# UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL INSTITUTO DE CIÊNCIAS BÁSICAS DA SAÚDE CURSO DE GRADUAÇÃO EM BIOMEDICINA

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Ativação de Ureases: Inferências Evolutivas via Filogenia

Porto Alegre

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### **RESUMO**

Ureases são enzimas de grande importância histórica, médica e agrícola que catalisam a hidrólise de ureia em amônia e carbamato. Estas enzimas são amplamente distribuídas em plantas, fungos e bactérias. Em todos esses organismos, a ligação de um grupo de proteínas acessórias é necessária para o real funcionamento da enzima, atuando na modificação do sítio ativo e permitindo a inserção dos íons de níquel essenciais para a catálise. O níquel é inserido no sítio ativo da urease, num processo dependente de GTP, com o auxílio de UreD / UreH, UreE, UreF, e UreG. Estas proteínas acessórias orquestram a ativação da apoproteína, fornecendo o metal apropriado e facilitando alterações conformacionais. O mecanismo de ativação e as funções de cada proteína acessória na maturação da urease não se encontram completamente elucidadas. Para obter-se uma visão geral das relações evolutivas que podem ser estabelecidas entre as proteínas acessórias e as próprias ureases, no presente estudo foi realizada uma ampla análise das sequências de aminoácidos de proteínas acessórias destas enzimas. Considerando que os resultados são preliminares, conclui-se que as relações filogenéticas aqui apresentadas demonstram que as árvores obtidas para proteínas acessórias de urease parecem seguir topologias similares àquelas obtidas para árvores da própria enzima. Além disso, os resultados de árvores quiméricas de UreG sugerem que a análise filogenética pode estar sofrendo Atração de Longos Ramos.

# 1. INTRODUÇÃO

### 1.1 Histórico: Ureases

Ureases (EC 3.5.1.5) são enzimas que catalisam a hidrólise de ureia em amônia e carbamato, que em seguida se decompõe em outra molécula de amônia e bicarbonato (**Figura 1**). Estas enzimas são amplamente distribuídas em plantas, fungos e bactérias. Em todos esses organismos, a ligação de um grupo de proteínas acessórias é necessária para a ativação da enzima (Yukl & Wilmot 2013), atuando na modificação do sítio alvo e permitindo a inserção dos íons de níquel essenciais para a catálise (Schenk et al. 2012).

$$O \\ \parallel \\ H_2N\text{-}C\text{-}NH_2\text{+}H_2O \xrightarrow{} NH_3\text{+}H_2N\text{-}C\text{-}OH$$
Urease

Figura 1: Representação simplificada da reação de hidrólise da ureia. Reação de hidrólise da ureia em amônia e carbamato, representando a catálise por urease (reação secundária de decomposição do carbamato em amônia e bicarbonato omitida). Adaptado de Mobley e colaboradores (Mobley et al. 1995).

Ureases são biomoléculas de grande importância em atividades como medicina e agricultura. Além de possuírem alto potencial de aplicabilidade tecnológica, possuem um papel histórico relevante para o desenvolvimento das ciências básicas em áreas como enzimologia, cristalografia e biologia molecular e estrutural.

Como exemplo de importância médica, pode se citar a bactéria Gram-negativa Helicobacter pylori, que se encontra presente no estômago de mais da metade da população humana (Kusters et al. 2006). Esse patógeno gástrico depende da urease para que a produção de amônia eleve localmente o pH no estômago e, dessa forma, se desfaçam as interações fracas no gel de mucina encontrado na parede do estômago (Scott et al. 2002). Sem esse gel, o patógeno consegue acessar as células epiteliais do hospedeiro e a mucosa gástrica sofre lesão devido a exposição ao ácido estomacal. Também, tem sido demostrado que a urease pode ser um fator de virulência essencial para várias doenças, incluindo doenças de longa duração/crônicas (Konieczna et al. 2012) até mesmo de natureza extragástricas (Banić et al. 2012).

Na agricultura, ureases vegetais degradam a ureia, metabólito de plantas e fertilizante, agindo como importante fator no ciclo do nitrogênio (Krajewska 2009); no entanto, a ureia também pode ser metabolizada pelas bactérias do solo, o que pode levar a volatilização de amônia e consequente alquilação do solo, um processo danoso para o desenvolvimento de plantas (Bremner 1995). Recentemente, Polacco e colaboradores (Polacco et al. 2013), discutiram a relação entre o níquel e as ureases em plantas e constataram que os estudos devem ser expandidos a outras plantas, uma vez que até o momento muito do que se sabe das enzimas provém de estudos com soja, batata e Arabidopsis.

Também de interesse, a urease de sementes de feijão-de-porco (*Canavalia ensiformis*) foi a primeira enzima a ser cristalizada (Sumner 1926) e a primeira proteína que se mostrou conter níquel (Dixon 1975). Dessa forma, a urease é um modelo importante que fez avançar a nossa compreensão dos mecanismos de montagem dos sítios de inserção de metais em enzimas. No entanto, tais mecanismos continuam não completamente elucidados e ainda se encontram lacunas no estudo da ativação das ureases (Polacco et al. 2013).

### 1.2 Ativação: Proteínas Acessórias de Urease

A biossíntese do metalocentro de níquel em ureases geralmente requer a participação de diversas proteínas acessórias. O níquel é inserido no sítio ativo da urease, num processo dependente de GTP, com o auxílio de UreD / UreH, UreE, UreF, e UreG (Carter et al. 2009). Estas proteínas acessórias orquestram a ativação da apoproteína, fornecendo o metal apropriado, facilitando alterações conformacionais da proteína e, eventualmente, modificações pós-traducionais necessárias (Farrugia et al. 2013).

Os mecanismos propostos até agora para cada uma das proteínas acessórias, com exceção da UreG que também foi descrita em outros modelos, foram mais estudados em *Klebsiella aerogenes* e *Helicobacter pylori*. A seguir, são resumidos alguns mecanismos e atividades associados as proteínas acessórias de urease:

- UreD/UreH: Proteína scaffold que recruta outras proteínas e facilita a ligação das mesmas e a consequente formação do complexo de ativação. Estudos demonstram que UreD/UreH se liga à urease. Embora o complexo não possua cristal depositado em bancos de dados, um modelo foi proposto através da análise de outras evidencias (Ligabue-Braun, Real-Guerra, et al. 2013) (*Figura 2*).
- UreE: Principal candidata a metalochaperona, pode estar envolvida no fornecimento de níquel de urease devido a sequência da proteína em K. aerogenes, que revela 10 resíduos de histidina na porção C-terminal de 15 resíduos, sítio conhecido de ligação de Ni<sup>2+</sup> e outros metais (Mulrooney & Hausinger 1990).
- UreF: A proteína age como um túnel da atividade de GTPase da UreG,
   de forma que integre a hidrólise de GTP a biossíntese do metalocentro,

processo que garante a fidelidade da ativação da urease (Boer & Hausinger 2012).

• UreG: Enquanto presente no complexo de ativação da urease, se observa atividade de GTPase. Quando ocorre a substituição de um resíduo chave na alça-P do motivo em que o GTP se liga em *K. aerogenes* ou *H. pylori,* a capacidade da célula produzir urease ativa é perdida (Moncrief & Hausinger 1997; Mehta et al. 2003).

