

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
CENTRO DE BIOTECNOLOGIA
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOLOGIA CELULAR E MOLECULAR

**ESTRUTURA DO GENE *VvAGL11* E ANÁLISE DA EXPRESSÃO DURANTE A
MORFOGÊNESE DA SEMENTE DE VIDEIRA**

Dissertação de Mestrado

Jaiana Malabarba

Porto Alegre, outubro de 2014

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Dissertação submetida ao Programa de Pós-graduação em Biologia Celular e Molecular do Centro de Biotecnologia da Universidade Federal do Rio Grande do Sul como requisito parcial para obtenção do título de Mestre.

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“There are many hypotheses in science which are wrong.
That is perfectly all right: it’s the aperture to finding out what’s right.
Science is a self-correcting process.”

Carl Sagan

“There's real poetry in the real world.
Science is the poetry of reality.”

Richard Dawkins

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LISTA DE ABREVIATURAS, SÍMBOLOS E UNIDADES

% - percentagem

3' - extremidade 3' -hidroxila de DNA ou RNA

5' - extremidade 5' -fosfato de DNA ou RNA

A – adenilato ou ácido adenílico

AG - gene *AGAMOUS*

AGL6 – gene *AGAMOUS-LIKE 6*

AGL11 – gene *AGAMOUS-LIKE 11*

AGL15 – gene *AGAMOUS-LIKE 15*

AGL17 – gene *AGAMOUS-LIKE 17*

AP1 – gene *APETALA 1*

AP2 – gene *APETALA 2*

AP3 – gene *APETALA 3*

AtAGL11 - gene *AGAMOUS-like* de *Arabidopsis thaliana*

AVAG2 - gene *AGAMOUS 2* de *Asparagus*

BPC1 - gene *BASIC PENTACYSTEINE 1*

BS - gene *B SISTER*

C – citidilato ou ácido citidílico

cDNA – DNA complementar (do inglês, *complementary DNA*)

Col-0 - *Arabidopsis thaliana* acesso Columbia-0

DNA – ácido desoxirribonucleico (do inglês, *deoxyribonucleic acid*)

DEF - gene *DEFICIENS*

DNase – desoxirribonuclease

EgMADS1 - gene *MADS-Box 1* de *Eustoma grandiflorum*

ESTs – etiquetas ou marcas de sequências expressas (do inglês, *Expressed Sequence Tags*)

F1 - primeira geração filial (do inglês, *First Filial Generation*)

FAO – Organização das Nações Unidas para Agricultura e Alimentação (do inglês, *Food and Agriculture Organization of the United Nations*)

FBP7 - gene codificador da *FLORAL BINDING PROTEIN 7*

FBP11 - gene codificador da *FLORAL BINDING PROTEIN 11*

FLC - gene *FLOWERING LOCUS C*

FUL - gene *FRUITFULL*

G – guanidilato ou ácido guanidílico

INDEL- mutação de inserção ou deleção de nucleotídeos (do inglês *Insertion e Deletion*)

LMADS2 – gene *MADS-Box 2* de *Lilium*

Mcm1 - gene *minichromosome maintenance 1*

MAS - seleção assistida por marcadores (do inglês, *marker assisted selection*)

mRNA – RNA mensageiro (do inglês, *messenger RNA*)

OIV - Organização Internacional da Uva e do Vinho (do francês, *Organization Internationale de la Vigne e du Vin*)

OsMADS13 - gene *MADS-Box 13* de *Oryza sativa*

pb – par (es) de bases

PCR – reação em cadeia da DNA polimerase (do inglês, *polymerase chain reaction*)

PhalAG2 - gene *AGAMOUS 2* de *Phalaenopsis*

PI - gene *PISTILLATA*

Place – do inglês, *Plant Cis-acting Regulatory DNA Elements*

QTL- locus de característica quantitativa (do inglês, *quantitative trait locus*)

RNA – ácido ribonucleico (do inglês, *ribonucleic acid*)

RT-qPCR – PCR quantitativa precedida de transcrição reversa (do inglês, *reverse transcription-quantitative PCR*)

SdI - inibidor do desenvolvimento da semente (do inglês *Seed Development Inhibitor*)

SEP1 – gene *SEPALLATA 1*

SEP2 - gene *SEPALLATA 2*

SEP3 - gene *SEPALLATA 3*

SEP4 - gene *SEPALLATA 4*

SHP1 - gene *SHATTERPROOF 1*

SHP2 - gene *SHATTERPROOF 2*

SNP- polimorfismo de nucleotídeo único (do inglês *Single Nucleotide Polymorphism*)

SOC - gene *SUPPRESSOR OF OVEREXPRESSION OF CONSTANCE 1*

SRF - gene *SERUM RESPONSE FACTOR*

STK - gene *SEEDSTICK*

stk - mutante que não expressa o gene *STK/AtAGL11*

SVF - gene *SHORT VEGETATIVE PHASE*

T – timidilato ou ácido timidílico

UTR - região não traduzida (do inglês, *untranslated region*)

VvAG3 - gene *AGAMOUS 3* de *Vitis vinifera* (*VvAGL11*)

VvAGL11 - gene *AGAMOUS-LIKE 11* de *Vitis vinifera* (*VvAG3*)

WT - tipo-selvagem (do inglês, *wild-type*)

ZAG2 - gene *AGAMOUS 2* de *Zea mays*

ZMM1 - gene *MADS-BOX 1* de *Zea mays* (*ZMADS1*)

RESUMO

A videira é uma frutífera de origem antiga e atualmente é a mais cultivada do mundo, sendo de extrema importância econômica para diversos países. Certos genótipos de *Vitis vinifera* L., como a ‘Sultanina’, produzem bagas com apenas traços de sementes. Esta característica, denominada de apirenia, é classificada como estenoespermocarpia uma vez que a fecundação ocorre para a formação do fruto, porém é seguida de aborto do embrião devido à degeneração do endosperma. A apirenia é uma das características mais apreciadas para a uva de mesa e a compreensão dos mecanismos genéticos e moleculares controladores dessa característica é justificada pelo crescimento e pela exigência do mercado de uvas *in natura*. Estudos anteriores de nosso grupo permitiram identificar o gene *VvAGL11* como principal candidato atuante no controle do desenvolvimento de sementes em videira. Foi nosso objetivo, pelo presente trabalho, avaliar o gene *VvAGL11* nas cultivares pirênica Chardonnay e apirênica Sultanina, visando uma maior compreensão de seu papel durante a morfogênese da semente. Por meio de sequenciamento alelo-específico, foi possível caracterizar as variações alélicas do gene, o qual apresenta duas isoformas transcritas na cultivar pirênica Chardonnay. Em ‘Sultanina’, uma das isoformas identificadas possui mutações únicas. Pela avaliação do perfil transcricional de *VvAGL11* em diferentes estádios de desenvolvimento, da flor e do fruto, foi observado um grande acúmulo de transcritos em sementes de 2, 4 e 6 semanas em comparação com tecidos de flor e polpa. Na cultivar Sultanina, a expressão de *VvAGL11* foi extremamente baixa em todos os estádios e tecidos analisados. Análises morfo-anatômicas foram empregadas para comparar a semente e o traço de semente. Foram identificadas alterações anatômicas que sugerem a perda da identidade da camada de integumento médio do traço de semente de Sultanina, uma vez que esta camada não apresentou o padrão de diferenciação normal. O padrão de expressão espaço-temporal de *VvAGL11* foi determinado pela técnica de hibridização *in situ*. Foram utilizadas amostras das cultivares Chardonnay e Sultanina nos mesmos estádios de desenvolvimento analisados por RT-qPCR. Os níveis de transcritos de *VvAGL11* aumentaram significativamente em frutos de ‘Chardonnay’ com 2 e 4 semanas de desenvolvimento, especificamente na camada dupla do integumento médio da semente. Em ‘Sultanina’, não houve detecção de transcritos do gene em quaisquer estádios de desenvolvimento.

ABSTRACT

Grapevine have ancient origins and is currently the most widely cultivated fruit in the world, with high economic importance to many countries. Certain genotypes of *Vitis vinifera* L. like 'Sultanina' produce berries with only seed traces. This characteristic, termed apireny, is classified as stenospermocarpy since fertilization occurs aiming the formation of fruits but it is followed by the abortion of the embryo due to endosperm degeneration. Seedlessness is one of the most appreciated features for table grapes. Thereby the understanding of genetic and molecular mechanisms that control this feature is justified by the growth and the demand of *in natura* grapes' market. Our previous studies allowed us to identify a candidate gene, *VvAGL11*, possibly involved in the control of seed development in grapevine. Our objective with the present study was to evaluate *VvAGL11* in a pirenic cultivar, Chardonnay, and in an apirenic cultivar, Sultanina, in order to better understand its role during seed morphogenesis. By allele-specific sequencing, it was possible to characterize the allelic variations of *VvAGL11*, which exhibited two transcript isoforms in 'Chardonnay'. In 'Sultanina' one of the isoforms was identified with unique mutations. Evaluation of transcriptional profiles of *VvAGL11* during different developmental stages, from flowers to mature fruits, it showed a large accumulation of transcripts in seeds of 2, 4 and 6 weeks old compared to flower and pulp tissues in the Chardonnay cultivar. In 'Sultanina' the expression of *VvAGL11* was extremely low in all stages and tissues examined. Anato-morphological analyzes were performed in order to compare the seed and the seed trace. It was identified a loss of identity of the medium integument layer in the Sultanina seed trace since this layer neither elongated nor doubled in size as it should. The spatial-temporal expression pattern of *VvAGL11* was determined by an *in situ* hybridization technique. Samples of 'Chardonnay' and 'Sultanina' in the same stages of development analyzed by RT-qPCR were employed. *VvAGL11* transcript levels were significantly increased in 'Chardonnay' seeds with 2 and 4 weeks of development, specifically in the dual layer medium integument of the seed. In 'Sultanina', gene transcripts were undetectable during all stages of development.

1 INTRODUÇÃO

O desenvolvimento e a seleção de variedades de uva de mesa (*Vitis vinifera*) são considerados investimentos de longo prazo e de alto custo. A técnica de melhoramento clássico comumente utilizada envolve a geração de centenas de indivíduos F1 (do inglês, *First Filial Generation*) a cada ano, os quais necessitam de muito espaço para avaliação em campo e de longo período de cultivo para a obtenção de frutos (CABEZAS *et al.*, 2006). Todo este processo gera perdas de produtividade pela demora excessiva, de até seis anos, para a avaliação da qualidade dos frutos. Visando a inovação deste sistema, a geração de plantas que apresentem caracteres de interesse por técnicas de engenharia genética é vantajosa para os grandes mercados produtores de frutíferas de desenvolvimento prolongado, como a videira, além da utilização de ferramentas biotecnológicas, como marcadores moleculares, que auxiliem na aceleração da geração e seleção de novas cultivares.

Muitos programas de melhoramento têm mantido o foco na geração de uvas de mesa que combinem ausência de sementes com outros caracteres de interesse agrônômico no fruto tais como aumento do tamanho da baga, sabor moscatel e crocância (LOOMIS & WEINBERG, 1979; CAMARGO *et al.*, 2005). A compreensão dos mecanismos genéticos e moleculares controladores da apirenia ou ausência de sementes em uvas de mesa possui grande relevância econômica, uma vez que o mercado consumidor desta fruta vem crescendo significativamente nas últimas décadas, além de ter aumentado sua preferência por uvas apirênicas (VITTI, 2009).

