreatectomy in the control group, except in one patient with a cystic tumor. Two control patients were excluded after their biopsies revealed severe chronic pancreatitis. Severe spontaneous intracranial hemorrhage was the cause of BD in 13 patients and cardiac arrest resulting in severe anoxic encephalopathy in 4 patients. Fifty-six organs were retrieved for transplantation (2 hearts, 4 lungs, 16 livers, and 34 kidneys). Age, body mass index (BMI), plasma glucose or glycated hemoglobin (HbA_{1C}) did not differ significantly between the groups. Brain-dead patients were mostly men, while controls were mainly women. As expected, brain-dead patients spent more hours on mechanical ventilation, had more episodes of cardiac arrest and persistent hypotension, and more frequently developed hypernatremia and low platelet levels than controls. Desmopressin and steroids were used as part of the care of potential organ donors in 10 patients.

Serum TNF-α, IL-6, IL-1β, IFN-γ, and TF quantifications by ELISA

Blood samples and pancreas biopsies were obtained at different time points in brain-dead patients and controls (median 12 hours [10-18] vs. 3.5 hours [2.6-4], respectively; P<0.001). As shown in Fig. 1, brain-dead patients had higher concentrations of TNF- α (12.03 pg/mL [6.2-23.6] vs. 3.8 pg/mL [3.4-6.7]; P=0.005) and IL-6 (1127.1 pg/mL [335.7-4571.6] vs. 77.4 pg/mL [48.1-186.6]; P<0.00001) in comparison to controls. Serum IL-1 β , IFN- γ and TF were similar in brain-dead patients and controls (IL-1 β 0.1 pg/mL [0.1-92.2] vs. 0.1 pg/mL [0.1-0.1], P=0.516; IFN- γ 0.03 pg/mL [0.02-0.05] vs. 0.03 pg/mL [0.02-0.03], P=0.128; TF 126.8 pg/mL [80.6-291.4] vs. 76.9 pg/mL [63.6-102.5], P=0.170).

TNF- α , IL-6, IL-1 β , IFN- γ , and TF mRNA quantification in human pancreatic tissue by RT-qPCR

Real-time qPCR analysis was performed in 35 pancreatic biopsies (17 brain-dead patients and 18 controls), as shown in Fig. 2. TNF- α , IL-6, IL- 1β and IFN- γ mRNA levels in pancreatic tissue were similar in the two groups (TNF- α brain-dead 4.65 A.U [1.4-8.7] vs. controls 2.53 A.U [2.0-9.1], P=0.875; IL-6 brain-dead 0.47-fold [0.06-1.2] vs. controls 0.58-fold [0.03-1.5], P=0.887; IL- 1β brain-dead 0.16-fold [0.03-1.2] vs. controls 0.06-fold [0.001-3.1], P=0.148 and IFN- γ brain-dead 0.54-fold [0.04-2.4] vs. controls 0.94-fold [0.3-1.6], P=0.330). For TF analysis, we excluded three control patients considered to have advanced malignant disease. Unexpectedly, RT-qPCR revealed significant up-regulation of TF mRNA expression in control patients (brain-dead 0.39-fold [0.1-1.2] vs. control 1.38-fold [0.7-2.0], P=0.049).

TNF- α , IL-6, IL-1 β , IFN- γ , and TF protein immunohistochemistry quantifications in human pancreatic tissue

Immunohistochemical studies were conducted to quantify protein expression of BD-induced inflammatory cytokines and procoagulant activity in human pancreatic tissue (Table 3).

Immunohistochemical analysis showed that brain-dead patients had increased TNF- α protein expression compared to controls (16.81 ± 5.2 pixels vs. 11.57 ± 4.93 pixels; P<0.005), in agreement with the results obtained for serum TNF- α levels (r=0.451; P=0.014). Moreover, TNF- α protein was widely distributed in all pancreatic tissue, including islets (Figure 3). After controlling for possible confounding factors identified in the univariate analysis (sex, ventilation support, and vasopressor support),

BD remained independently associated with TNF- α protein up-regulation (beta= 7.64 [CI 0.81-14.48]; P=0.030).

IL-6 and IL-1 β proteins were identified in both ductal cells and islet cells, but no significant differences were observed between cases and controls (IL-6 15.65 \pm 6.3 pixels vs. 19.89 \pm 9.25 pixels, P=0.132; IL-1 β 12.89 \pm 6.2 pixels vs. 11.87 \pm 8.02 pixels, P=0.683). This indicates that IL-6 and IL-1 β expression in human pancreatic tissue was not affected by BD, as also demonstrated by mRNA quantification.

Staining of human pancreas sections with polyclonal antibodies against IFN- γ and TF showed that IFN- γ and TF were minimally present in pancreatic tissue. The mean \pm SD TF protein concentration for the entire pancreatic tissue was 15.71 ± 9.53 pixels. Moreover, islets of Langerhans were not stained, indicating no expression of TF in the endocrine pancreas. Because an increase in TF mRNA expression by RT-qPCR was observed in control patients compared to brain-dead patients, we also used 10 pancreas sections from necropsies as a second set of controls, and no significant differences were found among groups (brain-dead 15.63 ± 9.6 vs. control 16.39 ± 8.32 vs. necropsies 14.7 ± 12.01 , P=0.909).

Discussion

In this study, BD was associated with systemic inflammation, as demonstrated by an increase in TNF- α and IL-6 serum levels, as well as pancreatic inflammation, reflected by the up-regulation of TNF- α protein expression in pancreatic tissue of brain-dead subjects as compared to controls. BD-associated increases in IL-1 β , IFN- γ or TF were not detected.

It is well known that BD has a negative non-immunological effect on organ function (11, 35). Understanding the mechanisms by which BD affects pancreatic

function is of great interest to islet transplantation. To the best of our knowledge, the present case-control study was the first to show that damage to human pancreatic tissue is caused by BD before other stress factors related to isolation and transplantation procedures are triggered. The increase in TNF-α in the pancreatic tissue of brain-dead patients prior to islet isolation and transplantation suggests that BD itself determines the onset of the inflammatory response detected in human pancreas. These results are in accordance with the results of Birks et al, who compared the expression of TNF-α in myocardial biopsies of brain-dead and living donors immediately before transplantation, showing that the increase in TNF- α in cadaveric donors was a consequence of BD (36). Moreover, the outcomes of islet transplantation seem to have improved since the soluble TNF receptor antagonist etanercept was introduced as part of immunosuppression protocols (3, 22), suggesting that TNF- α may be a relevant factor damaging the islets. In fact, TNF-α induces nuclear factor-κB (NF-κB) activation, a pro-apoptotic mechanism contributing to pancreatic beta cell death in type 1 diabetes (37, 38), while blockage of NF-κB activation protects beta cells against TNF-α-induced apoptosis (39). In islet transplantation, TNF-α may act at different points in the process, leading to pathological signs of islet cell death: at the tissue level, before pancreas recovery, as demonstrated by us, during isolation procedures, as a consequence of cytokine production by islets (40), and at the graft site, as part of recipient inflammatory response (41).