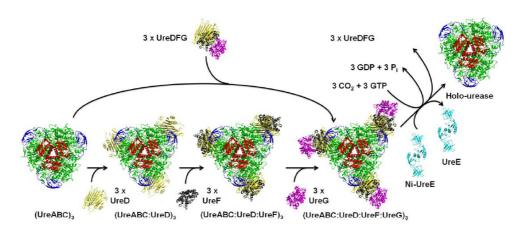


Figura 2: Modelo de ativação da urease através da formação do complexo UreABC:UreD:UreF:UreG:UreE. Modelo apresenta duas possíveis rotas de ativação: Através do acoplamento sequencial das proteínas acessórias de urease (representada nas reações abaixo na figura), ou da ligação do pré-complexo UreDFG diretamente a apoenzima (representada na reação acima na figura), seguido da ligação e transferência do Níquel pela UreE. (Farrugia et al. 2013) Baseado no modelo proposto por (Ligabue-Braun, Real-Guerra, et al. 2013).

### 1.3 Ancestralidade: Análise filogenética

São observadas algumas peculiaridades características nas ureases no que tange à sua estrutura. Geralmente, ureases de plantas e fungos apresentam estruturas de trímeros ou hexâmeros homo-oligoméricos, normalmente, com cerca de 840 resíduos de aminoácidos (Mobley et al. 1995). Ureases microbianas, por outro lado, são multímeros compostos de duas ou três subunidades ( $\alpha$ /C,  $\beta$ /B, e  $\gamma$ /A, variando entre 100 e 570 resíduos), normalmente formando trímeros de  $\alpha\beta\gamma$  ou  $\alpha\beta$  (Jabri et al. 1995; Mobley et al. 1995) (veja *Tabela 1*). Esse tipo de peculiaridade

chama atenção pela hipótese da possível fusão das subunidades, com a informação filogenética tida como ferramenta aliada na compreensão de seus possíveis caminhos evolutivos, como foi proposto por Ligabue-Braun et al. (2013).

K. aerogenes <sup>a</sup>	H. <i>pylori</i>	Plantas	Função
UreA/γ			Subunidade da enzima
UreB/β	UreA/ γ <sup>b</sup>		Subunidade da enzima
UreC/ α	UreB/β <sup>c</sup>	Urease <sup>d</sup>	Subunidade da enzima
UreD	UreH	UreD	Proteína s <i>caffold</i>
UreE	UreE	e	Metalochaperona
UreF	UreF	UreF	Possível potencializador de ligação
UreG	UreG	UreG	GTPase

**Tabela 1: Proteínas necessárias para a ativação de ureases e suas potenciais funções.** <sup>a</sup> Conjunto de proteínas comuns a maioria das bactérias. <sup>b</sup> Equivalente a fusão das subunidades A e C de *K. aerogenes.* <sup>c</sup> Equivalente a UreB de *K. aerogenes.* <sup>d</sup> Equivalente a fusão das subunidades A, B e C de *K. aerogenes.* <sup>e</sup> Nenhum ortólogo de UreE foi identificado em plantas. Adaptado de (Farrugia et al. 2013).

Para que potenciais caminhos evolutivos das proteínas acessórias de urease sejam inferidos, diferentes abordagens de análise filogenética podem ser utilizadas. A filogenética molecular, especificamente, usa como fonte de informação sequências de nucleotídeos ou aminoácidos. Esses dados são usados em diferentes modelos de algoritmos resultando em análises comparativas normalmente representadas na forma de uma árvore (filogenia ou árvore filogenética), que descreve as relações evolutivas entre as sequências. Dessa forma, é possível demonstrar a provável história evolutiva dos caracteres (proteínas, genes, genomas - chamados de unidades taxonômicas operacionais; OTUs, do inglês, *operational taxonomic units*) dos organismos incluídos nas análises. As árvores são representadas graficamente através de pontos (ou nós) ligados por linhas (ou ramos), de forma que a disposição dos mesmos defina a ancestralidade entre as OTUs (Figura 3). As OTUs, por sua vez, são representadas nos pontos terminais, unidas por linhas cujo nó interno representa

o ancestral comum mais recente desses *taxa*. Para que uma árvore evidencie o ancestral mais antigo do grupo a ser comparado, é necessária a identificação de uma raiz nas filogenias. Geralmente, o enraizamento se dá pela inclusão de uma ou diversas OTUs que representem grupos externos. Os grupos externos devem possuir ancestrais comuns com as OTUs em estudo, indicando caracteres presentes em organismos mais próximos aos ancestrais e provendo um direcionamento para a interpretação dos processos evolutivos (Braun et al. 2014)

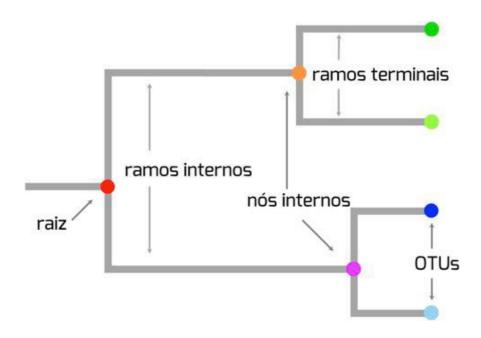


Figura 3: Modelo de nomenclatura de árvores filogenéticas. (Braun et al. 2014)

Finalmente, para que tais dados sejam analisados, é necessária a escolha dos algoritmos de reconstrução filogenética a serem usados. Vale ressaltar que a evolução é contínua e trata-se de um evento histórico, e não podemos observá-la exatamente como ocorreu ao compararmos sequências e outras características. Podemos, no entanto, inferir eventos de evolução através desses algoritmos, na chamada reconstrução filogenética. Para inferências de caráter evolutivo-molecular, podem ser

definidas quatro técnicas computacionais principais: agrupamento de vizinhos, máxima parcimônia, máxima verossimilhança e inferência Bayesiana (Yang 2006).

### 2. OBJETIVOS

O mecanismo de ativação e as funções de cada proteína acessória na maturação da urease não se encontram completamente elucidados e dependem da dedicação de diversas áreas de estudo, incluindo aplicações da bioinformática.

Para obter-se uma visão geral das relações evolutivas que podem ser estabelecidas entre as proteínas acessórias das ureases e as próprias ureases, na busca de padrões, similaridades e diferenças que possam auxiliar na predição e interpretação de resultados experimentais, o presente estudo pretende realizar uma ampla análise das sequências de aminoácidos de proteínas acessórias de ureases.

Especificamente, o trabalho visa buscar, extrair, editar e interpretar as sequências de diferentes proteínas acessórias de ureases em diferentes espécies. Dessa forma, objetiva-se construir árvores filogenéticas por meio de alinhamentos múltiplos de sequências, avaliar a similaridade e analisar possíveis regiões de conservação.

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3. ARTIGO CIENTÍFICO

Urease Activation: Evolutionary Inferences via Phylogeny

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Abstract

Ureases are enzymes are of great historic, agricultural, and medical importance. They catalyze the hydrolysis of

urea to ammonia and carbamate. These enzymes are widely distributed in plants, fungi and bacteria. In all of these

organisms, the binding of a group of accessory proteins is required for the proper functioning of the enzyme, acting

in the modification of the active site and permitting the insertion of a nickel ion, which is essential for catalysis.