Diversos autores e grupos de pesquisa buscaram compreender as bases genéticas e moleculares da apirenia. A presença de um alelo dominante na região do *locus SdI* (Inibidor do desenvolvimento da semente, do inglês, *Seed development Inhibitor*), na extremidade do cromossomo 18 de *Vitis vinifera*, foi identificada como o principal QTL (*QTL*, do inglês, *Quantitative Trait Locus*) associado à apirenia, responsável por 50 a 70% da variância fenotípica total (BOUQUET E DANGLLOT, 1996; LAHOGUE *et al.*, 1998; DOLIGEZ *et al.*, 2002; CABEZAS *et al.*, 2006; MEJÍA *et al.*, 2007; COSTANTINI *et al.*, 2008). O gene *VvAGL11* (*AGAMOUS-LIKE 11*; acesso no genoma em <http://genomes.cribi.unipd.it/grape/:Vv18s0041g01880>) está localizado no mesmo *contig* do *Locus SdI* e foi proposto como um forte candidato responsável pela ausência de sementes

em videira (COSTANTINI *et al.*, 2008; MEJÍA *et al.*, 2011; REVERS *et al.*, 2010). *VvAGL11* codifica um fator de transcrição do tipo MADS-box. Análises *in silico* permitiram sugerir uma possível ortologia entre *VvAGL11* e *SEEDSTICK* (*STK* ou *AtAGL11*) de *Arabidopsis thaliana* (acesso no genoma www.Arabidopsis.org: AT4G09960), o qual está envolvido no controle da identidade do óvulo (PINYOPICH *et al.*, 2003).

A identificação do gene *VvAGL11* localizado dentro de um *Locus* de Características Quantitativas significativamente associado à apirenia, aliado à sua ortologia com o gene *SEEDSTICK* de *Arabidopsis* justifica a caracterização deste gene no contexto da apirenia em videira. A compreensão de seu papel na morfogênese de sementes em uvas poderá ser útil no desenvolvimento de novas cultivares de uvas de mesa que atendam às exigentes demandas do mercado de consumo nacional e internacional.

1.1 MORFOGÊNESE DE SEMENTES E APIRENIA

1.1.1 Anatomia das Sementes em Desenvolvimento

As angiospermas apresentam uma estrutura especializada altamente complexa responsável pela propagação e manutenção de suas espécies, a semente. Esta é composta por três partes principais, cada qual com sua função: os integumentos, que protegem o endosperma e o embrião de situações adversas até que haja condições favoráveis para a germinação; o endosperma, que possui os nutrientes necessários para o desenvolvimento inicial do embrião; e o embrião, que se divide em duas partes, a radícula e o cotilédone, que germinarão para a formação de uma nova planta (Figura 1a; HAUGHN & CHAUDHURY, 2005).

As diversas camadas de integumentos que compõem a casca da semente (Figura 1b) possuem origem materna. Esta estrutura é formada, geralmente, por uma cutícula que se localiza acima de uma camada de epiderme e por três regiões tegumentares: integumento externo; uma camada dupla de integumento médio; e três camadas de integumento interno (PRATT, 1971; Figura 1b). Por outro lado, o endosperma e o embrião são gerados por dupla fecundação, sendo o primeiro triplóide, proveniente da fecundação da célula central

homodiplóide, e o segundo diploide, originado da fecundação da célula ovo haplóide (SITTE *et al.*, 2002).

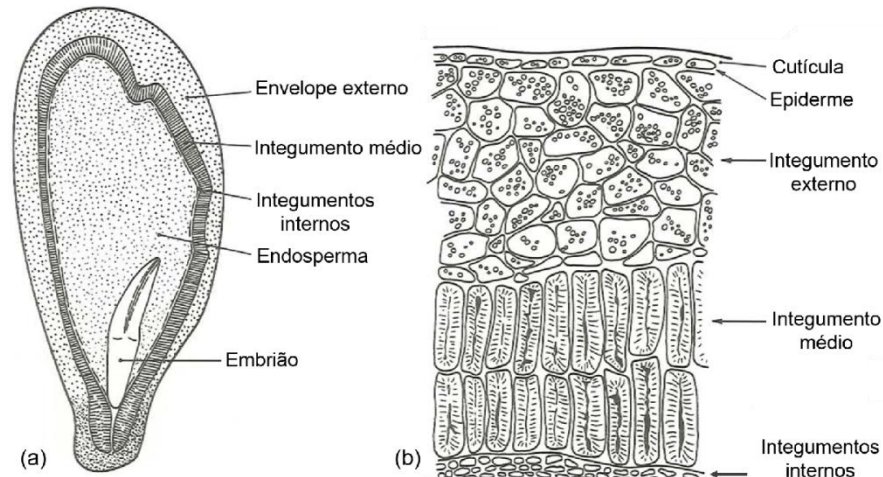


Figura 1. Ilustração da semente de *Vitis*. (a) A semente madura apresentada em corte longitudinal onde são visualizadas as camadas externas seguidas do integumento médio e de camadas de integumento interno. A porção central é composta pelo embrião circundado pelo endosperma. (b) Ilustração das camadas da casca da semente composta por uma camada de cutícula, uma camada de epiderme seguida pelo integumento externo e pelo integumento médio. Abaixo destes, iniciam-se as camadas de integumentos internos. Adaptado de PRATT (1971).

Em *A. thaliana*, o desenvolvimento da semente pode ser dividido em duas fases. A primeira é caracterizada pela proliferação e crescimento do endosperma, que aumenta o tamanho da semente em cerca de 100 vezes e traz grande contribuição para o seu tamanho final (VAROQUAUX *et al.*, 2000). Em contraste, o crescimento do embrião ocorre durante a segunda fase, quando há a expansão do endosperma (JÜRGENS & MAYER, 1994). O fluxo de nutrientes ocorre de forma centrípeta, do integumento externo para o interno, passando através de apoplastos para o endosperma e para o embrião em rotas que ainda não são completamente compreendidas (STADLER *et al.*, 2005). Ao fim da maturação da semente as camadas de integumentos morrem devido à sua lignificação, com exceção do endotélio (HAUGHN & CHAUDHURY, 2005). O crescimento da semente é, portanto, ditado pelo desenvolvimento conjunto do endosperma e dos integumentos (GARCIA *et al.*,

2005). Estudos já demonstraram que existe um forte efeito materno dos integumentos no controle e na determinação do tamanho da semente, por consequência de seu alongamento (BERGER *et al.* 2006). Deste modo, uma coordenação temporal e espacial de crescimento entre estes diferentes tecidos é crítica para o desenvolvimento correto da semente (UNGRU *et al.*, 2010).

1.1.2 Apirenia

Os processos que regem o desenvolvimento de sementes estão bem caracterizados do ponto de vista fisiológico (EGLI, 1998). Diversas culturas que possuem apelo comercial voltado para a produtividade de grãos dependem deste conhecimento, bem como do seu entendimento do ponto de vista genético e molecular, para a obtenção de boas safras ano a ano. Entretanto, em culturas onde o produto final é o fruto, uma das características mais apreciadas pelos consumidores é a ausência de sementes.

Uma planta é considerada apirênica quando é capaz de produzir frutos sem sementes, com traços de sementes ou com sementes reduzidas em número e tamanho (VAROQUAUX *et al.*, 2000). Duas formas de apirenia podem ser observadas dependendo do estágio em que a semente cessa o seu desenvolvimento, partenocarpia e estenoespermocarpia. Na partenocarpia, o ovário é capaz de se desenvolver em fruto sem que haja fecundação, apenas a partir do desenvolvimento dos tecidos maternos. Os frutos partenocárpicos são caracterizados pela ausência total de sementes. Por outro lado, na estenoespermocarpia, a fecundação ocorre normalmente e o desenvolvimento da semente se inicia. Porém, após o início de sua formação, ocorre aborto do embrião ainda imaturo devido à ausência ou má formação do endosperma, caracterizando a estenoespermocarpia pela produção de frutos com apenas traços de sementes (STOUT, 1936; BOUQUET & DANGLOT, 1996; VAROQUAUX *et al.*, 2000; MEJÍA *et al.*, 2011). Na maioria dos casos, o aborto do embrião ocorre oito semanas após a fecundação (EMERSHAD *et al.*, 1989).

Frutíferas altamente comercializadas como videiras, laranjeiras, tomateiros e caquizeiros realizam, com mais frequência, a estenoespermocarpia. Este fator agrega valor aos frutos produzidos. Por consequência, os mecanismos moleculares que regem o cessar do

desenvolvimento da semente na estenoespermocarpia são de grande importância para que haja um entendimento maior deste fenótipo.

1.1.3 Mecanismos Moleculares da Morfogênese de Sementes

O desenvolvimento dos órgãos florais e o processo de transição floral são bem conhecidos do ponto de vista molecular (SMACZNIAK *et al.*, 2012). Entre diversos componentes deste processo, os genes da família *MADS-box* possuem papel fundamental na regulação dos mecanismos que definem a identidade dos meristemas florais como sépalas, pétalas, estames, carpelos e óvulos, sendo que seus representantes são muito estudados na área de genética molecular vegetal com mais de 2.600 estudos publicados em revistas internacionais (SMACZNIAK *et al.*, 2012).

1.1.3.1 Os genes *MADS-box*

Os fatores de transcrição da família *MADS-box* possuem papéis de extrema importância no desenvolvimento de plantas, pois são responsáveis pelo controle da identidade dos órgãos florais. A origem do nome *MADS* deve-se à alta similaridade desses fatores de transcrição com a proteína humana *SRF* (do inglês, *Serum Response Factor*; NORMAN *et al.*, 1988), com o gene *Mcm1* (do inglês, *Minichromosome maintenance 1*) de *Saccharomyces cerevisiae* (PASSMORE *et al.*, 1988), com o gene *AGAMOUS* (*AG*; YANOFSKY *et al.*, 1990) de *A. thaliana* e com o gene *DEFICIENS* (*DEF*) de *Antirrhinum majus* (SOMMER *et al.*, 1990). Por este motivo, estes fatores de transcrição foram denominados de genes da família *MADS* (de *MCM1*, *AG*, *DEF* e *SRF*; SCHWARZ-SOMMER *et al.*, 1990).

Os fatores de transcrição *MADS-box* contêm domínios de ligação ao DNA e de dimerização conservados em diversas espécies, chamados de domínio *MADS* (SCHWARZ-SOMMER *et al.*, 1992; PELLEGRINI *et al.*, 1995; HUANG *et al.*, 2000). Os genes *MADS-box* são divididos em dois grupos tendo-se por base a estrutura dos seus domínios proteicos, denominadas de tipo I e de tipo II (SMACZNIAK *et al.*, 2012). Os genes *MADS-box* de estruturas do tipo I constituem um grupo bem diversificado, contendo apenas ~180 pares de bases (pb) de sequências do domínio *MADS* em comum (DE BODT *et al.*, 2003;

PARENICOVÁ *et al.*, 2003). Estudos recentes permitiram demonstrar que os genes pertencentes a este grupo possuem funções relacionadas com a gametogênese feminina e com o desenvolvimento de sementes (WUEST *et al.*, 2010).

Por outro lado, os genes *MADS-box* do tipo II foram mais bem estudados e caracterizados. Estes são genes homeóticos que atuam principalmente no desenvolvimento floral, na embriogênese e no desenvolvimento do fruto. Este grupo é caracterizado por sua estrutura denominada *MIKC*, na qual a porção N-terminal é composta pelo domínio MADS (M) de ligação ao DNA, seguido por dois domínios essenciais para a interação com proteínas, as porções I (*intervening*) e K (*keratin-like*). O domínio C-terminal é responsável pela formação e atividade de complexos proteicos e pela regulação transcricional (KAUFMANN *et al.*, 2005). Com base nas características estruturais deste domínio, os genes *MADS-box* do tipo *MIKC* foram ainda divididos em *MIKC^C* (forma canônica) e *MIKC**. Este último apresenta alterações no domínio K, possivelmente pela duplicação de éxons desta região, e forma uma rede de interação de proteínas responsável pela maturação do pólen (ADAMCZYK & FERNANDEZ, 2009).