IL-1 β induces endothelial activation, promoting leukocyte interactions with adhesion molecules expressed on the cell surface (42). Moreover, IL-1 β and IFN- γ induce beta cell apoptosis via activation of beta cell gene networks under the control of NF- κ B and STAT-1 (43). In our study, IL-1 β and IFN- γ were equally distributed in blood and pancreas tissue from brain-dead and control patients, suggesting that BD was not linked to the release of these cytokines. These results differ from those obtained

with an animal model of BD, which demonstrated an intense IL-1 β response in serum and myocardium of rats (42). Because our study was the first study to evaluate the behavior of IL-1 β after BD in humans, we believe this might be reflecting a difference between species. Weiss *et al* showed that IFN- γ mRNA expression was consistently higher in liver biopsies from brain-dead donors compared to living donors at the time of donor laparotomy (44), a finding not observed by us.

Contreras *et al* also reported the presence of a pro-inflammatory state shortly after the induction of BD in rats, as demonstrated by the up-regulation of cytokines, including TNF- α , IL-1 β , and IL-6, in serum and pancreatic tissue (21). Our study corroborates the up-regulation of TNF- α in human serum and pancreatic tissue. However, although IL-6 was increased in serum, IL-6, IL-1 β , and IFN- γ were not expressed in pancreatic tissue. A possible explanation for these differences lies on the fact that, in comparison to that previous study, our samples were collected later after the diagnosis of BD. However, it could also be that the difference is merely reflecting the unique characteristics of different species.

Also, differently from experimental models (18, 19), we did not find an increase in TF in human pancreas from brain-dead patients. Furthermore, our results demonstrated an increase in TF mRNA in controls, but this increase was not confirmed at the protein level, suggesting that TF gene expression undergoes an important post-transcriptional regulation. To investigate these unexpected findings, pancreas sections from necropsies were also studied, and no significant differences in TF expression related to BD were found. We believe that this could be explained by the specific conditions of our controls, since most had malignant tumors, and TF is suspected of being implicated with thromboembolic events in pancreatic cancer (34).

TF expression has been reported in islets in different experimental models (24, 25, 45) as well as in human islet isolates (46). It is also identified as the main trigger of IBMIR, which is one of the possible explanations for the poor engraftment following islet transplantation. So, we focused on the influence of BD on TF expression in human pancreas. However, the present results do not confirm this hypothesis, as BD was not associated with TF increase in pancreatic tissue. Saito *et al* reported similar findings in a rat model of BD in which TF was observed to increase in isolated islets, but not in pancreatic tissue before digestion procedures (18). That suggests that TF expression is triggered by cold ischemia and isolation procedures, rather than BD. We therefore believe that attempts to block TF expression should target the isolated islets and not the brain-dead donor.

This study has some limitations. First, the case-control design prevented us from establishing an ideal control group. Patients diagnosed with severe pancreatitis and advanced malignant tumors were excluded, because they have the potential to upregulate pro-inflammatory and procoagulant mediators (34, 47). Additionally, a second control group was used in TF analyses. Second, the median time to collection of blood and pancreas samples was different between groups, since the duration of a pancreatectomy is shorter than that needed to complete a multiorgan donation protocol. Nevertheless, this difference might weaken and not strengthen our findings, because Third, protein cytokines peak early after BD (21).quantification immunohistochemistry was not specifically determined in the islets, but rather in the total slide. However, knowing the exocrine cell-staining pattern is important because islet suspensions prepared for clinical transplantation contain up to 40% of nonendocrine duct cells (48).

In conclusion, our data suggest that brain death itself impacts IL-6 and TNF- α cytokine content before any transplantation procedures. This association of cytokine content and BD can explain, at least in part, the more favorable outcomes of living donation. Randomized controlled trials to test the performance of TNF- α blockers administered to brain-dead donors prior to organ harvesting procedures are therefore warranted.

Contributors

T.H.R participated in the study conception and design, data acquisition, analysis, and interpretation of data, drafting of the manuscript and revision of the manuscript. D.C and C.B.L participated in the study conception and design, analysis and interpretation of data, revision of the manuscript and statistical analysis. J.R, S.S.B, A.B.O, T.J.M.G.F and C.R.P.K participated in data acquisition and revision of the manuscript. J.L.G critically reviewed the manuscript for intellectual content.

Acknowledgements

This study was supported by Fundo de Incentivo à Pesquisa e Eventos (FIPE) from Hospital de Clínicas de Porto Alegre (Porto Alegre, RS, Brazil).

References

- Pihoker C, Gilliam LK, Hampe CS, Lernmark A. Autoantibodies in diabetes.
 Diabetes. 2005 Dec;54 Suppl 2:S52-61. PubMed PMID: 16306341.
- Shapiro AM, Lakey JR, Ryan EA, Korbutt GS, Toth E, Warnock GL, et al. Islet transplantation in seven patients with type 1 diabetes mellitus using a glucocorticoid-free immunosuppressive regimen. The New England journal of medicine. 2000 Jul 27;343(4):230-8. PubMed PMID: 10911004. Epub 2000/07/27. eng.
- 3. Hering BJ, Kandaswamy R, Ansite JD, Eckman PM, Nakano M, Sawada T, et al. Single-donor, marginal-dose islet transplantation in patients with type 1 diabetes. JAMA: the journal of the American Medical Association. 2005 Feb 16;293(7):830-5. PubMed PMID: 15713772. Epub 2005/02/17. eng.
- Mineo D, Pileggi A, Alejandro R, Ricordi C. Point: steady progress and current challenges in clinical islet transplantation. Diabetes care. 2009 Aug;32(8):1563-9.
 PubMed PMID: 19638527. Pubmed Central PMCID: 2713646. Epub 2009/07/30. eng.
- 5. Johansson H, Goto M, Dufrane D, Siegbahn A, Elgue G, Gianello P, et al. Low molecular weight dextran sulfate: a strong candidate drug to block IBMIR in clinical islet transplantation. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2006 Feb;6(2):305-12. PubMed PMID: 16426314. Epub 2006/01/24. eng.
- Ricordi C, Inverardi L, Kenyon NS, Goss J, Bertuzzi F, Alejandro R.
 Requirements for success in clinical islet transplantation. Transplantation. 2005
 May 27;79(10):1298-300. PubMed PMID: 15912093. Epub 2005/05/25. eng.