The metal is inserted into the active site of urease, in a GTP-dependent process, with the aid of UreD/UreH, UreE,

UreF, and UreG: accessory proteins which orchestrate the activation of the apoprotein. The mechanisms of

activation and function of each urease accessory protein in urease maturation, are not fully understood. To obtain

an overview of evolutionary relationships that can be established between the accessory proteins and urease itself,

in this study an extensive analysis of the amino acid sequences of the accessory proteins is performed. Whereas

the results are preliminary, we conclude that the phylogenetic relationships presented here demonstrate that the

accessory proteins seem to follow similar tree topologies to those obtained for urease. Furthermore, the results of

chimeric UreG trees phylogenetic analysis suggest that they may be suffering from Long Branch Attraction.

**Keywords:** Urease accessory proteins, Phylogeny, Molecular evolution, Urease activation

### 1. Introduction

Ureases (urea amidohydrolases, EC 3.5.1.5) are enzymes that catalyze the hydrolysis of urea to ammonia and carbamate, which then decomposes into bicarbonate and another ammonia molecule (Mobley et al. 1995). Ureases are biomolecules of great importance in technological activities such as medicine and agriculture. It has been shown, for example, that urease could be a key virulence factor for various long term and chronic diseases (Konieczna et al. 2012), (Banić et al. 2012). Besides having high potential for technological applicability, they have an important historical role in the development of basic sciences in areas such as enzymology, crystallography, molecular and structural biology. Urease from jack bean seeds (*Canavalia ensiformis*) was the first enzyme to be crystallized (Sumner 1926) and the first protein shown to contain nickel (Dixon 1975). Thus, urease is an important model for the advance of our understanding of the assembly mechanisms of metallocenters.

These enzymes are widely distributed in plants, fungi, archaea, and bacteria. In all urease positive organisms, the binding of a group of urease accessory proteins (UAP) is required for the activation of the enzyme (Yukl & Wilmot 2013), working on modifying the target site and allowing the insertion of nickel ions essential for catalysis (Schenk et al. 2012). Nickel is inserted into the active site of urease with the aid of the UAPs UreD/UreH, UreE, UreF, and ureG, through a GTP-dependent process (Carter et al. 2009). These accessory proteins orchestrate the activation of the apoprotein by providing the suitable metal and facilitating conformational changes of the protein and, eventually, necessary post-translational modifications (Farrugia et al. 2013).

The mechanism of activation and function of each accessory protein in the urease maturation are not fully elucidated and depend on the dedication of several areas of study, including bioinformatics applications. To obtain an overview of evolutionary relationships that can be established between the accessory proteins of urease and urease itself, this study intends to conduct a comprehensive analysis of the amino acid sequences of the urease accessory proteins through a phylogenetic approach. We expect that by searching for patterns, similarities, and differences, the results may assist in the prediction and interpretation of experimental data in the field of urease activation and metallocenter assembly.

### 2. Materials and Methods

The urease accessory proteins amino acid sequences were retrieved from the National Center for Biotechnology Information (Sayers et al. 2012) protein database. Lists of organisms representing a broad range of differences and shown to be effective for phylogenetic reconstruction for urease were used as a guide (Ligabue-Braun et al. 2013). The keyword search comprised the given UAP followed by the species or gender name (eg. UreD + *Canavalia ensiformis*). When the "UAP + species" combination did not return a valid sequence, the species segment would be deleted and a new search would be run looking for "UAP + genus" (eg. UreD + *Canavalia*). In case of another negative response, the organism would not be included in the study. In order to avoid prediction and annotation problems, manual filtering was conducted in the resulting sequences from the query. Sequences labeled as putative, predicted, or hypothetical were excluded, as well as sequences from uncultured organisms, or with incomplete sequences (number of AA lower than 50% of the protein consensus). Resulting sequences were grouped by protein: UreD/H, UreE, UreF, and UreG, and an abbreviation was created for each genus/species (Supp. Tables 1, 2, 3, and 4).

The ClustalW (McWilliam et al. 2013) algorithm was used for alignments. In order for the most appropriate model of amino acid substitution to be calculated for each group, the resulting alignments were subjected to analysis in the Evolutionary Molecular Genetic Analysis Package 6 (MEGA6) (Tamura et al. 2013) (Table 1). Following the AA substitution model calculated for each of the alignments, phylogenetic trees were estimated by the maximum likelihood (ML) method using MEGA6 with 1,000 replicates of bootstrap. All trees were rooted using proteins from BLAST searches (Altschul et al. 1990) of each accessory protein of *Klebsiella pneumoniae* with similarity below 30% (Table 2). The obtained trees were viewed and edited through the program FigTree (Rambaut 2009). For the analysis of conservation and sequence similarity the JALVIEW (Waterhouse et al. 2009) program was used. Chimeric sequences comprising only conserved sequence segments were built using Bioedit software (Hall 1999), using results from the sequence similarity analysis.

### 3. Results

Primarily, we were able to generate four phylogenetic trees, one for each UAP, through the application of ML method on the alignments for each set of full protein sequences (Figs. 1, 2, 3, and 4) with very low bootstrap values in the more internal nodes. Resulting models of amino acid substitution for the full sequence sets are shown in Table 1. Subsequently, a chimera set was created for UreG using conserved regions with at least 35% of conservation (Fig 5). Conservation of at least 35% was not evidenced when the analysis was carried in the other

protein sets, for that reason, the following analysis continued using only UreG. The set was submitted to ML analysis, using the LG + G model and resulted in a phylogenetic tree with similar low bootstrap values, especially in internal nodes (Fig 6). Finally, we decided to take a minimalistic approach, using less representatives from each group of organisms (3 from plants, 3 from archaea, 3 from fungi, and 3 from *Helicobacter*) for a new chimera set of UreG (Fig 7). This time, higher bootstrap values were shown resulting in a resolved tree.

### 4. Discussion

Because of low bootstrap values, trees with full AA sequences are not fully resolved. The overall topology found on the cited trees, however, resulted in groupings very similar to tree topology obtained for urease (Ligabue-Braun et al. 2013). In order to evaluate whether the low bootstrap values were due to lack of conservation on some regions and/or large number of gaps, we repeated the analysis in UreG using conserved areas only. The resulting tree still showed very low bootstrap values (Fig 6), suggesting that low conservation and gap-rich regions were not the bottleneck for the resolution of the tree. Finally, the results found in figure 7, in which a smaller number of UreG AA sequences was used is more resolved and shows higher reliability in terms of bootstrap.

We could observe in an extra tree (Figure 7) that as we include more sequences in the analysis, the bootstrap values tend to diminish. Such evidence suggests that urease accessory proteins, or at least UreG, are highly heterogeneous among species. Given that sequences vary so much among closely related groups, it can be difficult to separate them apart from further groups which do not share ancestors. Therefore, we propose that the phylogenetic analysis of urease accessory proteins could be under effect of the Long Branch Attraction (LBA) (Li et al. 2007) phenomenon. When this occurs, distant groups are considered to be more closely related, which causes the algorithm to investigate such relationship more frequently, resulting in a low bootstrap number (more topologies are erroneously considered likely to be representative, so the number of times the branch organization meets a certain criteria is divided by the other likely states.). Maximum likelihood inference, although robust and free from several biases when compared to other methods for phylogenetic reconstruction, still struggles with LBA. Also, there seems to exist few analytical solutions for ML, therefore the biases it suffers are not well understood (Parks & Goldman 2014). Autapomorphies rich samples, be it nucleotide or AA sequences, currently represent an obstacle when reconstructing phylogenies. Less sequences from brother groups diminish the inconsistency that high intra species variability may lead to. The very definition of inconsistency clearly states that adding more data will only strengthen the wrong tree (Sullivan & Swofford 1997), this way, having less sequences would lead to more information, or at least more precision in the obtained tree.