Na planta-modelo *A. thaliana*, 107 genes *MADS-box* foram identificados nos últimos 20 anos, sendo que os genes do tipo *MIKC^C* compõem o modelo de desenvolvimento floral ABCDE (THEISSEN, 2001; THEISSEN & SAEDLER, 2001; PARENICOVA *et al.*, 2003; SMACZNIAK *et al.*, 2012). Os genes da classe A (*APETALA1* - *API*) determinam a identidade de sépalas, os genes da classe A juntamente com genes da classe B (*APETALA3* - *AP3* - e *PISTILATTA* - *PI*) determinam a identidade das pétalas, ao passo que os genes da classe B somados aos genes da classe C (*AG*) são responsáveis pela formação de estames. Os genes da classe C possuem papel na identidade de carpelos (LOHMANN & WEIGEL, 2002), enquanto os genes da classe D (*SEEDSTICK-SHATTERPROOF1-SHATTERPROOF2*) atuam no desenvolvimento dos óvulos (Figura 2; COLOMBO *et al.*, 1995). Os genes *MADS-box* das classes A, B, C e D realizam interações com os genes da classe E (*SEPALATTA1-SEP2-SEP3-SEP4*), formando o modelo de quartetos florais (THEISSEN & SAEDLER, 2001 DORNELAS & DORNELAS, 2005). Interessantemente, as proteínas *MADS-box* formam complexos de alta ordem, normalmente em junções tetraméricas formadas por dois dímeros, como demonstrado na Figura 2.

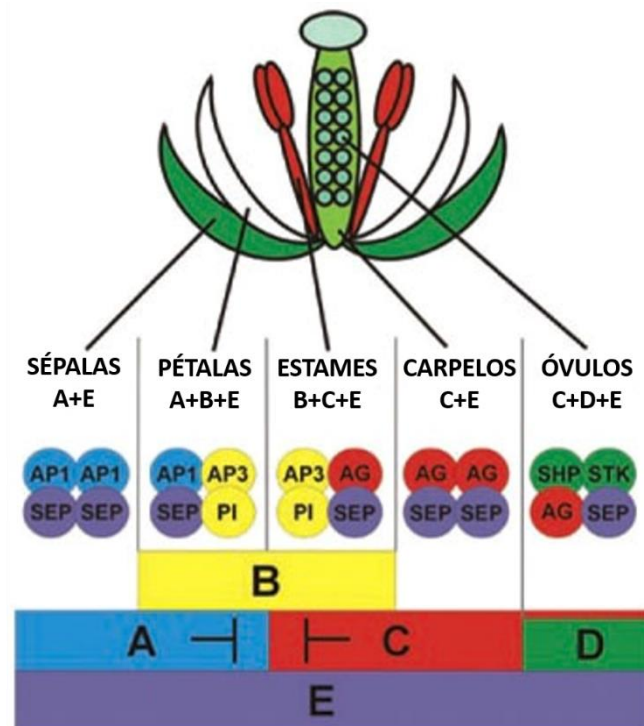


Figura 2. Modelo ABCDE de determinação da identidade dos órgãos florais por proteínas *MIKC^C* em *Arabidopsis thaliana*. Os genes *MIKC^C* da classe A (*API*) determinam a formação de sépalas; genes da classe B (*AP3* e *PI*), juntamente com genes da classe A, são necessários para a formação de pétalas. Os genes da classe B também são necessários para a formação de estames junto com os genes da classe C (*AG*), que por sua vez são necessários para a formação de carpelos. Os genes da classe D (*STK*, *SHP1* e *SHP2*) são responsáveis pelo desenvolvimento de óvulos. Todas as classes (ABCD) necessitam de, ao menos, um dos genes da classe E (*SEP1*, *SEP2*, *SEP3* e *SEP4*) para realizarem sua função. Adaptado de DORNELAS & DORNELAS (2005).

Os complexos quaternários das proteínas MADS-box normalmente ligam-se a duas sequências de DNA consenso denominadas CArG-box (CC(A/T)₆GG) separadas por até 300 pb (LIU *et al.*, 2008). Essa interação entre complexos MADS-box e CArG-box é responsável pela geração de um *loop* de DNA entre os dois sítios CArG-box (THEISSEN, 2001; THEISSEN & SAEDLER, 2001; MENDES *et al.*, 2013). A formação das diferentes interações proteicas determina a diferenciação e o crescimento dos órgãos florais criando, assim, inúmeras combinações de multimerização de proteínas que aumentam a diversificação de ligação dos complexos MADS-box ao DNA e, conseqüentemente, a

promoção da transcrição de genes por essas proteínas (MELZER & THEISSEN, 2009; SMACZNIAK *et al.*, 2012).

1.1.3.2 Genes *MIKC^C* responsáveis pelo desenvolvimento de óvulo e de semente

Os genes *MIKC^C* das classes C e D formam a subfamília monofilética AG, pertencendo a clados irmãos uma vez que foram gerados por um evento de duplicação ocorrido no início da evolução das angiospermas (KRAMER *et al.* 2004). As proteínas codificadas por genes destas duas classes possuem dois motivos na região C-terminal conhecidos como AGI e II, os quais atuam na formação de complexos quaternários das proteínas MADS-box (YUN *et al.*, 2004; SONG *et al.*, 2006).

Os genes *MIKC^C* da classe D primeiramente caracterizados foram *FBP7* e *FBP11* (*FLORAL BINDING PROTEIN 7 e 11*) de *Petunia hybrida*, expressos especificamente nos óvulos. A geração de duplos mutantes destes genes levou à formação alterada destas estruturas (ANGENENT *et al.*, 1995; COLOMBO *et al.*, 1995; ANGENENT & COLOMBO 1996; COLOMBO *et al.*, 1997). No estudo de COLOMBO *et al.* (1995), foi demonstrado que a expressão ectópica de *FBP7* ou de *FBP11* em *Petunia* induziu a formação de óvulos em pétalas e sépalas, mostrando o papel destes genes no desenvolvimento dos óvulos. Foram propostos ortólogos à *FBP7* e *FBP11* em outras espécies como *AGL11* (*STK*) em *A. thaliana* (ROUNSLEY *et al.*, 1995; PINYOPICH *et al.*, 2003), *ZAG2* e *ZMM1* em *Zea* (SCHMIDT *et al.*, 1993; THEISSEN *et al.*, 1995), *OsMADS13* em *Oryza* (LOPEZ-DEE *et al.*, 1999), *PhalAG2* em *Phalaenopsis* (SONG *et al.*, 2006), *AVAG2* em *Asparagus* (YUN *et al.*, 2004), *LMADS2* em *Lilium* e *EgMADS1* em *Lisianthus* (TZENG *et al.* 2002). Todos estes genes tendem a especificar a identidade dos tecidos da quarta camada da flor.

Em *A. thaliana*, o gene *AtAGL11* é responsável pela identidade do óvulo juntamente com os genes *SHATTERPROOF 1* e *SHATTERPROOF 2* (*SHP1*, *SHP2*), e seu mutante defectivo apresenta fenótipo de sementes diminutas (PINYOPICH *et al.*, 2003; BRAMBILLA *et al.*, 2007). O gene *AtAGL11* é responsável pela expansão e divisão celular do funículo, sendo que a ausência deste gene faz com que esta estrutura se torne mais larga e espessa (ROUNSLEY *et al.*, 1995; PINYOPICH *et al.*, 2003). Este gene é uniformemente

expresso nos integumentos dos óvulos e ao longo do funículo nas fases iniciais de desenvolvimento da semente. Na fase de maturação, *AtAGL11* é fortemente expresso no funículo e fracamente nos integumentos (ROUNSLEY *et al.*, 1995; PINYOPICH *et al.*, 2003; MATIAS-HERNANDEZ *et al.*, 2010). Seu mutante (*stk*), além de demonstrar diminuição no tamanho das sementes e aumento da espessura do funículo (Figura 3d-e), possui síliquas menores com apenas metade da quantidade normal de sementes (Figura 3a). O mutante *stk* também apresenta o fenótipo de não abscisão das sementes da rafe (Figura 3f), provavelmente pela estrutura diferenciada do funículo que as mantém fortemente ligadas à rafe (PINYOPICH *et al.*, 2003).

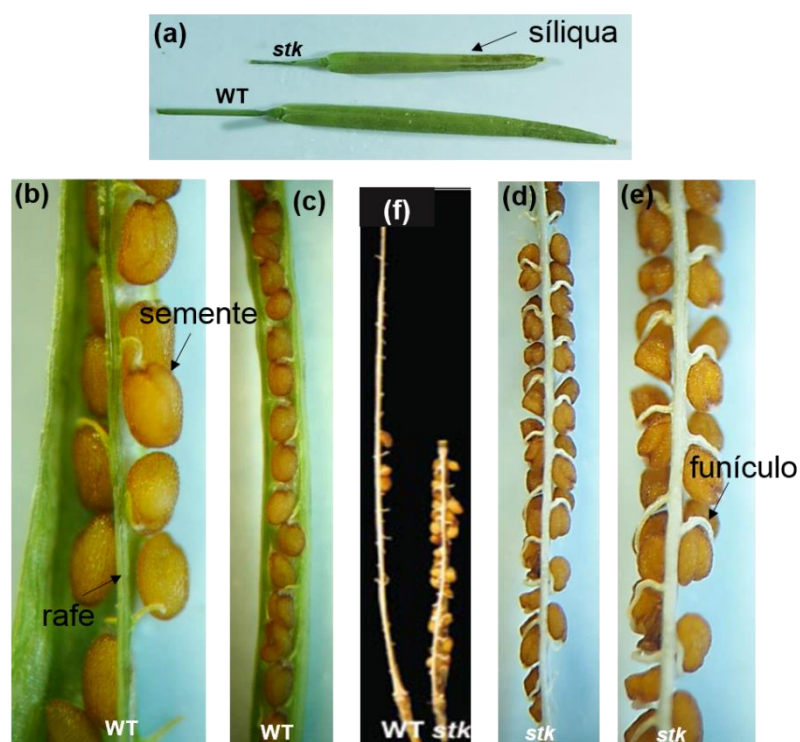


Figura 3. Comparação entre *Arabidopsis thaliana* Col-0 selvagem (WT, do inglês, *wild-type*) e *A. thaliana* Col-0 mutante para o gene *AtAGL11* (*stk*). (a) Diferença entre o tamanho de síliqua; (b-c) formato e tamanho da semente de WT e espessura e comprimento do funículo de WT; (d-e) formato e tamanho da semente de *stk* e espessura e comprimento do funículo de *stk*; (f) diferença entre WT e *stk* em relação à abscisão das sementes da rafe. Imagem f realizada por PINYOPICH *et al.* (2003).

A regulação da expressão tecido-específica do gene *AG* é dependente de elementos *cis* presentes no seu segundo íntron, conhecido por sua grande extensão (3 quilobases ou kb, do inglês, *kilobase*) (DEYHOLOS 2000; HONG *et al* 2003). Da mesma maneira, o gene *AtAG11* (*STK*) é regulado por elementos *cis* localizados em sua região promotora e em seu grande primeiro íntron (1,3 kb). Esses elementos regulatórios são ricos em G-A, no qual a proteína BASIC PENTACYSSTEINE 1 (BPC1) se liga e gera uma mudança conformacional do DNA que auxilia na formação do precomplexo de ativação da transcrição deste gene (KOOIKER *et al.*, 2005).