- 7. Kaddis JS, Danobeitia JS, Niland JC, Stiller T, Fernandez LA. Multicenter analysis of novel and established variables associated with successful human islet isolation outcomes. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2010 Mar;10(3):646-56. PubMed PMID: 20055802. Pubmed Central PMCID: 2860018. Epub 2010/01/09. eng.
- Leitao CB, Tharavanij T, Cure P, Pileggi A, Baidal DA, Ricordi C, et al.
 Restoration of hypoglycemia awareness after islet transplantation. Diabetes care.
 2008 Nov;31(11):2113-5. PubMed PMID: 18697903. Pubmed Central PMCID:
 2571057. Epub 2008/08/14. eng.
- Tharavanij T, Betancourt A, Messinger S, Cure P, Leitao CB, Baidal DA, et al.
 Improved long-term health-related quality of life after islet transplantation.

 Transplantation. 2008 Nov 15;86(9):1161-7. PubMed PMID: 19005394. Pubmed
 Central PMCID: 2741424. Epub 2008/11/14. eng.
- 10. Fiorina P, Shapiro AM, Ricordi C, Secchi A. The clinical impact of islet transplantation. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2008 Oct;8(10):1990-7. PubMed PMID: 18828765. Epub 2008/10/03. eng.
- 11. Terasaki PI, Cecka JM, Gjertson DW, Takemoto S. High survival rates of kidney transplants from spousal and living unrelated donors. The New England journal of medicine. 1995 Aug 10;333(6):333-6. PubMed PMID: 7609748. Epub 1995/08/10. eng.

- 12. Jassem W, Koo DD, Cerundolo L, Rela M, Heaton ND, Fuggle SV. Leukocyte infiltration and inflammatory antigen expression in cadaveric and living-donor livers before transplant. Transplantation. 2003;75(12):2001-7. Epub 2003/06/28.
- 13. Jung HS, Choi SH, Kim SJ, Lee KT, Lee JK, Jang KT, et al. A better yield of islet cell mass from living pancreatic donors compared with cadaveric donors. Clinical transplantation. 2007 Nov-Dec;21(6):738-43. PubMed PMID: 17988267. Epub 2007/11/09. eng.
- 14. Fisher AJ, Donnelly SC, Hirani N, Burdick MD, Strieter RM, Dark JH, et al. Enhanced pulmonary inflammation in organ donors following fatal non-traumatic brain injury. Lancet. 1999 Apr 24;353(9162):1412-3. PubMed PMID: 10227229. Epub 1999/05/05. eng.
- Eckhoff DE, Eckstein C, Smyth CA, Vilatoba M, Bilbao G, Rahemtulla FG, et al. Enhanced isolated pancreatic islet recovery and functionality in rats by 17beta-estradiol treatment of brain death donors. Surgery. 2004 Aug;136(2):336-45.
 PubMed PMID: 15300200. Epub 2004/08/10. eng.
- 16. Murugan R, Venkataraman R, Wahed AS, Elder M, Hergenroeder G, Carter M, et al. Increased plasma interleukin-6 in donors is associated with lower recipient hospital-free survival after cadaveric organ transplantation. Critical care medicine. 2008 Jun;36(6):1810-6. PubMed PMID: 18496370. Epub 2008/05/23. eng.
- 17. Barklin A. Systemic inflammation in the brain-dead organ donor. Acta anaesthesiologica Scandinavica. 2009 Apr;53(4):425-35. PubMed PMID: 19226294. Epub 2009/02/20. eng.
- 18. Saito Y, Goto M, Maya K, Ogawa N, Fujimori K, Kurokawa Y, et al. Brain death in combination with warm ischemic stress during isolation procedures induces the expression of crucial inflammatory mediators in the isolated islets. Cell

- transplantation. 2010;19(6):775-82. PubMed PMID: 20573302. Epub 2010/06/25. eng.
- Saito Y, Goto M, Maya K, Ogawa N, Fujimori K, Kurokawa Y, et al. The influence of brain death on tissue factor expression in the pancreatic tissues and isolated islets in rats. Transplantation proceedings. 2009 Jan-Feb;41(1):307-10.
 PubMed PMID: 19249541. Epub 2009/03/03. eng.
- Adrie C, Monchi M, Fulgencio JP, Cottias P, Haouache H, Alvarez-Gonzalvez A, et al. Immune status and apoptosis activation during brain death. Shock. 2010
 Apr;33(4):353-62. PubMed PMID: 20407403. Epub 2010/04/22. eng.
- 21. Contreras JL, Eckstein C, Smyth CA, Sellers MT, Vilatoba M, Bilbao G, et al. Brain death significantly reduces isolated pancreatic islet yields and functionality in vitro and in vivo after transplantation in rats. Diabetes. 2003 Dec;52(12):2935-42. PubMed PMID: 14633854. Epub 2003/11/25. eng.
- 22. Froud T, Ricordi C, Baidal DA, Hafiz MM, Ponte G, Cure P, et al. Islet transplantation in type 1 diabetes mellitus using cultured islets and steroid-free immunosuppression: Miami experience. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2005 Aug;5(8):2037-46. PubMed PMID: 15996257. Epub 2005/07/06. eng.
- 23. Johansson H, Lukinius A, Moberg L, Lundgren T, Berne C, Foss A, et al. Tissue factor produced by the endocrine cells of the islets of Langerhans is associated with a negative outcome of clinical islet transplantation. Diabetes. 2005

 Jun;54(6):1755-62. PubMed PMID: 15919797. Epub 2005/05/28. eng.
- 24. Moberg L, Johansson H, Lukinius A, Berne C, Foss A, Kallen R, et al. Production of tissue factor by pancreatic islet cells as a trigger of detrimental thrombotic