### 5. Conclusion

Given that the results are preliminary, we conclude that the phylogenies presented here demonstrate that urease accessory proteins seem to follow similar tree topologies when compared to urease. The results from chimera trees of UreG suggest that the phylogeny analysis can be suffering from Long Branch Attraction. In addition, we highlight the hardship found in working with such proteins, which seem to have accumulated many differences, even within closely related groups. Further analysis will replicate the methodology used in UreG, with less sequences, for the other accessory proteins.

### 6. Acknowledgements

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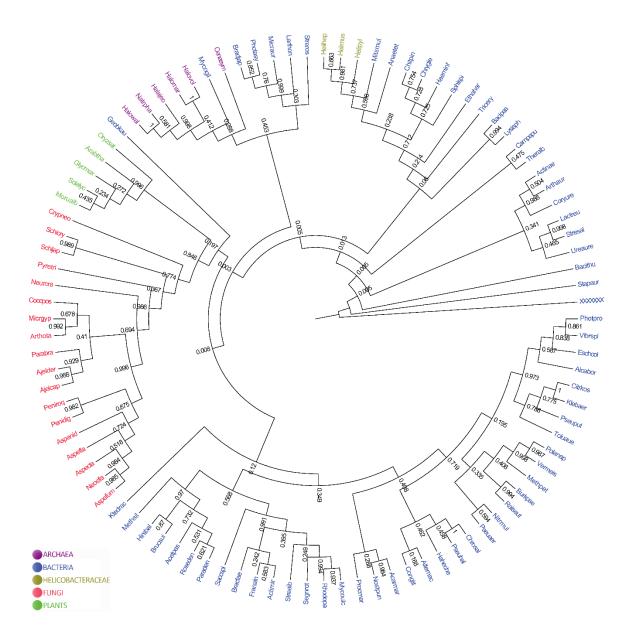
### 7. Conflict of Interest

The authors declare no conflict of interest regarding the work presented.

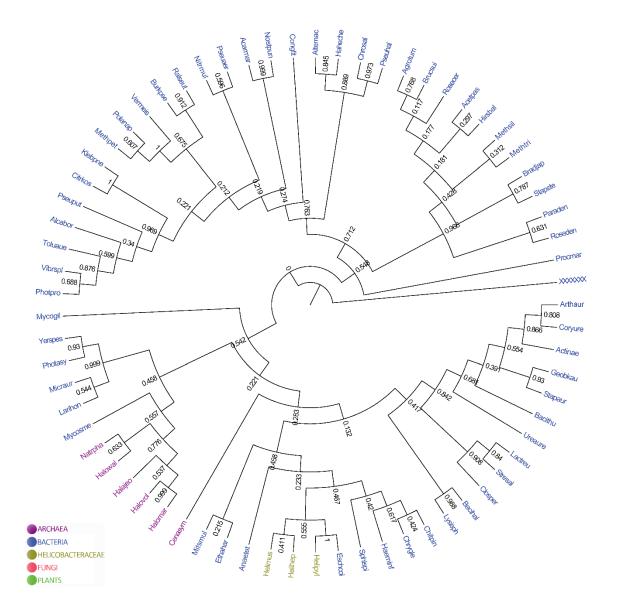
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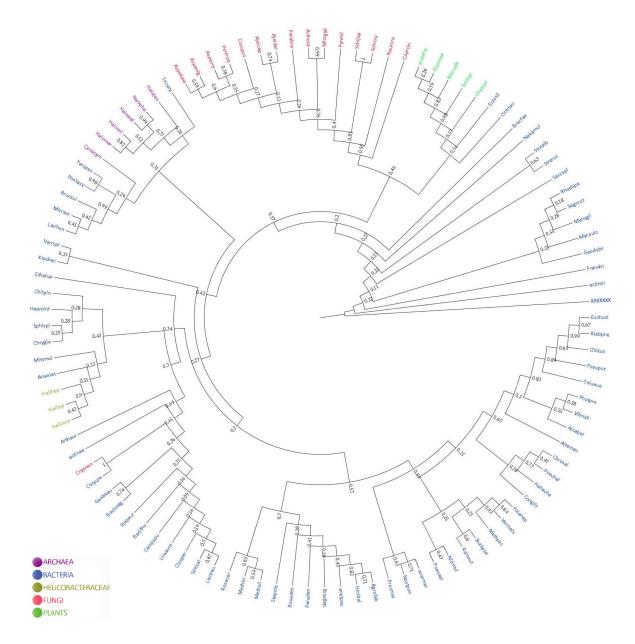
# Figures/Tables and legends



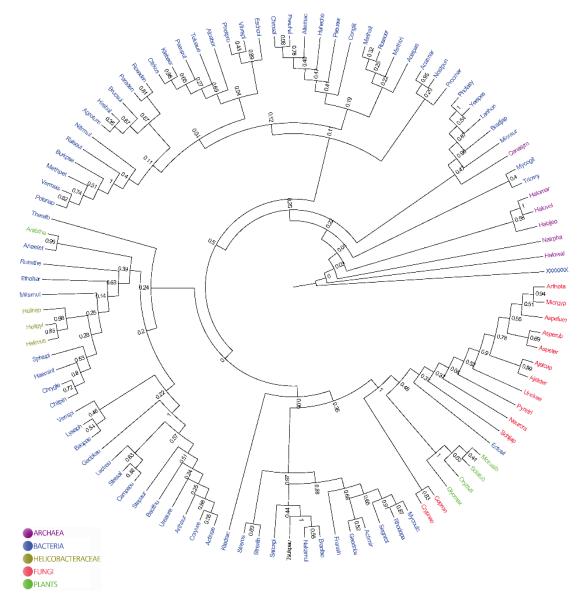
**Figure 1:** Molecular phylogenetic analysis of full UreD / H sequences by the Maximum Likelihood method. The evolutionary history was inferred using the ML method based on the model LG + G + F. Rooting sequence represented by "XXXXXXX" refers to *Methanobacterium sp.* Numbers on internodes represent the bootstrap values for the given topology.



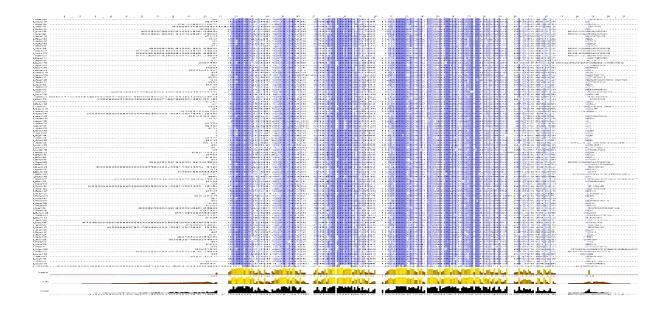
**Figure 2:** Molecular phylogenetic analysis of full UreE sequences by the Maximum Likelihood method. The evolutionary history was inferred using the ML method based on the model LG + G + I. Rooting sequence represented by "XXXXXXX" refers to *Aeromonas diversas*. Numbers on internodes represent the bootstrap values for the given topology.