A compreensão da regulação da expressão de genes responsáveis pelo desenvolvimento de semente em plantas modelos como *A. thaliana* avançou significativamente nos últimos anos (MATIAS-HERNANDEZ *et al.*, 2010). Do mesmo modo, estudos abordando este tema também vêm sendo conduzidos em outras espécies tais como videira, tomateiro, laranjeira, entre outros (VAROQUAUX *et al.*, 2000). Uma das plantas que têm sido extensivamente estudadas com o intuito de buscar uma maior compreensão do processo de apirenia é a videira. Nesta planta, a subfamília AG foi previamente caracterizada, sendo composta por três membros, dois deles mais similar a AG (*VvAG1* e *VvAG2*) e um terceiro, *VvAGL11* ou *VvAG3*, mais similar de *AGL11/STK* (BOSS *et al.*, 2001 e 2002). Porém, nenhum gene foi identificado como ortólogo de *SHP1* e *SHP2* (DÍAZ RIQUELME *et al.*, 2009). A confirmação da ortologia entre *AtAGL11* e *VvAGL11* auxiliaria no processo de compressão da ausência de semente em *V. vinifera*.

1.2 A CULTURA DA VIDEIRA

A videira (*Vitis* spp. L.) é uma planta de origem antiga, proveniente de áreas caucasianas e que se propagou inicialmente na área mediterrânea. Sua grande expansão mundial ocorreu graças a sua habilidade de se adaptar a diversos solos e condições climáticas (LACIRIGNOLA & DIGIARO, 1999). Atualmente, a videira é a frutífera mais cultivada mundialmente, cobrindo 7,8 milhões de hectares em 2011 e sendo a 5ª fruta mais produzida no mundo com 67,5 milhões de toneladas de frutos em 2012 (OIV, 2011; FAO, 2012). A videira possui grande apelo comercial uma vez que as suas bagas são utilizadas para as mais diversas finalidades tais como produção de vinhos (68%), consumo de uvas de mesa *in natura* (30%), produção de uvas passas (2%) e produtos de menor impacto como sucos,

geleias, etanol, vinagre, óleo de sementes, ácido tartárico e fertilizantes (FASOLI *et al.*, 2012).

A viticultura é uma atividade tradicional em países de clima temperado e, recentemente, tem ganhado grande significância em diversas regiões de clima subtropical e tropical. Esta característica abre perspectivas para uma grande expansão na viticultura tropical, principalmente para a produção de uvas de mesa (AMARAL *et al.*, 2012). A produção mundial de uva de mesa atingiu 2,1 milhões de toneladas em 2010, sendo que o Brasil ocupa o 6º lugar na produção mundial de uva de mesa (FAO, 2010).

A apirenia é uma das características mais apreciadas em uvas de mesa, em concomitância com frutos de tamanho elevado. Estas duas qualidades constituem principais objetivos de programas de melhoramento de uvas de mesa (VAROQUAUX *et al.*, 2000; CABEZAS *et al.*, 2006). Nos Estados Unidos da América, as uvas apirênicas já dominam o mercado enquanto que, na Europa, é crescente a demanda por uvas sem sementes. Dados de 2012 da Organização Internacional da Uva e do Vinho (OIV, do francês, *Organisation Internationale de la Vigne et du Vin* e da Organização das Nações Unidas para Agricultura e Alimentação (FAO, do inglês, *Food and Agriculture Organization of the United Nations*), permitiram comprovar que nas últimas duas décadas a comercialização mundial de uva *in natura* aumentou cerca de 26% ao ano, enquanto que a produção de uvas para esta finalidade cresceu a taxas anuais próximas de 13%. Essa expansão comercial gerou oportunidades de crescimento aos países produtores e a competitividade entre os mercados exportadores das melhores uvas apirênicas (LAZZAROTTO & FIORAVANÇO, 2012).

A produção de uvas brasileiras *in natura* está baseada, predominantemente, na cultivar Niágara Rosada, no caso de uvas de mesa comuns. Em contrapartida, as uvas finas de mesa são, essencialmente, as cultivares Itália, Rubi, Benitaka, Brasil e Red Globe, que são uvas pirênicas, e as cultivares Centennial Seedless, Superior Seedless, Sultanina, Catalunha e Crimson Seedless, que são uvas apirênicas (PROTAS, 2005). Dentre estas, a cultivar Sultanina destaca-se devido à sua boa adaptação ao cenário europeu e estado-unidense de cultivo, onde foi amplamente utilizada em cruzamentos visando a geração de novas cultivares apirênicas e, por isso, vêm sendo estudada como modelo de apirenia em videira.

1.2.1 A Cultivar Sultanina

Atualmente, diversas variedades de uvas apirênicas são produzidas no mundo, cada qual sendo escolhida por sua adaptabilidade às condições locais. As principais cultivares estenoespermocárpicas são, em sua maioria, descendentes de ‘Sultanina’, também chamada de ‘Thompson Seedless’. Esta é uma das cultivares mais importantes comercialmente e é, até hoje, utilizada em cruzamentos como doadora de seu fenótipo apirênico (ADAM-BLONDON *et al.*, 2001; GENOVA *et al.*, 2014). Na Figura 4, é possível visualizar o contraste entre uma cultivar pirênica, neste caso ‘Chardonnay’, e ‘Sultanina’, que é estenoespermocárpica e, portanto, origina frutos com traços de sementes pouco desenvolvidos e macios, imperceptíveis ao consumidor, sendo fortemente apreciada no mercado de uvas de mesa (STOUT, 1936). A mutante somática natural Sultanina possui coloração mais clara e estava entre um grupo de variedades inicialmente cultivadas em Nova York e, em 1872, em Marysville, Califórnia, por Willian Thompson do qual descende seu nome mais comum. Originalmente, a cultivar Sultanina é proveniente da Pérsia da Ásia Menor, atualmente áreas do Irã e da Turquia, onde era conhecida como ‘Oval Kechmish’ (CHRISTENSEN, 2000).

Para a determinação do fenótipo de apirenia de uma determinada cultivar de videira é utilizada uma classificação determinada pela OIV sobre a norma descritora 243 (Figura 5). De acordo com esta norma utiliza-se dados de matéria seca de 100 sementes aleatoriamente amostradas, classificadas pela seguinte ordem: classe 1 - muito baixa (≤ 10 mg/semente); classe 3 - baixa (21-29 mg/semente); classe 5 - média (36-44 mg/semente); classe 7 - alta (51-59 mg/semente) e classe 9 - muito alta (> 65 mg/semente) como mostrado na Figura 6. As variedades que se encaixam nas classes 1, 3 e 5 são consideradas apirênicas, enquanto que as cultivares com notas 7 e 9 são consideradas pirênicas.

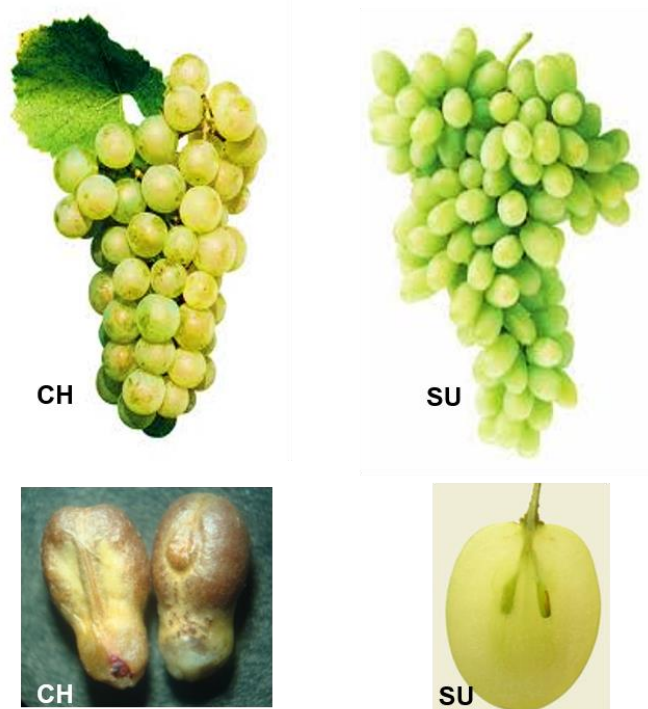


Figura 4. Comparação entre a cultivar pirênica Chardonnay (CH) e a cultivar apirênica Sultanina (SU), ambas em estágio de maturação dos frutos. ‘Chardonnay’ apresenta sementes de tamanho adequado e lignificadas enquanto que ‘Sultanina’ (baga) contém apenas o traço de uma semente. Imagem de ‘Chardonnay’ proveniente de Ternuta dei Mille (2011), e de ‘Sultanina’ proveniente de Agrofresh (2013). Semente de ‘Chardonnay’ com aumento de 40X.

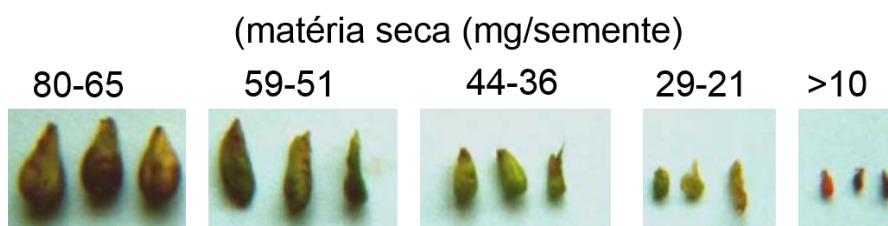


Figura 5. Classificação da pirenia e apirenia em videira segundo o descritor OIV 243. Distribuição das classes fenotípicas para a determinação de apirenia. Peso de matéria seca entre 44 - >10 mg/semente, abrangendo as classes 1, 3 e 5, classifica a cultivar como apirênica. Peso de matéria entre 51 - < 65, classes 7 e 9, determina que a cultivar é pirênica. Imagem proveniente de Acervo EMBRAPA Uva e Vinho. Adaptado de BOUQUET & DANGLLOT (1996).

1.2.2 Controle Genético e Molecular da Apirenia em Videira

V. vinifera possui seu genoma completamente sequenciado em sua forma quase homozigota e, também, em sua forma altamente heterozigota, ambas obtidas a partir da cultivar Pinot Noir (PN40024; JAILLON *et al.*, 2007; VELASCO *et al.*, 2007), caracterizando o genoma-referência de videira ([http:// http://genomes.cribi.unipd.it/grape/](http://genomes.cribi.unipd.it/grape/)). Este conjunto de dados genômicos auxilia em estudos de desenvolvimento de cultivares por métodos não tradicionais, concomitantemente com o entendimento da função de seus mais de 23 mil genes.

A identificação e a compreensão dos mecanismos genéticos e moleculares que regem o desenvolvimento de espécies com ausência de sementes são essenciais para a obtenção de culturas comerciais de alta qualidade e competitividade no mercado (CAMARGO *et al.*, 1999). Como alternativa biotecnológica, muitos pesquisadores focaram seus esforços na busca de marcadores moleculares que pudessem ser utilizados para a detecção do fenótipo de apirenia prematuramente, utilizando-os em estratégias de seleção assistida por marcadores (MAS, do inglês, *Marker Assisted Selection*) em populações segregantes (STRIEM *et al.*, 1996; LAHOGUE *et al.*, 1998; ADAM-BLONDON *et al.*, 2001; MEJÍA *et al.*, 2007; MEJÍA *et al.*, 2011).

O método de obtenção de mapas genéticos baseados na estratégia do duplo pseudocruzamento-teste, desenvolvido por GRATTAPAGLIA & SEDEROFF (1994), permitiu identificar QTLs com grandes efeitos fenotípicos. Esta estratégia é muito utilizada para espécies altamente heterozigotas e de demorado desenvolvimento como *V. vinifera*, sendo que os indivíduos avaliados são a progênie do cruzamento, plantas F1. Deste modo, a produção de mapas genéticos de videira aumentou consideravelmente (LODHI *et al.*, 1995; DALBO *et al.*, 2000; DOLIGEZ *et al.*, 2002; RIAZ *et al.*, 2003; ADAM-BLONDON *et al.*, 2004; FISCHER *et al.*, 2004; CABEZAS *et al.*, 2006).