- reactions in clinical islet transplantation. Lancet. 2002 Dec 21-28;360(9350):2039-45. PubMed PMID: 12504401. Epub 2002/12/31. eng.
- 25. Berman DM, Cabrera O, Kenyon NM, Miller J, Tam SH, Khandekar VS, et al. Interference with tissue factor prolongs intrahepatic islet allograft survival in a nonhuman primate marginal mass model. Transplantation. 2007 Aug 15;84(3):308-15. PubMed PMID: 17700154. Epub 2007/08/19. eng.
- 26. 9434 Ln. 1997.
- 27. Bustin SA, Benes V, Garson JA, Hellemans J, Huggett J, Kubista M, et al. The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. Clinical chemistry. 2009 Apr;55(4):611-22. PubMed PMID: 19246619. Epub 2009/02/28. eng.
- 28. Papa S, Skulachev VP. Reactive oxygen species, mitochondria, apoptosis and aging. Molecular and cellular biochemistry. 1997 Sep;174(1-2):305-19. PubMed PMID: 9309704. Epub 1997/10/06. eng.
- 29. Li B, Nolte LA, Ju JS, Han DH, Coleman T, Holloszy JO, et al. Skeletal muscle respiratory uncoupling prevents diet-induced obesity and insulin resistance in mice. Nature medicine. 2000 Oct;6(10):1115-20. PubMed PMID: 11017142. Epub 2000/10/04. eng.
- 30. Nagase I, Yoshida T, Kumamoto K, Umekawa T, Sakane N, Nikami H, et al. Expression of uncoupling protein in skeletal muscle and white fat of obese mice treated with thermogenic beta 3-adrenergic agonist. The Journal of clinical investigation. 1996 Jun 15;97(12):2898-904. PubMed PMID: 8675704. Pubmed Central PMCID: 507386. Epub 1996/06/15. eng.
- 31. Carroll AM, Haines LR, Pearson TW, Fallon PG, Walsh CM, Brennan CM, et al. Identification of a functioning mitochondrial uncoupling protein 1 in thymus. The

- Journal of biological chemistry. 2005 Apr 22;280(16):15534-43. PubMed PMID: 15695816. Epub 2005/02/08. eng.
- 32. Higuchi R, Fockler C, Dollinger G, Watson R. Kinetic PCR analysis: real-time monitoring of DNA amplification reactions. Biotechnology (N Y). 1993

 Sep;11(9):1026-30. PubMed PMID: 7764001. Epub 1993/09/01. eng.
- 33. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. Methods. 2001

 Dec;25(4):402-8. PubMed PMID: 11846609. Epub 2002/02/16. eng.
- 34. Kasthuri RS, Taubman MB, Mackman N. Role of tissue factor in cancer. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2009 Oct 10;27(29):4834-8. PubMed PMID: 19738116. Pubmed Central PMCID: 2764391. Epub 2009/09/10. eng.
- 35. Stangl M, Zerkaulen T, Theodorakis J, Illner W, Schneeberger H, Land W, et al. Influence of brain death on cytokine release in organ donors and renal transplants. Transplantation proceedings. 2001 Feb-Mar;33(1-2):1284-5. PubMed PMID: 11267293. Epub 2001/03/27. eng.
- 36. Birks EJ, Owen VJ, Burton PB, Bishop AE, Banner NR, Khaghani A, et al. Tumor necrosis factor-alpha is expressed in donor heart and predicts right ventricular failure after human heart transplantation. Circulation. 2000 Jul 18;102(3):326-31. PubMed PMID: 10899097. Epub 2000/07/19. eng.
- 37. Barthson J, Germano CM, Moore F, Maida A, Drucker DJ, Marchetti P, et al. Cytokines tumor necrosis factor-alpha and interferon-gamma induce pancreatic beta-cell apoptosis through STAT1-mediated Bim protein activation. The Journal of biological chemistry. 2011 Nov 11;286(45):39632-43. PubMed PMID: 21937453. Pubmed Central PMCID: 3234786. Epub 2011/09/23. eng.

- 38. Eizirik DL, Colli ML, Ortis F. The role of inflammation in insulitis and beta-cell loss in type 1 diabetes. Nature reviews Endocrinology. 2009 Apr;5(4):219-26. PubMed PMID: 19352320. Epub 2009/04/09. eng.
- 39. Ortis F, Pirot P, Naamane N, Kreins AY, Rasschaert J, Moore F, et al. Induction of nuclear factor-kappaB and its downstream genes by TNF-alpha and IL-1beta has a pro-apoptotic role in pancreatic beta cells. Diabetologia. 2008

 Jul;51(7):1213-25. PubMed PMID: 18463842. Epub 2008/05/09. eng.
- 40. Lund T, Fosby B, Korsgren O, Scholz H, Foss A. Glucocorticoids reduce proinflammatory cytokines and tissue factor in vitro and improve function of transplanted human islets in vivo. Transplant international: official journal of the European Society for Organ Transplantation. 2008 Jul;21(7):669-78. PubMed PMID: 18346012. Epub 2008/03/19. eng.
- 41. Hanley S, Liu S, Lipsett M, Castellarin M, Rosenberg L, Tchervenkov J, et al. Tumor necrosis factor-alpha production by human islets leads to postisolation cell death. Transplantation. 2006 Sep 27;82(6):813-8. PubMed PMID: 17006329. Epub 2006/09/29. eng.
- 42. Silva IA, Correia CJ, Simas R, Correia CD, Cruz JW, Ferreira SG, et al. Inhibition of autonomic storm by epidural anesthesia does not influence cardiac inflammatory response after brain death in rats. Transplantation proceedings.
 2012 Sep;44(7):2213-8. PubMed PMID: 22974957. Epub 2012/09/15. eng.
- 43. Cnop M, Welsh N, Jonas JC, Jorns A, Lenzen S, Eizirik DL. Mechanisms of pancreatic beta-cell death in type 1 and type 2 diabetes: many differences, few similarities. Diabetes. 2005 Dec;54 Suppl 2:S97-107. PubMed PMID: 16306347. Epub 2005/11/25. eng.

- 44. Weiss S, Kotsch K, Francuski M, Reutzel-Selke A, Mantouvalou L, Klemz R, et al. Brain death activates donor organs and is associated with a worse I/R injury after liver transplantation. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2007 Jun;7(6):1584-93. PubMed PMID: 17430397.
- 45. Ji M, Yi S, Smith-Hurst H, Phillips P, Wu J, Hawthorne W, et al. The importance of tissue factor expression by porcine NICC in triggering IBMIR in the xenograft setting. Transplantation. 2011 Apr 27;91(8):841-6. PubMed PMID: 21325994. Epub 2011/02/18. eng.
- 46. Moberg L, Olsson A, Berne C, Felldin M, Foss A, Kallen R, et al. Nicotinamide inhibits tissue factor expression in isolated human pancreatic islets: implications for clinical islet transplantation. Transplantation. 2003 Nov 15;76(9):1285-8.
 PubMed PMID: 14627904. Epub 2003/11/25. eng.
- 47. Norman JG, Fink GW, Franz MG. Acute pancreatitis induces intrapancreatic tumor necrosis factor gene expression. Arch Surg. 1995 Sep;130(9):966-70. PubMed PMID: 7661681. Epub 1995/09/01. eng.
- 48. Beuneu C, Vosters O, Movahedi B, Remmelink M, Salmon I, Pipeleers D, et al. Human pancreatic duct cells exert tissue factor-dependent procoagulant activity: relevance to islet transplantation. Diabetes. 2004 Jun;53(6):1407-11. PubMed PMID: 15161741. Epub 2004/05/27. eng.