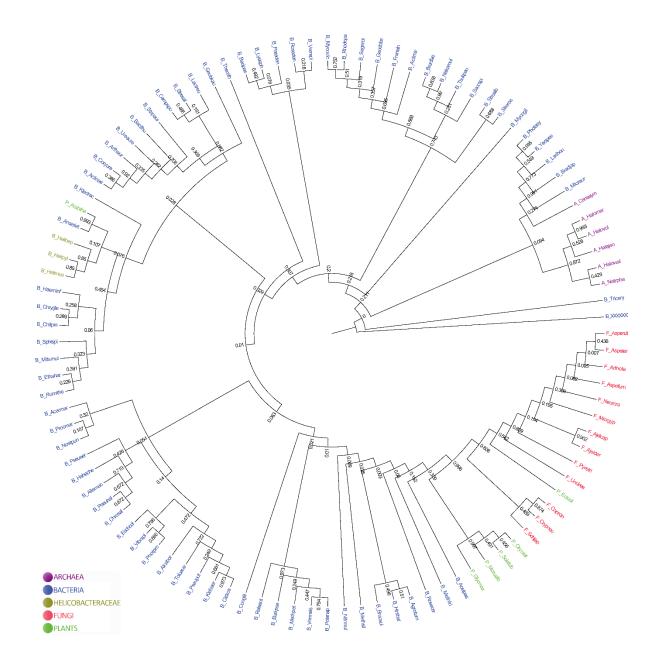
**Figure 3:** Molecular phylogenetic analysis of full UreF sequences by the Maximum Likelihood method. The evolutionary history was inferred using the ML method based on the model LG + G + I + F. Rooting sequence represented by "XXXXXXX" refers to *Aspergillus terreus*. Numbers on internodes represent the bootstrap values for the given topology.



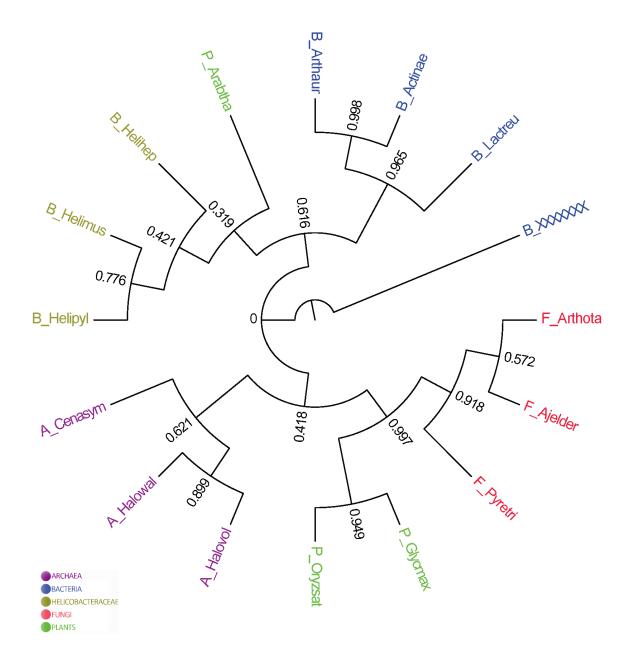
**Figure 4:** Molecular phylogenetic analysis of full UreG sequences by the Maximum Likelihood method. The evolutionary history was inferred using the ML method based on the model LG + G + I. Rooting sequence represented by "XXXXXXX" refers to *Bradyrhizobium sp.* Numbers on internodes represent the bootstrap values for the given topology.



**Figure 5:** Analysis of conserved regions of the protein UreG. Regions hatched in blue represent minimum 35% conservation.



**Figure 6:** Molecular phylogenetic analysis of chimeric UreG sequences by the Maximum Likelihood method. The evolutionary history was inferred using the ML method based on the model LG + G. Rooting sequence represented by "XXXXXXX" refers to *Bradyrhizobium sp.* Numbers on internodes represent the bootstrap values for the given topology.



**Figure 7:** Molecular phylogenetic analysis of minimal chimeric UreG sequences by the Maximum Likelihood method. The evolutionary history was inferred using the ML method based on the model LG + G. Rooting sequence represented by "XXXXXXX" refers to *Bradyrhizobium sp.* Numbers on internodes represent the bootstrap values for the given topology.

**Table 1:** Models of amino acid substitution calculated in MEGA6 program for each accessory protein group. LG: Le & Gascuel general amino acid replacement matrix (Le & Gascuel 2008). G: Gamma distribution to account for evolutionary rate differences among sites. I: Considers some sites invariable. F: Takes in consideration observed AA frequencies.

Protein Group	Substitution Model
UreD/H	LG + G + F
UreE	LG + G + I
UreF	LG + G + I + F
UreG	LG + G + I
UreG Chimera	LG + G

**Table 2:** Sequences used as external groups for rooting of each accessory protein group.

Protein Group	Root Protein and Source Organism
UreD/H	Radical SAM domain-containing protein (Methanobacterium sp.)
UreE	ABC transporter ATP-binding protein (Aeromonas diversas)
UreF	Succinyl-CoA ligase beta-chain (Aspergillus terreus)
UreG and UreG Chimera	Hydrogenase nickel incorporation protein HypB (Bradyrhizobium sp.)

# **Supplementary Tables**

**Table 1:** Table of used organisms from which UreD sequences were prospected. Sequence records are identified by GI numbers.

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Campylobacter sputorumCampspu260162329Cenarchaeum symbiosumCenasym190411588Chitinophaga pinensisChitpin502446313Chromohalobacter salexigensChrosal122419607Chryseobacterium gleumChrygle300502356Citrobacter koseriCitrkos673535173Clostridium perfringensClosper170710936Coccidioides posadasiiCoccpos320036446Congregibacter litoralisConglit495571008Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Brucella suis	Brucsui	190411640
Cenarchaeum symbiosum Chitinophaga pinensisCenasym190411588Chromohalobacter salexigens Chryseobacterium gleum Chryseobacterium gleum Citrobacter koseri CitrkosChrygle Citrkos300502356 673535173Clostridium perfringens Coccidioides posadasii Congregibacter litoralis Corynebacterium urealyticum Cryptococcus neoformans Escherichia coli Ethanoligenens harbinense Frankia alniConasym Consymbacterium urealyticum Cryptococcus neoformans EthaharCorynebacterium urealyticum Crypneo448277877 448277877Ethanoligenens harbinense Frankia alni Geobacillus kaustophilusEthahar Franaln Geobkau503251842 81703969	Burkholderia pseudomallei	Burkpse	190411638
Chitinophaga pinensisChitpin502446313Chromohalobacter salexigensChrosal122419607Chryseobacterium gleumChrygle300502356Citrobacter koseriCitrkos673535173Clostridium perfringensClosper170710936Coccidioides posadasiiCoccpos320036446Congregibacter litoralisConglit495571008Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Campylobacter sputorum	Campspu	260162329
Chromohalobacter salexigensChrosal122419607Chryseobacterium gleumChrygle300502356Citrobacter koseriCitrkos673535173Clostridium perfringensClosper170710936Coccidioides posadasiiCoccpos320036446Congregibacter litoralisConglit495571008Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Cenarchaeum symbiosum	Cenasym	190411588
Chryseobacterium gleumChrygle300502356Citrobacter koseriCitrkos673535173Clostridium perfringensClosper170710936Coccidioides posadasiiCoccpos320036446Congregibacter litoralisConglit495571008Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Chitinophaga pinensis	Chitpin	502446313
Citrobacter koseri Clostridium perfringens Closper Closper Closper Closper Closper T70710936 Coccidioides posadasii Coccpos Congregibacter litoralis Conglit Corynebacterium urealyticum Coryure Cryptococcus neoformans Escherichia coli Ethanoligenens harbinense Frankia alni Franaln Ceobacillus kaustophilus Citrkos Closper 170710936 Coccpos 320036446 Coryure 448277877 Cryptococcus neoformans Crypneo 405120208 Eschcol 418163 Ethanoligenens harbinense Frankia alni Franaln 205830842	Chromohalobacter salexigens	Chrosal	122419607
Clostridium perfringensClosper170710936Coccidioides posadasiiCoccpos320036446Congregibacter litoralisConglit495571008Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Chryseobacterium gleum	Chrygle	300502356
Coccidioides posadasii Coccpos 320036446 Congregibacter litoralis Conglit 495571008 Corynebacterium urealyticum Coryure 448277877 Cryptococcus neoformans Crypneo 405120208 Escherichia coli Eschcol 418163 Ethanoligenens harbinense Ethahar 503251842 Frankia alni Franaln 205830842 Geobacillus kaustophilus Geobkau 81703969	Citrobacter koseri	Citrkos	673535173
Congregibacter litoralisConglit495571008Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Clostridium perfringens	Closper	170710936
Corynebacterium urealyticumCoryure448277877Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Coccidioides posadasii	Coccpos	320036446
Cryptococcus neoformansCrypneo405120208Escherichia coliEschcol418163Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Congregibacter litoralis	Conglit	495571008
Escherichia coli Eschcol 418163  Ethanoligenens harbinense Frankia alni Franaln 205830842  Geobacillus kaustophilus Geobkau 81703969	Corynebacterium urealyticum	Coryure	448277877
Ethanoligenens harbinenseEthahar503251842Frankia alniFranaln205830842Geobacillus kaustophilusGeobkau81703969	Cryptococcus neoformans	Crypneo	405120208
Frankia alni Franaln 205830842 Geobacillus kaustophilus Geobkau 81703969	Escherichia coli	Eschcol	418163
Geobacillus kaustophilus Geobkau 81703969	Ethanoligenens harbinense	Ethahar	503251842
•	Frankia alni	Franaln	205830842
Glycine max Glycmax 351727327	Geobacillus kaustophilus	Geobkau	81703969
	Glycine max	Glycmax	351727327