Diversos graus de tamanho de sementes podem ser observados em cultivares de videiras estenospermocárpicas e em suas progênies, o que demonstra que a estenospermocarpia é um caractere quantitativo. BOUQUET & DANGLOT (1996) geraram um modelo de controle desta característica. Neste modelo, alelos recessivos de três *loci* independentes são regulados por um alelo dominante em um único *locus*, o qual inibiria

o desenvolvimento da semente. Posteriormente, este *locus* foi denominado *SdI* (Inibidor do desenvolvimento da semente, do inglês, *Seed development Inhibitor*) (LAHOGUE *et al.*, 1998).

Em estudos genéticos com populações geradas a partir de variedades de videira pirênicas (doadora materna) e apirênicas (doador paterno), foram detectados QTLs para tamanho de baga, peso e tamanho de sementes em *V. vinifera*. Os QTLs estão localizados no grupo de ligação 18, na porção distal do cromossomo, e explica de 50 a 70% da variância fenotípica para esses caracteres (DOLIGEZ *et al.*, 2002; FANIZZA *et al.*, 2005; CABEZAS *et al.*, 2006; MEJÍA *et al.*, 2007; CONSTANTINI *et al.*, 2008; REVERS *et al.*, 2010). Este QTL corresponde ao *locus SdI* proposto anteriormente no modelo de BOUQUET & DANGLLOT (1996) (MEJÍA *et al.*, 2011).

A partir deste *QTL*, o gene *AGAMOUS-LIKE 11* (*VvAGL11*) foi mapeado *in silico* no mesmo *contig* que o *locus SdI* e também colocalizou com o marcador molecular SSR (repetição de sequência simples, do inglês, *Simple Sequence Repeat*) VMC7F2, que se encontra 463 pb a montante do códon de início de tradução de *VvAGL11*. Este marcador é altamente polimórfico, identificando diferenças entre os alelos do gene *VvAGL11* por amplificar uma região que apresenta tamanhos desiguais. Assim, é possível detectar cada um dos dois alelos de ‘Sultanina’, no caso um dos alelos é idêntico ao do genoma referência (‘Pinot Noir’) e é chamado de alelo selvagem, enquanto que o outro alelo, chamado de alelo mutado, possui diferenças em sua região promotora. VMC7F2 é utilizado com 95% de eficácia para detecção de apirenia em videira (CONSTANTINI *et al.*, 2008). Pelo emprego deste marcador na técnica de MAS é possível identificar o genótipo de cada indivíduo, podendo este ser homocigoto para o alelo normal (pirênico), heterocigoto (apirênico) ou homocigoto para o alelo mutado (apirênico).

VvAGL11 (acesso no genoma em <http://genomes.cribi.unipd.it/grape/Vv18s0041g01880>), também chamado por BOSS *et al.* (2002) de *VvMADS5* e por DÍAZ-RIQUELME *et al.* (2009) de *VvAG3*, é um possível ortólogo do gene *AtAGL11/STK* de *A. thaliana* que, por sua vez, atua no controle da identidade do óvulo (FAVARO *et al.*, 2003; PINYOPICH *et al.*, 2003). Em um estudo contemplando a família MADS-box em videira, DÍAZ-RIQUELME *et al.* (2009) demonstrou que *VvAGL11* é induzido em tecidos florais e em frutos, enquanto que é reprimido em raiz, ramo, folha, gavinha e gema. Ainda nesse estudo, foi demonstrada uma possível ortologia entre *VvAGL11* (*VvAG3* na Figura 6),

AtAGL11 e *PMADS21* de álamo. Estes genes agruparam-se na subfamília AG (Figura 6), na qual os genes *MADS-box MIKC* de videira, álamo e *A. thaliana* foram agrupados em uma árvore filogenética que se mostrou dividida em 13 subfamílias (SEPALLATA, AGAMOUS-LIKE 6, APETALA1-FRUITFULL, FLOWERING LOCUS C, AGAMOUS, AGAMOUS-LIKE 12, TM 8, AGAMOUS-LIKE 15, AGAMOUS-LIKE 17, SUPPRESSOR OF OVEREXPRESSION OF CONSTANS 1, SHORT VEGETATIVE PHASE, APETALA-PISTILLATA, B SISTER) dentro de 10 clados maiores.

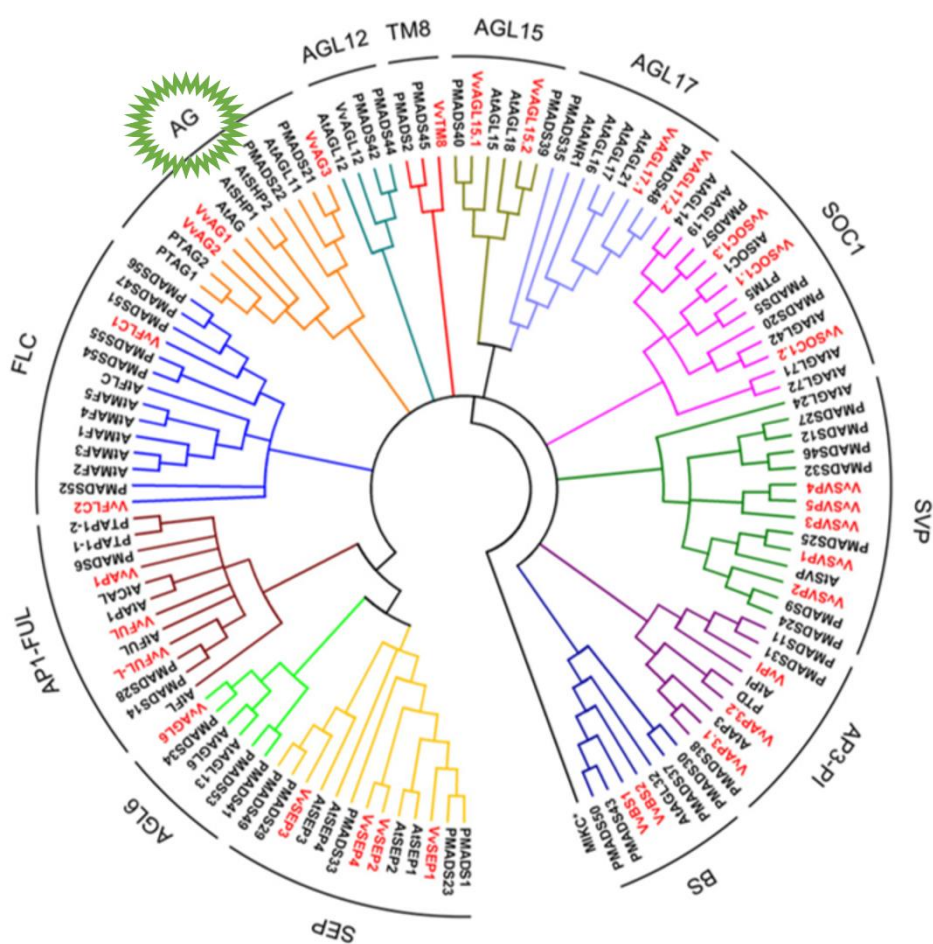


Figura 6. Árvore filogenética da família de genes *MIKC* de *Vitis* (em vermelho), *Arabidopsis* e *Populus*. Árvore gerada pelo alinhamento das sequências completas de aminoácidos das proteínas de cada gene. O método junção de vizinhos (NJ, do inglês, *neighbor-joining*) foi utilizado para o alinhamento. As proteínas *MIKC* agruparam-se em 13 subfamílias. As proteínas *MADS-box MIKC** foram utilizadas como um grupo externo. Adaptado de DÍAZ-RIQUELME *et al.* (2009).

Com o objetivo de caracterizar a expressão do gene *VvAGL11*, MEJÍA *et al.* (2011), por meio de ensaios de PCR quantitativa precedida de transcrição reversa (*RT-qPCR*, do inglês, *reverse transcription-quantitative PCR*), observaram que transcritos de *VvAGL11* são 25 vezes mais abundantes no estágio de fruto em comparação com os estádios de prefloração e floração em genótipos de videira sem sementes. Outro dado interessante refere-se ao fato de que, no estágio de fruto, a expressão do gene é acentuada em genótipos homocigotos para o alelo normal de *VvAGL11*, mostrando uma expressão 25 vezes maior do que em genótipos homocigotos para o alelo mutado, enquanto que os genótipos heterocigotos demonstraram um nível intermediário da expressão do gene. O sequenciamento da região promotora e codificadora de *VvAGL11* realizado por estes autores não permitiu concluir se as variações alélicas identificadas são responsáveis pelo fenótipo de apirenia e não foram realizadas análises funcionais neste trabalho (MEJÍA *et al.*, 2011).

Paralelamente a estes estudos, nosso grupo analisou uma progênie de videira gerada a partir do cruzamento de uma cultivar apirênica ‘Crimson Seedless’ com o híbrido complexo resistente ao míldio ‘Villard Blanc’. Os resultados obtidos a partir do mapa genético construído entre estas duas cultivares permitiram confirmar o QTL para a ausência de sementes dentro do *locus Sd1* na extremidade do cromossomo 18 e com LOD (do inglês, *Linked Open Data*) máximo associado ao marcador molecular microssatélite VMC7F2 (REVERS *et al.*, 2010).

Na presente Dissertação de Mestrado, a hipótese de trabalho foi fundamentada na seguinte afirmação: a extrema diminuição da expressão de *VvAGL11* encontrada em genótipos apirênicos de videira é a causa do fenótipo de apirenia, uma vez que o gene é essencial para a morfogênese da semente em uvas.

2 OBJETIVOS

2.1 OBJETIVO GERAL

Caracterizar as estruturas do gene *VvAGL11* na cultivar pirênica Chardonnay e na cultivar apirênica Sultanina por meio do sequenciamento de suas variações alélicas e da exploração da sua expressão gênica diferencial, visando uma maior compreensão de seu papel durante a morfogênese das sementes em videira.

2.2 OBJETIVOS ESPECÍFICOS

- Caracterizar as variações alélicas do gene *VvAGL11* em videira apirênica e pirênica;
- Caracterizar o perfil transcricional do gene *VvAGL11* em diferentes estádios de desenvolvimento do fruto;
- Analisar morfológicamente a semente de ‘Chardonnay’ e o traço de semente de ‘Sultanina’;
- Determinar o padrão de expressão espaço-temporal de *VvAGL11* em videira apirênica e com semente durante o desenvolvimento reprodutivo.

3 CAPÍTULO I

THE *AGAMOUS-LIKE 11* TRANSCRIPTION IS ESSENTIAL FOR SEED MORPHOGENESIS IN GRAPEVINE

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Title

The *AGAMOUS-LIKE 11* transcription is essential for seed morphogenesis in grapevine

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Abstract

Despite the wide and extensive appreciation of seedless grapes, little is known about the mechanisms that drive apireny. *VvAGL11* is a candidate gene involved in *Vitis vinifera* seed morphogenesis and studies showed that its expression is mainly localized in fruits of seeded grapes, while seedless varieties exhibit significantly lower levels of transcripts. Grapevine *AGL11* is a MADS-box gene, possibly orthologous to *Arabidopsis AGL11*, with an important role in ovule development. Our study aimed to characterize *VvAGL11* allelic variations in a seeded (Chardonnay) and in a seedless (Sultanina) grapevine cultivars, allowing us to observe several mutations that might be responsible for gene's loss of function. *VvAGL11* transcript levels corresponding to each phenotype were evaluated by RT-qPCR during grapevine flower and fruit development. *VvAGL11* relative expression was significantly higher in seeds of 2, 4 and 6 weeks after fruit-set of the seeded variety whereas in the seedless grape its transcription was extremely low in all stages analyzed. We further characterized the patterns of *VvAGL11* spatial and temporal expression via *in situ* hybridization. Transcript accumulation was specifically detected in the dual medium integument layer of the seeds, which is responsible for the elongation of seed coat and necessary to determine final seed size. Nevertheless, in seedless grape, no hybridization signals were visible and a morphological analysis showed an apparent loss of identity of the medium integument layer of the seed traces. Our data provide insights of the important role of *VvAGL11* in seed morphogenesis suggesting that depletion of its expression may be the main responsible for the erroneous development of a highly essential seed layer therefore culminating in the typical apirenic phenotype.