Table 1. Primer sequences used for *TNF-\alpha*, *IL-6*, *IL-1\beta*, *IFN-\gamma* and *TF* gene expression analyses.

Sequences

TNF - α gene ^a	F 5'- CCCAGGGACCTCTCTCAATCA -3'		
	R 5'- GGTTTGCTACAACATGGGCTACA -3'		
IL-6 gene ^a	F 5'- AGCCCTGAGAAAGGAGACATGTA -3'		
	R 5'- TCTGCCAGTGCCTCTTTGCT -3'		
IL - 1β gene ^a	F 5'- TGATGTCTGGTCCATATGAACTGAA -3'		
	R 5'- GGACATGGAGAACACCACTTGTT -3'		
IFN-γ gene ^a	F 5'- CCAACGCAAAGCAATACATGA -3'		
	R 5'- TCCTTTTTCGCTTCCCTGTTT -3'		
TF gene ^a	F 5'- TGTTCAAATAAGCACTAAGTCAGGAGAT -3'		
	R 5'- TCGTCGGTGAGGTCACACTCT -3'		
β-actin gene ^a	F 5'- GCGCGGCTACAGCTTCA -3'		
	R 5'- CTTAATGTCACGCACGATTTCC -3'		

F= forward primer; R= reverse primer.

^a= Primers were designed using published human gene sequences and the Primers Express 3.0 Software (Life Technologies). They were projected to target two consecutive exons of a gene in order to prevent the amplification of any contaminating genomic DNA.

Table 2. Baseline characteristics of brain-dead patients and controls.

	Brain-dead (n=17)	Controls (n=20)	p value
Cause of brain death or pancreatectomy	13 strokes and 4 anoxic injuries	6 benign tumors and 14 malignant tumors	-
Age (years)	54 ± 11	58 ± 13	0.34
Men (n, %)	12 (70.6)	7 (35)	0.03
BMI ⁺	25.9 ± 3.6	24.1 ± 3.6	0.13
Hypothermia (n, %)	2 (11.8)	0	0.115
Ventilation support (hours)	72 (48-114)	6 (5-6.7)	< 0.001
Time from BD to pancreas harvest or time to biopsy (hours)	12 (10-18)	3.5 (2.6-4)	< 0.001
Episode of cardiac arrest (n, %)	5 (29.4)	0	0.009
Vasopressor support (n, %)	17 (100)	6 (30)	< 0.001
Persistent hypotension (n, %)	4 (23.5)	0	0.022
Use of desmopressin (n, %)	10 (58.8)	0	< 0.001
Use of steroids (n, %)	10 (58.8)	0	< 0.001
Plasma sodium (mEq/L)	158 ± 10	141 ± 3	< 0.001
Plasma glucose (mg/dL)	174 ± 51	156 ± 67	0.40
HbA_{1C} (%)	6.2 ± 2.03	5.5 ± 1.42	0,2
Platelet count (x10 ⁹ /L)	185 ± 66	250 ± 56	0.002
Serum amylase (UI/L)	42 (30-199)	90 (49-114)	0.42
Serum lipase (UI/L)	22 (17-26)	73 (32-171)	0.01

All values were recorded at the time of brain death diagnosis or on the day before surgery for controls.

BMI: body mass index; HbA_{1C} : glycosylated hemoglobin.

^{+:} calculated as weight in kilograms divided by the square of height in meters.

Table 3. TNF- α , IL-6, IL-1 β , IFN- γ and TF protein immunohistochemistry quantifications* in human pancreatic tissue.

	Brain-dead (n=17)	Controls (n=18)	Necropsies (n=10)	p value
TNF-α	16.81 ± 5.2	11.57 ± 4.93	-	p<0.005
IL-6	15.65 ± 6.3	19.89 ± 9.25	-	p=0.132
IL-1β	12.89 ± 6.2	11.87 ± 8.02	-	p=0.683
IFN-γ	5.72 ± 3.74	3.99 ± 2.11	-	p=0.107
TF	15.63 ± 9.6	16.39 ± 8.32	14.7 ± 12.01	p=0.909

^{*} Values presented in pixels.

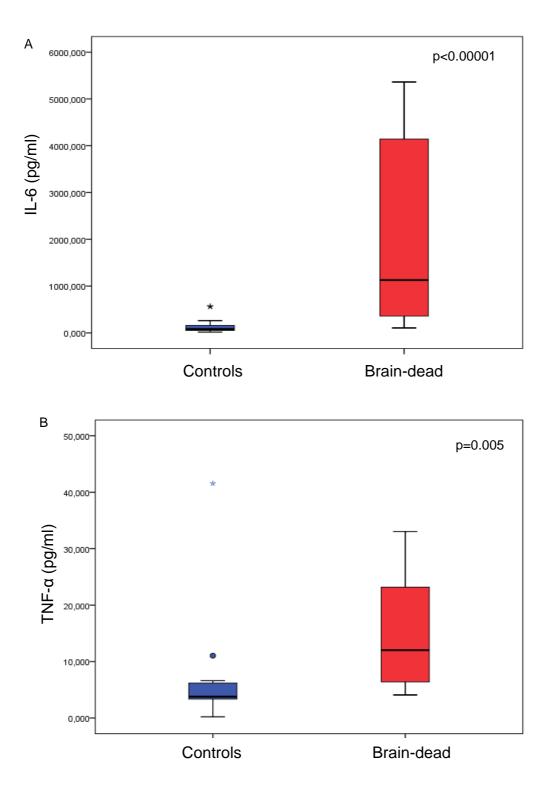


Figure 1. A. Serum IL-6 levels in brain-dead and controls. B. Serum TNF- α in brain-dead and controls.

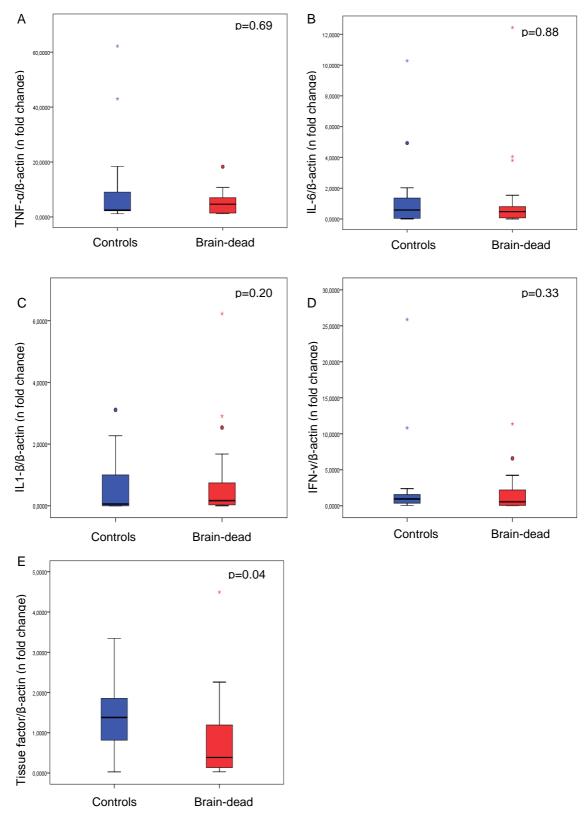


Figure 2. Cytokines quantifications by RT-qPCR in human pancreatic tissue, expressed in A.U (A. TNF- α) and in n-fold change related to calibrator sample (B. IL-6. C. IL-1 β . D. INF- γ . E. Tissue factor).