Haemophilus influenzae Hahella chejuensis	Haheche	
		190411635
Halalkalicoccus jeotgali	Halajeo	495693486
Haloarcula marismortui	Halomar	34419200
Haloferax volcanii	Halovol	291372705
Haloquadratum walsbyi	Halowal	121689508
Helicobacter hepaticus	Helihep	14579325
Helicobacter mustelae	Helimus	502787689
Helicobacter pylori	Helipyl	332672906
Hirschia baltica	Hirsbal	506308782
Klebsiella aerogenes	Klebaer	731078
Ktedonobacter racemifer	Ktedrac	495193921
Lactobacillus reuteri	Lactreu	659901712
Laribacter hongkongensis	Larihon	654310527
Lysinibacillus sphaericus	Lysisph	205830835
Methylibium petroleiphilum	Methpet	190411631
Methylocella silvestris	Methsil	501586734
Micromonospora aurantiaca	Micraur	503052676
Microsporum gypseum	Micrgyp	311343242
Mitsuokella multacida	Mitsmul	260850227
Morus alba	Morualb	222143566
Mycobacterium gilvum	Mycogil	190411605
Mycobacterium ulcerans	Mycoulc	118570527
Natronomonas pharaonis	Natrpha	121695537
Neosartorya fischeri	Neosfis	119409225
Neurospora crassa	Neurcra	28922161
Nitrosospira multiformis	Nitrmul	190411630
Nostoc punctiforme	Nostpun	205830823
Oryza sativa	Oryzsat	301344549
Paracoccidioides brasiliensis	Parabra	226291919
Paracoccus denitrificans	Paraden	190411606
Penicillium digitatum	Penidig	425769194
Penicillium roqueforti	Peniroq	584412396
Photorhabdus asymbiotica	Photasy	506314736
Photobacterium profundum	Photpro	493270411
Polaromonas	Polanap	190411607
naphthalenivorans Prochlorococcus marinus	Procmar	7839378
Pseudomonas aeruginosa	Pseuaer	81539701
Pseudoalteromonas	Pseuhal	123588265
haloplanktis		
Pseudomonas putida	Pseuput	81440943
Pyrenophora tritici_repentis	Pyretri	187982937
Ralstonia eutropha	Ralseut	7388345
Rhodococcus opacus	Rhodopa	226365152
Roseobacter denitrificans	Roseden	123172087
Saccharopolyspora spinosa	Saccspi	41350150

Schizosaccharomyces cryophilus	Schicry	528316377
Schizosaccharomyces japonicus	Schijap	213410395
Segniliparus rotundus	Segnrot	502903350
Solanum lycopersicum	Solalyc	350537735
Sphingobacterium spiritivorum	Sphispi	300760796
Staphylococcus aureus	Stapaur	384231297
Streptomyces albus	Strealb	664085801
Streptosporangium roseum	Streros	665590112
Streptococcus salivarius	Stresal	2501636
Thermocrinis albus	Theralb	502756781
Tolumonas auensis	Toluaue	506358985
Trichodesmium erythraeum	Tricery	122965194
Ureaplasma urealyticum	Ureaure	254797563
Verminephrobacter eiseniae	Vermeis	190411617
Vibrio splendidus	Vibrspl	518658520
Yersinia pestis	Yerspes	3901302

**Table 2:** Table of used organisms from which UreE sequences were prospected. Sequence records are identified by GI numbers.

Species	Abbreviation	GI
Acaryochloris marina	Acarmar	501111920
Acetobacter pasteurianus	Acetpas	256652743
Actinomyces naeslundii	Actinae	9789805
Agrobacterium tumefaciens	Agrotum	635308178
Alcanivorax borkumensis	Alcabor	110835579
Alteromonas macleodii	Altemac	226695780
Anaerococcus tetradius	Anaetet	490976022
Arthrobacter aurescens	Arthaur	167012849
Bacillus halodurans	Bacihal	81788163
Bacillus thuringiensis	Bacithu	595878209
Bradyrhizobium japonicum	Bradjap	654719640
Brucella suis	Brucsui	597795047
Burkholderia pseudomallei	Burkpse	157939456
Cenarchaeum symbiosum	Cenasym	503247054
Chitinophaga pinensis	Chitpin	256420520
Chromohalobacter salexigens	Chrosal	499826866
Chryseobacterium gleum	Chrygle	300502353
Citrobacter koseri	Citrkos	673535177
Clostridium perfringens	Closper	170662092
Congregibacter litoralis	Conglit	563352888
Corynebacterium urealyticum	Coryure	226695783
Escherichia coli	Eschcol	485333
Ethanoligenens harbinense	Ethahar	503250833
Geobacillus kaustophilus	Geobkau	81703968
Haemophilus influenzae	Haeminf	16272482
Hahella chejuensis	Haheche	83635257
Halalkalicoccus jeotgali	Halajeo	495693487
Haloarcula marismortui	Halomar	34419201
Haloferax volcanii	Halovol	291372442
Haloquadratum walsbyi	Halowal	385804940
Helicobacter hepaticus	Helihep	14579322
Helicobacter mustelae	Helimus	291276541
Helicobacter pylori	Helipyl	485333
Hirschia baltica	Hirsbal	506308777
Klebsiella pneumoniae	Klebpne	499532364
Lactobacillus reuteri	Lactreu	659901709
Laribacter hongkongensis	Larihon	654310525
Lysinibacillus sphaericus	Lysisph	226695790
Methylibium petroleiphilum	Methpet	189037565
Methylocella silvestris	Methsil	501586730
Methylosinus trichosporium	Methtri	489708565
Micromonospora aurantiaca	Micraur	503052679
Mitsuokella multacida	Mitsmul	260850224
Mycobacterium gilvum	Mycogil	500222475