Keywords: apireny, grapevine, *in situ* hybridization, seed, Sultanina, *VvAGL11*.

Introduction

Seedlessness is one of the most appreciated features in table grapes along with enlarged fruit size, both being main goals in breeding programs (Varoquaux *et al.*, 2000; Cabezas *et al.*, 2006; Lazzarotto and Fioravanço, 2012). In the past two decades, the worldwide market of *in natura* grapes increased about 26% per year, while the production of such grapes grew at a 13% annual rate (OIV, 2012; FAO, 2012). In grapevine, the absence of seeds, also called apireny, is divided in two biological processes. One is known as parthenocarpy, when fecundation does not occur and there is no seed formation, as in raisins. The second process is called stenospermocarpy, in which fertilization takes place to form the fruit but it is followed by embryo abortion due to the cease of endosperm development, normally after four weeks of fruit growth. The consequence of stenospermocarpy is the production of berries with reduced seed size, known as seed traces (Bouquet and Danglot, 1996; Mejía, 2011).

Several studies have shown that the presence of a dominant allele in the region of the *Seed Development Inhibitor (SdI)* locus, located in *Vitis vinifera* distal portion of chromosome 18, is responsible for 50 to 90% of the total phenotypic variance of seedlessness (Bouquet and Danglot, 1996; Lahogue *et al.*, 1998; Cabezas *et al.*, 2006; Costantini *et al.*, 2008; Doligez *et al.*, 2002; Mejía *et al.*, 2007). *In silico* analysis allowed the identification of a gene, named *Vitis vinifera AGAMOUS-LIKE 11 (VvAGL11)*, that mapped to the same locus contig *SdI* (Constantini *et al.*, 2008). The same work has also co-located *VvAGL11* with the molecular microsatellite marker VMC7F2, which was known to be a highly polymorphic marker capable of an efficiency of 95% in the detection of seedlessness in grapes (Constantini *et al.*, 2008).

The *VvAGL11* gene (Vv18s0041g01880; Boss *et al.*, 2002) was proposed as a strong candidate gene involved in the absence of seeds in grapevine (Costantini *et al.*, 2008; Mejía *et al.*, 2011; Revers *et al.*, 2010). This MADS-box gene was shown to be a possible ortholog of *AGL11 (STK)* of *Arabidopsis thaliana*, which acts on the identity and control of ovule and seed development (Favaro *et al.*, 2003; Pinyopich *et al.*, 2003; Díaz-Riquelme *et al.*, 2009). *AtAGL11* mutant (*stk*) presents reduced number and size of seeds (Pinyopich *et al.*, 2003; Brambilla *et al.*, 2007).

The transcription factors of the MADS-box family have extremely important roles in the development of plants. They are responsible for controlling the identity of floral organs (Smaczniak *et al.*, 2012). In a study covering the MADS-box family in grapevine, Díaz-Riquelme *et al.* (2009) demonstrated that *VvAGL11* is induced in floral and fruit tissues, whereas it is repressed in roots, branches, leaves, buds and tendrils. More recently, another study showed that *VvAGL11* is 25 times more expressed at the stage of fruit compared to flower stages in genotypes homozygous for seedless in grapes. In heterozygous genotypes, an intermediate level of *VvAGL11* expression was observed (Mejía *et al.*, 2011).

We therefore hypothesized that *VvAGL11* might be responsible for controlling seed morphogenesis in grapevine since the decrease of its expression would cause the apirenic phenotype. In this work we report the identification of differences between *VvAGL11* allele sequences in apirenic and pirenic grapevines, observing several mutations in intragenic regions and a set of *SNPs* in the coding region of the gene exclusively in the apirenic genotypic background. We characterized the *VvAGL11* transcript levels, observing a 65-fold increase in expression in grapevine seeds compared to apirenic grapevine fruits. We also described its spatial and temporal expression patterns in reproductive tissues by *in situ* hybridization that allowed us to establish a correlation between the development of a specific seed layer with normal or decreased *VvAGL11* expression.

Materials and methods

Plant material

All samples were harvested from grapevines located at Embrapa Grape and Wine experimental areas in Bento Gonçalves, Rio Grande do Sul, Brazil (29° 09' 48" S, 51° 31' 42" O and 616 m altitude). The phenological scale followed was the one described by Baggiolini (1952; Supplementary Table 2). 'Chardonnay' was chosen for comparison with 'Sultanina' because of its similar phenological development. For *VvAGL11* transcriptional profile, 'Chardonnay' (seeded) and 'Sultanina' (seedless) organ and tissue samples were harvested at the stages of preanthesis flower, fruit set and fruits at 2, 4 and 6 weeks after fruit set in 2010/2011. For *in situ* hybridization and morphological assays, 'Chardonnay' and 'Sultanina' samples were harvested at H1, preanthesis flower, fruit set and fruits at 2, 4 and

6 weeks after fruit set in 2012/2013 harvest. Both cultivars were grafted on Paulsen 1103 rootstock.

Sequencing

Twenty-two sets of primers were designed to amplify sequences in the promoter and in exon/intron regions of *VvAGL11* (Supplementary Table 1) in order to sequence the whole gene. Each region exhibited a 300-500 bp overlapping sequence. PCR amplifications were performed with 10 to 25 ng of grapevine genomic DNA using Platinum[®] Pfx DNA polymerase (Applied Biosystems) as recommended by the manufacturer. RACE amplifications were done with the SMARTer[™] RACE cDNA Amplification Kit (Clontech). PCR products were cloned into pGEM[®]-T Easy Vector (Promega) following the manufacturer instructions. Plasmid DNA was extracted by the alkaline lysis miniprep protocol modified by a CIA step (Maniatis, Fritsch and Sambrook, 1987). PCR products were treated with the Illustra ExoProStar reagent (GE Healthcare) as recommended by the manufacturer, and were sequenced in an ABI Prism[®] 310 Genetic Analyser (Applied Biosystems) using standard sequencing protocols described by Falavigna *et al.*, 2014.

Identification of Putative CArG Sequences

Genomic regions located 3 kb upstream of the *VvAGL11* ATG start codon, 377 bp downstream of the stop codon, and exons and introns were analyzed to identify CArG-Box sequences with the New Place bioinformatic program available at the Sogo website (<http://sogo.dna.affrc.go.jp>).

RNA extraction

Total RNA samples were extracted from frozen material (approximately 200 mg) by LiCl precipitation using the Zeng and Yang (2002) protocol modified by a purification scale adapted to 2 mL microcentrifuge tubes. Each extraction was conducted using three tubes that had their volumes pooled before the LiCl precipitation step. Genomic DNA in total RNA samples was removed using the TURBO DNA-free Kit (Ambion) according to the manufacturer's protocol. RNA integrity and quantity was monitored by agarose gel electrophoresis and spectrophotometric quantitation, respectively.

RT-qPCR analyses

Complementary DNAs were synthesized using the GeneAmp RNA PCR Core Kit (Applied Biosystems) according to manufacturer's instructions. Gene-specific primer pairs were designed for the two forms of the candidate gene (*VvAGL1IF* 5'-CACTTAATGGGTGAT TCCTTGGC-3', *VvAGL1IR* 5'-AGCAACTCATGCTTCTTCGACC-3'; and *VvAGL3F* 5'-ATTGTTCATCTGGGCATTTTCG-3', *VvAGL3R* 5'-GGAGATGAAGTTGGCGGATA-3') and evaluated by Oligo Analyzer (IDT, <http://www.idtdna.com>). RT-qPCR was performed in a StepOnePlus Real-Time PCR System (Applied Biosystems). SYBR Green (Invitrogen) was used to monitor dsDNA synthesis and ROX (Invitrogen) was employed as passive fluorescence reference. Each biological sample was analyzed in technical triplicates. Cycling protocol consisted of one step at 95 °C for 10 min followed by 40 cycles of 95 °C for 15 s, 60 °C for 1 min, and finished by a dissociation curve between 60 °C and 95 °C. The specificity of PCR amplifications was assessed by the presence of a single peak in melting curves and by the visualization of single amplification products of expected size in 1% ethidium bromide gel electrophoresis. Primer efficiency was calculated by LinRegPCR (version 11.0, Ruijter *et al.*, 2009). Mean relative gene expression was calculated by the Pfaffl (2001) method with the *actin* gene (GenBank EC969944) as reference (Reid *et al.*, 2006). Statistical analysis was performed using Prism 5.1 with One-way ANOVA and Tukey test.

Seed morphological assay

Seeds and seed traces were transferred into a fixation solution under vacuum (McDowell and Trump, 1976). Thereafter, samples were dehydrated in an increasing gradient of ethanol as described by Gabriel (1982), and embedded in 2-hydroxyethyl methacrylate resin according to Gerrits and Smid (1983). The 5 µm sections were obtained by a Leica RM 2255 microtome. The metachromatic reagent Toluidine Blue O (Feder and O'Brien, 1968) was used to determine seed structure. For the staining procedure, slides with sections were placed on a hot plate at 45 °C, and the reagent was added. After 15 min, slides were rinsed with 95% ethanol and dried at room temperature.

In situ hybridization analysis

‘Chardonnay’ and ‘Sultanina’ samples were fixed in 4% formaldehyde for 16 hours. To submerge the samples, vacuum was applied (approximately 600 mmHg) for 20 min. The material was stored under refrigeration (4 to 10 °C) after fixation. Thereafter, the tissues were dehydrated in ethanol series and embedded in paraffin. Longitudinal and transversal sections (8-10 µm) were prepared with a rotary microtome and mounted on silanized microscope slides. Gene-specific sense and antisense probes were designed at 3’UTR region (*VvAGL3F* 5’-ATTGTTTCATCTGGGCATTTTCG-3’, *VvAGL3R* 5’-GGAGATGAAGTTGGCGGATA-3’) corresponding to 185 nucleotides of the 3’ UTR of *VvAGL11* gene. Probes were generated by digoxigenin (DIG)-labeling using T7 or SP6 RNA polymerase of the DIG RNA Labeling Kit (Roche). After the detection of the hybridization signals by immunostaining, slides were washed, dehydrated, and mounted using Entellan® (Merck).

Results

Sequencing of VvAGL11

To further characterize the structural organization of the *VvAGL11* gene, allelic variations were PCR amplified with specific primers from genomic DNA extracted from an apirenic grapevine, ‘Sultanina’, and a pirenic grapevine, ‘Chardonnay’ as previously described. A total of six PCR fragments were assembled, formed by overlapping primers sets showed in figure 1. The gene sequence is composed by the promoter, coding and intragenic regions (8 exons and 7 introns). After allele specific isolation and sequencing, we were allowed to identify two alleles for each cultivar. Because ‘Chardonnay’ is a direct offspring of Pinot Noir, the comparison with the ‘Pinot Noir’ (PN40024) sequence in the publicly available genome base was possible (http://genomes.cribi.unipd.it/gb2/gbrowse/public/Vitis_vinifera/).