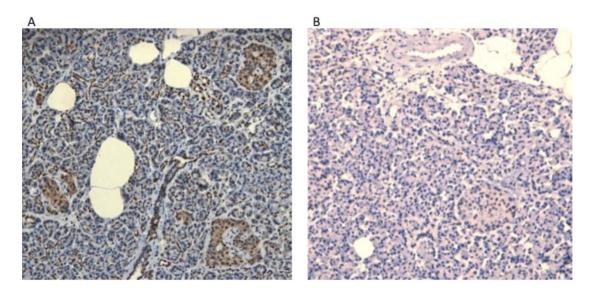


Figure 3. Staining of human pancreas with anti-TNF- α polyclonal antibody showing islets of Langerhans in brain-dead patients (A) and controls (B).

CAPÍTULO 4

Perspectivas futuras

Desde a década de 1990, quando o primeiro transplante de ilhotas em humanos foi realizado com sucesso (1), o transplante de ilhotas pancreáticas tornou-se uma terapia promissora para pacientes com DM tipo 1 com controle metabólico instável. Contudo, nos anos subsequentes, quando esse tipo de transplante passou a ser realizado em diversos centros ao redor do mundo, as taxas de sucesso relatadas não atingiram níveis satisfatórios (2, 3).

O desfecho de um transplante é resultado de uma complexa interação entre todos os eventos que acontecem desde o insulto que culmina com a ME até o período após o implante de órgão no receptor. O doador de múltiplos órgãos é uma peça fundamental dessa intrincada relação.

Há muito o que estudar no que concerne aos cuidados com o doador de órgãos. Este grupo de pesquisa tem focado seus esforços no estudo da ME e de suas consequências inflamatórias no pâncreas, como forma de aperfeiçoar o transplante de células β pancreáticas.

Recentemente, um alvo de pesquisas em DM tem sido o *glucagon-like peptide-1* (GLP-1). O GLP-1 é um hormônio peptídico liberado pelas células L do intestino delgado, cuja secreção é dependente da presença de nutrientes no lúmen do intestino. Sua liberação na corrente sanguínea estimula a secreção de insulina dependente de glicose e inibe a secreção de glucagon (4, 5). As propriedades antidiabetogênicas desse

hormônio levaram à pesquisa e ao desenvolvimento de um análogo do GLP-1, chamado exenatida. A exenatida, aprovada para uso clínico no tratamento do DM tipo 2 no Brasil, funciona como um forte agonista do receptor de GLP-1 (6-8).

Além do seu uso terapêutico no tratamento de pacientes com DM, a exenatida demonstra propriedades citoprotetoras para ilhotas pancreáticas (9, 10). A estimulação do receptor de GLP-1 produz efeitos diretos em células β, estimulando a regeneração e proliferação celular e reduzindo a apoptose, tanto em modelos animais como em ilhotas humanas isoladas (11, 12). Assim, seu potencial como agente citoprotetor no transplante de ilhotas foi testado clinicamente. Os resultados positivos desses estudos clínicos sugerem que, em breve, a exenatida poderá ser incluída como parte do tratamento dado a pacientes transplantados de ilhotas (13, 14).

A exenatida também possui atividade anti-inflamatória em linhagens de células β e ilhotas pancreáticas isoladas. Alguns estudos mostram a ação protetora da exenatida contra a atividade pró-inflamatória de citocinas como a IL-1β (15, 16). Por fim, um estudo publicado recentemente por Cechin *et al.* demonstrou que o tratamento com exenatida *in vitro* reduz a produção de citocinas e FT, além de ativar cinases, tais como Akt e ERK 1/2, em ilhotas humanas isoladas (17). Tais cinases fazem parte de diferentes vias de sinalização intracelular com funções pró-proliferativas e antiapoptóticas.

Desta forma, acreditamos que as propriedades anti-inflamatórias e próproliferativas da exenatida possam ser utilizadas para amenizar os danos causados pela ME nas ilhotas pancreáticas, melhorando a qualidade das células a serem transplantadas e reduzindo a perda da massa de ilhotas. Estamos testando essa hipótese através de um modelo experimental de ME em ratos Wistar, projeto atualmente em desenvolvimento no Centro de Pesquisa Experimental do HCPA. Em um futuro próximo, planejamos reproduzir esse mesmo projeto com o uso de bloqueadores de TNF-α.

Além disso, consideramos que a ME possa estar envolvida na perda precoce de ilhotas causada por apoptose das células β , através da ativação da via do fator de transcrição NF- κ B.

O fator de transcrição NF-κB é um heterodímero composto de duas subunidades, uma de 50 kDa e outra de 65 kDa. É encontrado no citoplasma da célula, associado a uma proteína inibitória, a IκB. O estímulo de citocinas inflamatórias resulta na fosforilação da IκB mediada pelo complexo da cinase da IκB, levando à degradação da IκB e consequente translocação do NF-κB para o núcleo (18). O NF-κB tem sido implicado como uma molécula sinalizadora do controle do balanço entre o ciclo celular normal e a apoptose (19, 20).

Em células β pancreáticas, o NF-κB controla diversas redes de genes que contribuem para a apoptose por ativação do estresse do retículo endoplasmático (21). Sabe-se que a exposição das células β a IL-1β *in vitro* induz a disfunção celular e, em combinação com IFN-γ ou TNF-α, provoca a morte celular, principalmente por apoptose. A inibição *in vitro* do NF-κB nessas células leva à diminuição da morte celular induzida por citocinas (22).

Chen *et al.* demonstraram, em ilhotas de porcos, que a atividade do NF-κB induzida por citocinas e sob condições de hipóxia desempenha um papel negativo sobre as ilhotas pancreáticas, levando à apoptose (23). Um estudo realizado em camundongos transgênicos que expressavam um super-repressor do NF-κB relatou uma diminuição do desenvolvimento de diabetes em resposta a múltiplas doses de estreptozotocina (24).