Mycobacterium smegmatis	Mycosme	118472340
Natronomonas pharaonis	Natrpha	88909693
Nitrosospira multiformis	Nitrmul	123544644
Nostoc punctiforme	Nostpun	186686176
Paracoccus denitrificans	Paraden	226695792
Photobacterium profundum	Photpro	493270419
Photorhabdus asymbiotica	Photasy	506314739
Polaromonas naphthalenivorans	Polanap	189037567
Prochlorococcus marinus	Procmar	7839379
Pseudoalteromonas haloplanktis	Pseuhal	123588264
Pseudomonas aeruginosa	Pseuaer	15600084
Pseudomonas putida	Pseuput	313499134
Ralstonia eutropha	Ralseut	9789778
Roseobacter denitrificans	Roseden	109457062
Roseomonas cervicalis	Rosecer	296263240
Sphingobacterium spiritivorum	Sphispi	300760793
Staphylococcus aureus	Stapaur	384231294
Stappia stellulata	Stapste	656018176
Streptococcus salivarius	Stresal	363548486
Tolumonas auensis	Toluaue	259710157
Ureaplasma urealyticum	Ureaure	7316078
Verminephrobacter eiseniae	Vermeis	189037574
Vibrio splendidus	Vibrspl	518658517
Yersinia pestis	Yerspes	6686076

**Table 3:** Table of used organisms from which UreF sequences were prospected. Sequence records are identified by GI numbers.

Species	Abbreviation	GI
Acaryochloris marina	Acarmar	205830115
Acetobacter pasteurianus	Acetpas	256652742
Actinosynnema mirum	Actimir	502426242
Actinomyces naeslundii	Actinae	4249615
Agrobacterium fabrum	Agrofab	205830125
Ajellomyces capsulatus	Ajelcap	225554742
Ajellomyces dermatitidis	Ajelder	327354766
Alcanivorax borkumensis	Alcabor	499909253
Alteromonas macleodii	Altemac	522831428
Anaerococcus tetradius	Anaetet	227216937
Arabidopsis thaliana	Arabtha	15219116
Arthrobacter aurescens	Arthaur	205830117
Arthroderma otae	Arthota	238842926
Aspergillus kawachii	Aspekaw	358369688
Aspergillus niger	Aspenig	317035317
Aspergillus oryzae	Aspeory	317148887
Bacillus megaterium	Bacimeg	294499714
Bacillus thuringiensis	Bacithu	595878208
Brachybacterium faecium	Bracfae	502488432
Bradyrhizobium japonicum	Bradjap	658448727
Brucella suis	Brucsui	648132136
Burkholderia pseudomallei	Burkpse	81379229
Campylobacter sputorum	Campspu	260162334
Cenarchaeum symbiosum	Cenasym	503247053
Chitinophaga pinensis	Chitpin	502446311
Chromohalobacter salexigens	Chrosal	122419611
Chryseobacterium gleum	Chrygle	300502354
Citrobacter koseri	Citrkos	673535178
Clostridium perfringens	Closper	170662151
Coccidioides posadasii	Coccpos	320034975
Congregibacter litoralis	Conglit	495571013
Coprinopsis cinerea	Coprcin	169861560
Corynebacterium urealyticum	Coryure	501329375
Cryptococcus neoformans	Crypneo	448277875
Ectocarpus siliculosus	Ectosil	298711027
Escherichia coli	Eschcol	15830580
Ethanoligenens harbinense	Ethahar	503251844
Frankia alni	Franaln	111220897
Geobacillus kaustophilus	Geobkau	81703969
Geodermatophilus obscurus	Geodobs	502711312
Glycine max	Glycmax	351726506
Haemophilus influenzae	Haeminf	30995384

Hahella chejuensis	Haheche	123531909
Halalkalicoccus jeotgali	Halajeo	495693488
Haloarcula marismortui	Halomar	34419202
Haloferax volcanii	Halovol	291371470
Haloquadratum walsbyi	Halowal	385804941
Helicobacter hepaticus	Helihep	14579323
Helicobacter mustelae	Helimus	291276542
Helicobacter pylori	Helipyl	485334
Hirschia baltica	Hirsbal	506308776
Klebsiella pneumoniae	Klebpne	641638415
Ktedonobacter racemifer	Ktedrac	495193920
Lactobacillus reuteri	Lactreu	659901710
Laribacter hongkongensis	Larihon	254797573
Lysinibacillus sphaericus	Lysisph	659841406
Methylibium petroleiphilum	Methpet	205830511
Methylocella silvestris	Methsil	501586729
Methylosinus trichosporium	Methtri	639846881
Micromonospora aurantiaca	Micraur	503052678
Microsporum gypseum	Micrgyp	311344260
Mitsuokella multacida	Mitsmul	260850225
Morus alba	Morualb	222143564
Mycobacterium gilvum	Mycogil	503236987
Mycobacterium ulcerans	Mycoulc	118570529
Nakamurella multipartita	Nakamul	502530429
Natronomonas pharaonis	Natrpha	76557520
Neurospora crassa	Neurcra	28923098
Nitrosospira multiformis	Nitrmul	123727174
Nostoc punctiforme	Nostpun	501380802
Oryza sativa	Oryzsat	301344551
Ostreococcus tauri	Ostrtau	308812025
Paracoccidioides brasiliensis	Parabra	226294758
Paracoccus denitrificans	Paraden	205830515
Penicillium roqueforti	Peniroq	584407181
Photorhabdus asymbiotica	Photasy	253989857
Photobacterium profundum	Photpro	493270423
Polaromonas	Polanap	500124378
naphthalenivorans Prochlorococcus marinus	Procmar	7839380
subsp. pastoris	110011111	, 00,000
Pseudomonas aeruginosa	Pseuaer	15600085
Pseudoalteromonas	Pseuhal	205830548
haloplanktis Pseudomonas putida	Pseuput	81440939
Pyrenophora tritici-repentis	Pyretri	187973256
Ralstonia eutropha	Ralseut	123329427
Rhodococcus opacus	Rhodopa	226365150
Roseomonas cervicalis	Rosecer	296263241
Roseobacter denitrificans	Roseden	123172086
noscooncier activity teams	110000011	1231,2000

Saccharopolyspora spinosa	Saccspi	498382100
Schizosaccharomyces cryophilus	Schicry	528315656
Schizosaccharomyces japonicus	Schijap	213410148
Segniliparus rotundus	Segnrot	502903714
Solanum lycopersicum	Solalyc	350535509
Sphingobacterium spiritivorum	Sphispi	300760794
Staphylococcus aureus	Stapaur	384231295
Stappia stellulata	Stapste	656018177
Streptomyces albus	Strealb	664085795
Streptosporangium roseum	Streros	665607552
Streptococcus salivarius	Stresal	2501638
Tolumonas auensis	Toluaue	506358979
Trichodesmium erythraeum	Tricery	123161225
Ureaplasma urealyticum	Ureaure	7272375
Verminephrobacter eiseniae	Vermeis	500133480
Verrucomicrobium spinosum	Verrspi	497646624
Vibrio splendidus	Vibrspl	518658515
Yersinia pestis	Yerspes	270338254

**Table 4:** Table of used organisms from which UreG sequences were prospected. Sequence records are identified by GI numbers.