The two *VvAGL11* alleles of ‘Chardonnay’ showed 99% sequence identity with the corresponding ‘Pinot Noir’ genome sequence for *VvAGL11*. One allele from ‘Sultanina’ also exhibited 99% identity with PN40024. However the other ‘Sultanina’ *VvAGL11* allele, the one with the molecular microsatellite marker VMC7F2 described with shorter length (198 bp; Constantini *et al.*, 2008), presented a group of polymorphisms that differed this allele

from the others, and due to that it was named the mutant allele (*Sultanina mut*), as shown in Fig. 1. The group of polymorphisms was composed by 28 INDELs and 105 SNPs.

Among the observed alterations in *Sultanina mutant VvAGL11 allele*, two SNPs give rise to two amino acid substitutions (R590L and T628A) as shown in Fig. 2. A sequence alignment of the four *VvAGL11* alleles, PN40024, and the newest sequence of *VvAGL11* from *Sultanina* (Di Genova *et al.*, 2014) is presented in Supplementary Fig. 1. With high fidelity amplification and sequencing, we were confident to define all *VvAGL11* alleles' modifications. We also searched for CArG-box sequences, perfect CArG-boxes and CArG sequences with one mismatch deduced by a probability matrix. These *cis* elements are known to be the sites of MADS-box protein complex binding (Smaczniak *et al.*, 2012). We identified ten putative CArG sequences in *VvAGL11*. One of them is located in the promoter region, eight in the second intron and one in the 3'-UTR region. The seventh CArG-box putative sequence of '*Sultanina*' *VvAGL11 mut* has a nucleotide modification (Supplementary Fig. 1, black arrow). Sequence modification in these *cis* elements could disrupt *VvAGL11* transcription activation.

VvAGL11 expression isoforms

The 3'- and 5'-RACE experiments allowed us to visualize the mRNA isoforms of *VvAGL11* in apirenic and pirenice grapevines. Two products of amplification differing in size were obtained from '*Chardonnay*' and one product was obtained from '*Sultanina*'. After sequencing them, it was possible to confirm the specificity of the amplicons and to characterize each transcript form present in the cultivars.

The results revealed that '*Chardonnay*' has two isoforms of *VvAGL11* mRNA. One isoform, named CH-1, is longer with 1,454 bp, being 732 bp of coding region. This alternative form increases the coding region in 60 nucleotides. The second '*Chardonnay*' allele, named CH-2, has 958 bp and a coding sequence of 672 pb. '*Sultanina*' also has two transcript forms, both with the same length (958 bp) and coding sequence (672 bp). The dissemblance of the two *VvAGL11* isoforms derived from '*Sultanina*' is the ten SNPs found in the mutant allele (*SU mut*). The coding regions of CH-2 and '*Sultanina*' allele 1 are identical to the *VvAGL11* gene predicted in the Pinot Noir reference genome. However, none of the *Sultanina* isoforms are completely identical to the '*Pinot Noir*' gene since their 5'-UTR are different in size, as shown in Fig. 3.

Transcription profile of VvAGL11

Aiming to evaluate the expression of *VvAGL11* and its isoforms, we performed RT-qPCR analysis with mRNA samples extracted from the reproductive organs of ‘Chardonnay’ and ‘Sultanina’ at critical developing stages. Besides a pair of primers designed for regions of the actin reference gene, we employed two sets of primers in RT-qPCR studies to differentiate *VvAGL11* mRNA isoforms. The first *VvAGL11* primer set was composed of a forward primer designed to anneal in the last intronic region and a reverse primer that annealed in the 3'-UTR, therefore allowing the amplification of the longer CDS isoform (CH1). The second pair of primers was designed to anneal in the first (forward) and in the second (reverse) exons that allowed us to amplify simultaneously the two isoforms of *VvAGL11* (the shorter (CH1) and longer (CH2) CDS). Considering that in ‘Sultanina’ the isoforms differ in point mutations (10 SNPs) only, we employed the primer set flanking the first and second exons for the analysis of both isoforms (SU1 and SU *mut*).

The reproductive tissues evaluated were divided into initial and advanced developmental stages. The initial stages of development were represented by flowers at preanthesis and fruit-set. The advanced stages were represented by fruits at 2, 4 and 6 weeks of development after fruit establishment (fruit-set). Seeds were separated from the pulp in ‘Chardonnay’ berry samples and the whole berry was evaluated in ‘Sultanina’ samples. The amplicons were sequenced for determination of their specificity. Actin gene *VvACT* (Gen Bank EC969944) was used as reference.

Our RT-qPCR results showed that *VvAGL11* relative expression was similarly low at all stages sampled from the ‘Sultanina’ variety for both isoforms SU1 and SU *mut*. ‘Chardonnay’ flowers and berry pulps also demonstrated low transcript levels of *VvAGL11* for both isoforms CH1 and CH2. RT-qPCR with primers designed to the intronic region was also performed with ‘Sultanina’ mRNA samples and a minor level of expression was detected, probable descendent from residual from pre-mRNA during cDNA synthesis. Nevertheless, seeds of ‘Chardonnay’ from fruits at 2, 4 and 6 weeks after fruit set exhibited much higher accumulation of *VvAGL11* CH2 transcripts, an increase 15-fold higher than CH2 transcripts in berries at fruit set and 65-fold higher than ‘Sultanina’ whole berries. ‘Chardonnay’ CH1 allele relative expression was unstable through seed stages, fluctuating from 5 to 11 fold higher than fruit-set (Fig. 4).

Morphological analysis of seeds from apirenic and pirenice grapevines

V. vinifera seeds possess a particular shape resembling a pear, being triangular in cross sections (Fig. 5). The seeds are composed of a cuticle, an epidermis, two layers corresponding to the outer integument and the medium integument, both forming the seed coat at the ripening stage, and yet three inner integument layers that are positioned around the endosperm and embryo (Ribereau-Gayon and Peynaud, 1980). The structure of the seeds from ‘Chardonnay’ and seed traces from ‘Sultanina’ were found to be very different in shape, size and hardness (Fig. 5). For that reason, a morphological analysis was conducted to better characterize the development of endosperm, embryo and seed layers of both cultivars at 2, 4 and 6 weeks of fruit development. Descriptions of the structures as they evolve, examined with the metachromatic reagent toluidine blue O staining, are presented below and in Fig. 6.

Endosperm and Embryo. A normal and complete development of the endosperm and the embryo was observed in ‘Chardonnay’ seeds after 6 weeks of fruit development. ‘Sultanina’ seed traces showed an unevolved endosperm and no embryo could be distinguished at that stage.

Outer Integument. Parenchyma cells are the main constituent of the outer integument and cell layers are thicker in the ventral face of the seed. At the time of ripening there is an increase in parenchyma cell volume in ‘Chardonnay’, but the number of cells do not seem to change (Fig. 6A, 6C and 6E). In ‘Sultanina’ seed traces, no differences were observed in this layer, since there is no visible evolution of the seeds in this variety (Fig. 6B, 6D and 6F). Along with the outer integument, the epidermis covers the outer integument in both varieties.

Medium Integument. After 2 weeks of fruit development, one layer of rectangular palisade cells was observed in the seeds of both varieties (Fig. 6A and 6B). After 4 weeks, two layers of this integument were observed in ‘Chardonnay’ seeds (Fig. 6C), but this duplication was not observed in ‘Sultanina’ seed traces (Fig. 6D). After 6 weeks of fruit development, ‘Chardonnay’ seeds showed a sclerified and thicker medium integument (Fig. 6E) opposed to ‘Sultanina’ seed traces that exhibited the same structures of the initial stage of development.

Inner Integument. Three layers of inner integument were observed in the seeds at the initial stage of fruit development in both varieties (Fig. 6A and 6B), although only two layers

were present in later stages in ‘Chardonnay’ (Fig. 6C and 6E) and in ‘Sultanina’ (Fig. 6D and 6F).

Spatial and temporal accumulation of VvAGL11 transcripts

In situ hybridization (ISH) of the reproductive tissues of both grapevine varieties were performed in order to better characterize *VvAGL11* spatial and temporal expression patterns at the level of transcript accumulation. The stages of development analyzed were those described by Baggiolini (1952; Supplementary Table 2) including flower formation (H1), preanthesis (PA), fruit-set (FS), seeds at 2, 4 and 6 weeks after flower fertilization.

After ISH with the complementary (antisense) probe, no expression of *VvAGL11* was detected at the stages H1 (Fig. 7B), PA (Fig. 7C), FS (Fig. 7D) and at 6 weeks of fruit development (Fig. 7L) in ‘Chardonnay’. *VvAGL11* transcripts exhibited a high accumulation in seeds after 2 (Fig. 7E and 7F) and 4 weeks (Fig. 7G to 7J) of development in ‘Chardonnay’. The ISH signal was clearly visible in the medium integument layer of the seeds (Fig. 7K). In ‘Sultanina’, no gene expression was observed at any reproductive tissue/organ or stage of development (Fig. 8A to 8F). Important to mention, the ISH slides with the antisense probe were always compared with slides hybridized with the sense *VvAGL11* probe as control in the analysis of ‘Chardonnay’ and ‘Sultanina’ (Supplementary Fig. 2 and 3, respectively).

Discussion

Spontaneous somatic variants have been selected all the way through grapevine domestication, hence creating new and different cultivars with interesting traits such as berry size, taste, color, and seedlessness (This *et al.*, 2006). Grapevine somatic variants have also been useful tools for studies of gene function, whereas they result from single mutation events in a given genetic background (Fernandez *et al.*, 2010). *VvAGL11* was proposed as a candidate gene involved in the absence of seeds (Costantini *et al.*, 2008; Revers *et al.*, 2010; Mejía *et al.*, 2011). With the attempt to better evidence the essentiality of *VvAGL11* in grapevine apireny, we described here *VvAGL11* gene structure in apirenic (‘Sultanina’) and pirenice (‘Chardonnay’) grapevines, its transcript isoforms and profiles of accumulation, and

its spatial-temporal expression pattern during reproductive organ/tissue development and seed morphogenesis.

By means of a PCR strategy based on a high fidelity polymerase, sequencing of *VvAGL11* allelic variants from ‘Sultanina’ and ‘Chardonnay’ was performed, allowing the identification of a group of polymorphisms (Fig. 1). No transposon or large sequence insertion or deletion was found in ‘Sultanina’ that could explain the gene loss of transcription observed in previous data (Mejía *et al.*, 2011). Sequences were compared with the grapevine reference (‘Pinot Noir’) genome and to a recent work that reported the whole genome sequence of ‘Sultanina’, producing a catalog of structural variants for grapevine (Di Genova *et al.*, 2014). We found two amino acid modifications in ‘Sultanina’s’ *VvAGL11* coding region (Fig. 2). Arginine and threonine were substituted by two more hydrophobic amino acids, leucine and alanine, respectively. These amino acid substitutions were localized in the C-terminal portion of the *VvAGL11* protein, a well known region responsible for activity and ternary complex formation (Egea-Cortines *et al.*, 1999). Furthermore, 17 INDELS and 53 additional SNPs were identified in the second intron of the ‘Sultanina’ mutant allele (Supplementary Fig. 1). The second intron of MADS-box AG subfamily members is known to present a quite large size (*AtAG* ~3 kb, *AtAGL11* ~1,8 kb, and *VvAGL11* ~4 kb) and to be a region with important *cis* elements that may control tissue specific expression of AG and AG-like genes (Deyholos, 2000; Hong *et al.*, 2003). Interestingly, MADS-box proteins form complexes of high order, often in tetrameric junctions made by two dimers that bind to two DNA consensus sequences called CArG-box [CC(A/T)₆GG] separated by up to 300 bp (Liu *et al.*, 2008). A CArG-box modification was observed in ‘Sultanina’ *mut* allele, in the seventh putative sequence. This *cis* element is only 84 bp distant from another CArG-box what would suggest their relevance in gene regulation.