Os estudos que relacionam o fator de transcrição NF-κB com a apoptose das células β são direcionados à compreensão da fisiopatologia do DM tipo 1 (25, 26).

Entretanto, nenhum estudo avaliou o papel da ME na ativação do NF-κB no tecido pancreático humano e seu potencial efeito na apoptose das ilhotas. Desta forma, a amostra de pacientes do estudo caso-controle objeto desta tese está sendo ampliada, com o objetivo de comparar os efeitos da ME na apoptose pela ativação da via do NF-κB no tecido pancreático.

Por fim, a ME está implicada no desenvolvimento de disfunção primária dos enxertos (DPE) transplantados (27, 28), de maneira não completamente compreendida e provavelmente subestimada. A injúria de isquemia-reperfusão é uma causa reconhecida de DPE (29, 30). A atividade inflamatória induzida pela ME parece ter um papel intensificador da lesão de isquemia-reperfusão, por aumento de espécies reativas de oxigênio. Especula-se que os agravos da ME possam tornar os órgãos mais suscetíveis aos efeitos deletérios da isquemia-reperfusão (31).

A proteína desacopladora 2 (do inglês *uncoupling protein 2*, UCP-2) é uma proteína inserida na membrana mitocondrial interna e faz parte de uma superfamília de proteínas transportadoras. A UCP-2 tem uma distribuição tecidual ampla e atua desacoplando a oxidação dos substratos da síntese de ATP, dissipando a energia do potencial de membrana e, consequentemente, diminuindo a produção de ATP pela mitocôndria. Esse desacoplamento está associado à regulação do metabolismo de ácidos graxos livres e à diminuição da formação de espécies reativas de oxigênio pela mitocôndria (32, 33).

O aumento da expressão da UCP-2 parece ter um efeito antiapoptótico na maioria dos tipos celulares, uma vez que reduz a produção de espécies reativas de oxigênio. Entretanto, acredita-se que o papel da UCP-2 possa ser tanto pró-apoptótico quanto antiapoptótico, dependendo da regulação transcricional, do tipo celular e dos diferentes estímulos bioquímicos (34). De fato, o papel da UCP-2 na apoptose das

células β pancreáticas é ainda bastante controverso. Alguns estudos indicam que a expressão aumentada da UCP-2 teria uma função antiapoptótica, por proteger as células do estresse oxidativo (35-37). Outros estudos relatam justamente o contrário: que o bloqueio da UCP-2 teria um efeito pró-apoptótico (38, 39).

Considerando-se a indefinição do papel da UCP-2 e de sua resposta aos estímulos inflamatórios da ME nas células β, decidimos avaliar o efeito do bloqueio e da superexpressão da UCP-2 na viabilidade, na função e na expressão de genes pró e antiapoptóticos de células INS-1E e de ilhotas pancreáticas humanas submetidas a diferentes condições pró-inflamatórias e pró-apoptóticas.

Frente a todo o exposto, concluímos que ainda há muito a estudar sobre a ME e seus efeitos no tecido pancreático, especialmente nas células β.

Referências

- Scharp DW, Lacy PE, Santiago JV, McCullough CS, Weide LG, Falqui L, et al. Insulin independence after islet transplantation into type I diabetic patient. Diabetes. 1990;39(4):515-8. Epub 1990/04/01.
- 2. Robertson RP. Islet transplantation: travels up the learning curve. Curr Diab Rep. 2002;2(4):365-70. Epub 2003/03/20.
- Robertson RP. Islet transplantation as a treatment for diabetes a work in progress.
 N Engl J Med. 2004;350(7):694-705. Epub 2004/02/13.
- 4. Holst JJ, Orskov C, Nielsen OV, Schwartz TW. Truncated glucagon-like peptide I, an insulin-releasing hormone from the distal gut. FEBS Lett. 1987;211(2):169-74. Epub 1987/01/26.
- 5. Kreymann B, Williams G, Ghatei MA, Bloom SR. Glucagon-like peptide-1 7-36: a physiological incretin in man. Lancet. 1987;2(8571):1300-4. Epub 1987/12/05.
- DeFronzo RA, Ratner RE, Han J, Kim DD, Fineman MS, Baron AD. Effects of exenatide (exendin-4) on glycemic control and weight over 30 weeks in metformintreated patients with type 2 diabetes. Diabetes Care. 2005;28(5):1092-100. Epub 2005/04/28.
- 7. Heine RJ, Van Gaal LF, Johns D, Mihm MJ, Widel MH, Brodows RG. Exenatide versus insulin glargine in patients with suboptimally controlled type 2 diabetes: a randomized trial. Ann Intern Med. 2005;143(8):559-69. Epub 2005/10/19.
- 8. Kendall DM, Riddle MC, Rosenstock J, Zhuang D, Kim DD, Fineman MS, et al. Effects of exenatide (exendin-4) on glycemic control over 30 weeks in patients with type 2 diabetes treated with metformin and a sulfonylurea. Diabetes Care. 2005;28(5):1083-91. Epub 2005/04/28.

- 9. Farilla L, Bulotta A, Hirshberg B, Li Calzi S, Khoury N, Noushmehr H, et al. Glucagon-like peptide 1 inhibits cell apoptosis and improves glucose responsiveness of freshly isolated human islets. Endocrinology. 2003;144(12):5149-58. Epub 2003/09/10.
- 10. Hui H, Nourparvar A, Zhao X, Perfetti R. Glucagon-like peptide-1 inhibits apoptosis of insulin-secreting cells via a cyclic 5'-adenosine monophosphate-dependent protein kinase A- and a phosphatidylinositol 3-kinase-dependent pathway. Endocrinology. 2003;144(4):1444-55. Epub 2003/03/18.
- 11. Gedulin BR, Nikoulina SE, Smith PA, Gedulin G, Nielsen LL, Baron AD, et al. Exenatide (exendin-4) improves insulin sensitivity and β-cell mass in insulinresistant obese fa/fa Zucker rats independent of glycemia and body weight. Endocrinology. 2005;146(4):2069-76. Epub 2004/12/25.
- 12. Xu G, Kaneto H, Lopez-Avalos MD, Weir GC, Bonner-Weir S. GLP-1/exendin-4 facilitates beta-cell neogenesis in rat and human pancreatic ducts. Diabetes Res Clin Pract. 2006;73(1):107-10. Epub 2006/01/13.
- 13. Ghofaili KA, Fung M, Ao Z, Meloche M, Shapiro RJ, Warnock GL, et al. Effect of exenatide on beta cell function after islet transplantation in type 1 diabetes. Transplantation. 2007;83(1):24-8. Epub 2007/01/16.
- 14. Faradji RN, Tharavanij T, Messinger S, Froud T, Pileggi A, Monroy K, et al. Longterm insulin independence and improvement in insulin secretion after supplemental islet infusion under exenatide and etanercept. Transplantation. 2008;86(12):1658-65. Epub 2008/12/24.
- 15. Ferdaoussi M, Abdelli S, Yang JY, Cornu M, Niederhauser G, Favre D, et al. Exendin-4 protects beta-cells from interleukin-1 beta-induced apoptosis by