Species	Abbreviation	GI
Acaryochloris marina	Acarmar	158304294
Acetobacter pasteurianus	Acetpas	256652741
Actinomyces naeslundii	Actinae	4249616
Actinosynnema mirum	Actimir	502426241
Agrobacterium tumefaciens	Agrotum	635308180
Ajellomyces capsulatus	Ajelcap	150411856
Ajellomyces dermatitidis	Ajelder	327348970
Alcanivorax borkumensis	Alcabor	123149169
Alteromonas macleodii	Altemac	226731813
Anaerococcus tetradius	Anaetet	227216938
Arabidopsis thaliana	Arabtha	227216938
Arthrobacter aurescens	Arthaur	119949672
Arthroderma otae	Arthota	238841889
Aspergillus ruber	Asperub	599158687
Aspergillus fumigatus	Aspefum	666427728
Aspergillus terreus	Aspeter	114196305
Bacillus pasteurii	Bacipas	75420971
Bacillus thuringiensis	Bacithu	595878207
Brachybacterium faecium	Bracfae	502488430
Bradyrhizobium japonicum	Bradjap	654722573
Brucella suis	Brucsui	673468322
Burkholderia pseudomallei	Burkpse	686978736
Campylobacter sputorum	Campspu	260162328
Cenarchaeum symbiosum	Cenasym	205830783
Chitinophaga pinensis	Chitpin	502446312
Chromohalobacter salexigens	Chrosal	499826864
Chryseobacterium gleum	Chrygle	300502355
Citrobacter koseri	Citrkos	673535179
Ruminoclostridium		
thermocellum	Rumithe	551234031
Congregibacter litoralis	Conglit	495571014
Coprinopsis cinerea	Coprcin	299743607
Corynebacterium urealyticum	Coryure	226731818
Cryptococcus neoformans	Crypneo	405118032
Ectocarpus siliculosus	Ectosil	299470337
Escherichia coli	Eschcol	418165
Ethanoligenens harbinense	Ethahar	503251843
Frankia alni	Franaln	123338675
Geobacillus kaustophilus	Geobkau	47076813
Geodermatophilus obscurus	Geodobs	502711313
Glycine max	Glycmax	351721504
Haemophilus influenzae	Haeminf	81336461
Hahella chejuensis	Haheche	123531908

Halalkalicoccus jeotgali	Halajeo	502964396
Haloarcula marismortui	Halomar	34419199
Haloferax volcanii	Halovol	291370919
Haloquadratum walsbyi	Halowal	121689509
Helicobacter hepaticus	Helihep	14579324
Helicobacter mustelae	Helimus	291276543
Helicobacter pylori	Helipyl	485335
Hirschia baltica	Hirsbal	506308775
Klebsiella aerogenes	Klebaer	137099
Ktedonobacter racemifer	Ktedrac	495193935
Lactobacillus reuteri	Lactreu	689724955
Laribacter hongkongensis	Larihon	226714896
Lysinibacillus sphaericus	Lysisph	226731825
Methylibium petroleiphilum	Methpet	205830793
Methylocella silvestris	Methsil	254797581
Methylosinus trichosporium	Methtri	489708561
Micromonospora aurantiaca	Micraur	302571835
Microsporum gypseum	Micrgyp	311340034
Mitsuokella multacida	Mitsmul	260850226
Morus alba	Morualb	222143562
Mycobacterium gilvum	Mycogil	205830796
Mycobacterium ulcerans	Mycoulc	226732038
Nakamurella multipartita	Nakamul	502530430
Natronomonas pharaonis	Natrpha	121721884
Neurospora crassa	Neurcra	28918042
Nitrosospira multiformis	Nitrmul	123544643
Nostoc punctiforme	Nostpun	186468626
Oryza sativa	Oryzsat	301344553
Paracoccus denitrificans	Paraden	205830798
Photobacterium profundum	Photpro	493270425
Photorhabdus asymbiotica	Photasy	253989856
Polaromonas		
naphthalenivorans	Polanap	226732041
Prochlorococcus marinus Pseudoalteromonas	Procmar	7839381
haloplanktis	Pseuhal	499647704
Pseudomonas aeruginosa	Pseuaer	674745349
Pseudomonas putida	Pseuput	313499131
Pyrenophora tritici_repentis	Pyretri	187983079
Ralstonia eutropha	Ralseut	123329424
Rhodococcus opacus	Rhodopa	254797584
Roseobacter denitrificans	Roseden	109457064
Roseomonas cervicalis	Rosecer	296263242
Saccharopolyspora spinosa	Saccspi	41350151
Schizosaccharomyces	-	
japonicus	Schijap	213409990
Segniliparus rotundus	Segnrot	502903349

Solanum tuberosum Sphingobacterium
spiritivorum
Staphylococcus aureus
Streptococcus salivarius
Streptomyces albus
Streptosporangium roseum
Thermocrinis albus
Tolumonas auensis
Trichodesmium erythraeum
Tsukamurella paurometabola
Uncinocarpus reesii
Ureaplasma urealyticum
Verminephrobacter eiseniae
Verrucomicrobium spinosum
Vibrio splendidus

Solatub	13161904
Sphispi	300760795
Stapaur	384231296
Stresal	2501641
Strealb	664085798
Streros	665607548
Theralb	502756782
Toluaue	259710309
Tricery	123161226
Tsukpau	502893014
Unciree	237907169
Ureaure	7272376
Vermeis	500133479
Verrspi	656243735
Vibrspl	515641537

# **3 CONCLUSÃO E PERSPECTIVAS**

Dado que os resultados são preliminares, conclui-se que as relações filogenéticas apresentadas demonstram que as proteínas acessórias de urease parecem seguir as topologias das árvores obtidas na reconstrução filogenética para ureases. Os resultados de árvores quiméricas para UreG sugerem que a análise filogenética pode estar sofrendo atração de ramos longos (LBA, *long branch attraction*) (Li et al. 2007). Além disso, destacam-se as dificuldades encontradas no trabalho com essas proteínas, as quais parecem ter acumulado muitas alterações, dentro de seus próprios grupos homólogos.

Como perspectivas para este trabalho, uma análise mais aprofundada poderá replicar a metodologia utilizada para UreG, com menos sequências, para outras proteínas acessórias, mesmo que nestas não se encontrem regiões tão conservadas. Além disso, também poderão ser feitas análises de inferência bayesiana e a identificação caso a caso de OTUs que possam estar causando vieses nas análises (especialmente aquelas com potencial a causar LBA).

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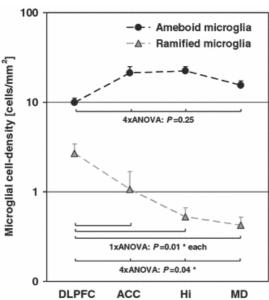
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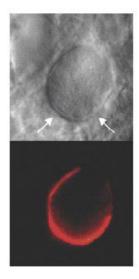
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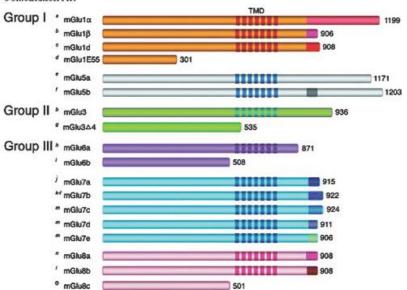
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