- interfering with the c-Jun NH2-terminal kinase pathway. Diabetes. 2008;57(5):1205-15. Epub 2008/02/07.
- 16. Kang JH, Chang SY, Jang HJ, Kim DB, Ryu GR, Ko SH, et al. Exendin-4 inhibits interleukin-1beta-induced iNOS expression at the protein level, but not at the transcriptional and posttranscriptional levels, in RINm5F beta-cells. J Endocrinol. 2009;202(1):65-75. Epub 2009/04/29.
- 17. Cechin SR, Perez-Alvarez I, Fenjves E, Molano RD, Pileggi A, Berggren PO, et al. Anti-inflammatory properties of exenatide in human pancreatic islets. Cell Transplant. 2011. Epub 2011/06/07.
- 18. Chen LF, Greene WC. Shaping the nuclear action of NF-kappaB. Nat Rev Mol Cell Biol. 2004;5(5):392-401. Epub 2004/05/04.
- Hayden MS, Ghosh S. Signaling to NF-kappaB. Genes Dev. 2004;18(18):2195-224. Epub 2004/09/17.
- 20. Oeckinghaus A, Hayden MS, Ghosh S. Crosstalk in NF-kappaB signaling pathways. Nat Immunol. 2011;12(8):695-708. Epub 2011/07/21.
- 21. Eizirik DL, Cardozo AK, Cnop M. The role for endoplasmic reticulum stress in diabetes mellitus. Endocr Rev. 2008;29(1):42-61. Epub 2007/12/01.
- 22. Ortis F, Pirot P, Naamane N, Kreins AY, Rasschaert J, Moore F, et al. Induction of nuclear factor-kappaB and its downstream genes by TNF-alpha and IL-1beta has a pro-apoptotic role in pancreatic beta cells. Diabetologia. 2008;51(7):1213-25. Epub 2008/05/09.
- 23. Chen C, Moreno R, Samikannu B, Bretzel RG, Schmitz ML, Linn T. Improved intraportal islet transplantation outcome by systemic IKK-beta inhibition: NF-kappaB activity in pancreatic islets depends on oxygen availability. Am J Transplant. 2011;11(2):215-24. Epub 2011/01/12.

- 24. Eldor R, Yeffet A, Baum K, Doviner V, Amar D, Ben-Neriah Y, et al. Conditional and specific NF-kappaB blockade protects pancreatic beta cells from diabetogenic agents. Proc Natl Acad Sci U S A. 2006;103(13):5072-7. Epub 2006/03/23.
- Eizirik DL, Mandrup-Poulsen T. A choice of death--the signal-transduction of immune-mediated beta-cell apoptosis. Diabetologia. 2001;44(12):2115-33. Epub 2002/01/17.
- 26. Pirot P, Cardozo AK, Eizirik DL. Mediators and mechanisms of pancreatic betacell death in type 1 diabetes. Arq Bras Endocrinol Metabol. 2008;52(2):156-65. Epub 2008/04/29.
- 27. Pratschke J, Wilhelm MJ, Laskowski I, Kusaka M, Paz D, Tullius SG, et al. The influence of donor brain death on long-term function of renal allotransplants in rats. Transplant Proc. 2001;33(1-2):693-4. Epub 2001/03/27.
- 28. Birks EJ, Owen VJ, Burton PB, Bishop AE, Banner NR, Khaghani A, et al. Tumor necrosis factor-alpha is expressed in donor heart and predicts right ventricular failure after human heart transplantation. Circulation. 2000;102(3):326-31. Epub 2000/07/19.
- 29. Avlonitis VS, Fisher AJ, Kirby JA, Dark JH. Pulmonary transplantation: the role of brain death in donor lung injury. Transplantation. 2003;75(12):1928-33. Epub 2003/06/28.
- 30. Lee JC, Christie JD. Primary graft dysfunction. Clin Chest Med. 2011;32(2):279-93. Epub 2011/04/23.
- 31. de Perrot M, Liu M, Waddell TK, Keshavjee S. Ischemia-reperfusion-induced lung injury. Am J Respir Crit Care Med. 2003;167(4):490-511. Epub 2003/02/18.

- 32. Dalgaard LT, Pedersen O. Uncoupling proteins: functional characteristics and role in the pathogenesis of obesity and Type II diabetes. Diabetologia. 2001;44(8):946-65.
- 33. Souza BM, Assmann TS, Kliemann LM, Gross JL, Canani LH, Crispim D. The role of uncoupling protein 2 (UCP2) on the development of type 2 diabetes mellitus and its chronic complications. Arq Bras Endocrinol Metabol. 2011;55(4):239-48. Epub 2011/07/23.
- 34. Jezek P. Possible physiological roles of mitochondrial uncoupling proteins--UCPn. Int J Biochem Cell Biol. 2002;34(10):1190-206. Epub 2002/07/20.
- 35. Bai Y, Onuma H, Bai X, Medvedev AV, Misukonis M, Weinberg JB, et al. Persistent nuclear factor-kappa B activation in Ucp2-/- mice leads to enhanced nitric oxide and inflammatory cytokine production. J Biol Chem. 2005;280(19):19062-9. Epub 2005/03/11.
- 36. Affourtit C, Brand MD. On the role of uncoupling protein-2 in pancreatic beta cells. Biochim Biophys Acta. 2008;1777(7-8):973-9. Epub 2008/04/25.
- 37. Collins P, Jones C, Choudhury S, Damelin L, Hodgson H. Increased expression of uncoupling protein 2 in HepG2 cells attenuates oxidative damage and apoptosis. Liver Int. 2005;25(4):880-7. Epub 2005/07/07.
- 38. Zhang D, Shen M, Mikita A, Zhang W, Liu Y, Liu Q, et al. Targeting uncoupling protein-2 improves islet graft function. Cell Transplant. 2011;20(3):421-9. Epub 2010/08/20.
- 39. Nino Fong R, Fatehi-Hassanabad Z, Lee SC, Lu H, Wheeler MB, Chan CB. Uncoupling protein-2 increases nitric oxide production and TNFAIP3 pathway activation in pancreatic islets. J Mol Endocrinol. 2011;46(3):193-204. Epub 2011/02/16.