We performed the first analysis of *VvAGL11* transcript isoforms and compared them to the genome gene-predicted models (Fig. 3). The transcription profile of *VvAGL11* isoforms provided complete data for each expression isoform during the development of flowers, fruits and seeds. ‘Sultanina’ alleles presented a low level of expression in all organs and stages of development, including berries, while the ‘Chardonnay’ alleles were expressed at very low levels in the pulp, but highly expressed in seeds (Fig. 4). Of the two ‘Chardonnay’ alleles, one possesses a larger coding region, able to encode 20 additional amino acids (CH1). The sequence of CH1 transcripts was also found in a ‘Cabernet Sauvignon’ cDNA

library derived from berries (GenBank CB974197.1). This could cause conformational changes in the C-terminal portion of VvAGL11 protein, yet no abnormal seed morphogenesis is observed in Chardonnay neither in Cabernet Sauvignon cultivars. Thus it is clear that this 60 nucleotides may not be relevant to seed phenotype, therefore the biological relevance of these 20 additional amino acids in the C-terminal VvAGL11 protein remains to be elucidated. The relative expression of CH1 allele in seeds showed a fluctuation between 5 to 11-fold compared to fruit-set (Fig. 4). This instability was possibly due to the random amplification of *VvAGL11* pre-mRNA since primers employed allowed the amplification of intronic regions. Taken together, our results suggested that *VvAGL11* has indeed a major role in seed development. Without its expression (transcript accumulation), there is no normal seed formation.

The interaction of proteins encoded by MADS-box genes of classes A, B, C, D, and E forms the model of floral quartets (Theissen and Saedler, 2001; Dornelas and Dornelas, 2005). The AGAMOUS subfamily of *MADS-box* genes is known to have redundant functions in *A. thaliana* since *AtAGL11* is the major responsible gene in the regulation of ovule development along with other two genes of class D, *SHATTERPROOF1* and *SHATTERPROOF2* (Colombo *et al.*, 1995; Pinyopich *et al.*, 2003). The *VvAGL11* gene was proposed as a possible ortholog of *AtAGL11*, however no gene has been identified yet as orthologous to *SHP1* or *SHP2* in grapevine (Díaz-Riquelme *et al.*, 2009). These data suggest that *VvAGL11* might be the only class D gene in grapevine and a possible model of floral quartet would be composed by *VvAG1* and *VvAG2* (*AGAMOUS* genes), and one of the *SEPALLATA* genes of grapevine (*VvSEP1*, *VvSEP2*, *VvSEP3*, and *VvSEP4*) as previously described by Boss *et al.* (2002) and Díaz-Riquelme *et al.* (2008). Additional studies of the *VvAGL11* protein are necessary to confirm this hypothesis.

Previous studies have described grapevine's seed anatomy (Pratt, 1971; Ribereau-Gayon and Peynaud, 1980) as well as the changes of internal layers towards seed ripening (Cadot *et al.*, 2006), but, as much as we are concerned, no study has demonstrated detailed morphological parameters in seeds of stenospermocarpic grapevines, specially seed traces. The main objective of our morphological assay was to compare normal seeds from 'Chardonnay' and seed traces from 'Sultanina' as they resemble so differently from each other (Fig. 5). Our analysis showed that the most substantial difference was at the medium integument of the seed coat, which did not develop normally in 'Sultanina' seed traces,

preserving its initial features until the complete maturation of the fruits. As expected, 'Chardonnay' seeds developed correctly with the elongation and duplication of the medium integument, along with normal endosperm growth (Fig. 6). The nutrient flux in seeds occurs in a centripetal way, from the outer integument to the inner, going through apoplasts to the endosperm and to the embryo (Stadler *et al.*, 2005). Garcia *et al.* (2005) have previously shown that there is a strong maternal effect of the integuments in the control and determination of final seed size, due to the elongation of these layers and, in case of elongation absence, there is a compensation effect promoted by the duplication of these layers. Seed traces formation are likely originated due to disruption of seed tissues normal development. Probably the erroneous formation of the medium integument seed layer that occurs in stenospermocarpic grapevines could interfere in seed nutrient intake and cause the degeneration of some tissues during seed development, like the endosperm.

ISH assays were performed for better characterization of *VvAGL11* spatial and temporal expression patterns, in 'Chardonnay' and 'Sultanina', during seed morphogenesis in order to identify its transcription specific tissue localization. At the time of ISH assays the transcription isoforms were not known and the probe used for hybridization was designed according to the *VvAGL11* gene predicted model (GSVIVT01025945001) available at <http://www.genoscope.cns.fr>. Therefore, the whole probe (185 bp) hybridizes with CH1 mRNA isoform while 38 nucleotides hybridizes with CH2, SU1 and SU *mut* mRNA isoforms. The transcripts accumulation were localized in the medium integument layer of 'Chardonnay' seeds of 2 and 4 weeks (Fig. 7), while no accumulation of transcripts was found in 'Sultanina' seed traces (Fig. 8). After 6 weeks of 'Chardonnay' seed development, no hybridization signals were detected (Fig. 7L), this is due probably because the medium integument layer has already started lignification process, once it will be part of the mature and hard seed coat (Haughn & Chaudhury, 2005). This data is in agreement with *VvAGL11* RT-qPCR transcriptional profile, where 'Sultanina' shows almost no expression in any of the stages evaluated. The particular accumulation of *VvAGL11* transcripts in a specific layer of the seed coat in Chardonnay cultivar, combined with the morphological differences in this same layer when comparing Chardonnay and Sultanina cultivars, suggest that this gene is essential for the elongation and duplication of the medium integument of the seed coat. Furthermore, seed growth is dictated by the coordinated development of endosperm and

integuments and, hence, a spatial and temporal coordination of endosperm and integument growth is critical for normal seed development (Garcia *et al.*, 2005).

Taken together, our results allow us to propose that the absence of *VvAGL11* expression, a MADS-box class D gene of grapevine, is responsible for the erroneous development of seeds. The extreme diminution of final seed size could be due to the non-differentiation of the medium integument seed layer, causing the cease of the elongation and duplication of the seed coat. Within this loss of identity, the endosperm would have no normal development as well, stopping its growth and causing the death of the embryo leading to the formation of a seed trace.

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FIGURES

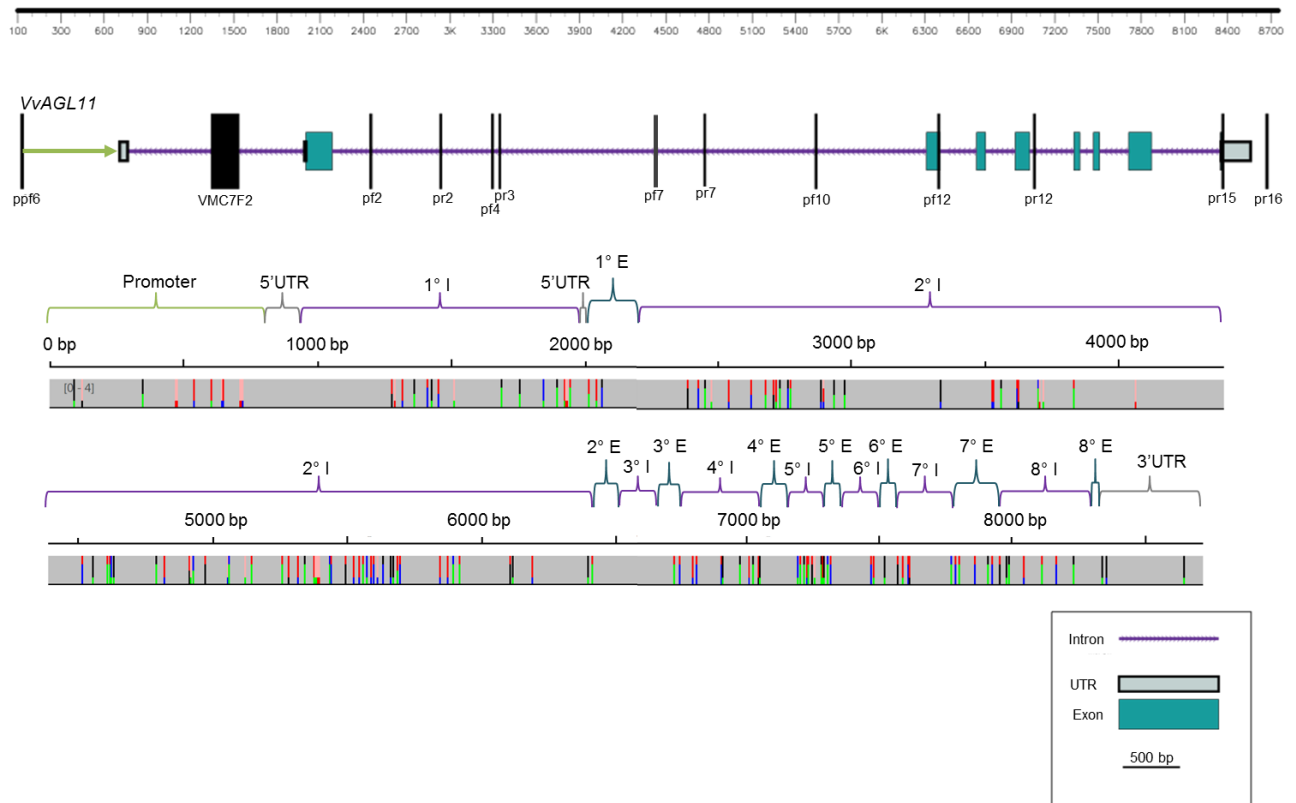


Fig. 1. Representative image of *VvAGL11*. DNA structure is composed by a promoter region, 5'-UTR, 8 introns, 8 exons and 3'-UTR. The black lines represents the localization of the primers used for allele amplification. Below, the alignment with the set of polymorphisms derived from 'Sultanina' and 'Chardonnay' *VvAGL11* alleles. Colors stripes represent SNPs (A-green, C-blue, T-red, G-black), and the salmon color indicates INDELS.

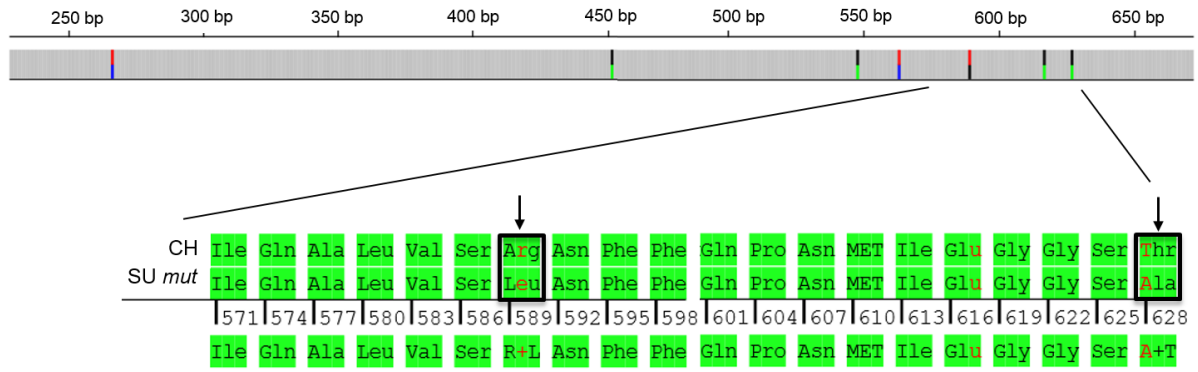


Fig. 2. Alignment of the coding regions of *VvAGL11* alleles where two amino acid modifications were observed. Alleles from ‘Pinot Noir’ (PN40024), ‘Chardonnay’ (both *VvAGL11* alleles) and ‘Sultanina’ normal and mutant allele. Colors stripes represent SNPs (A-green, C-blue, T-red, G-black).

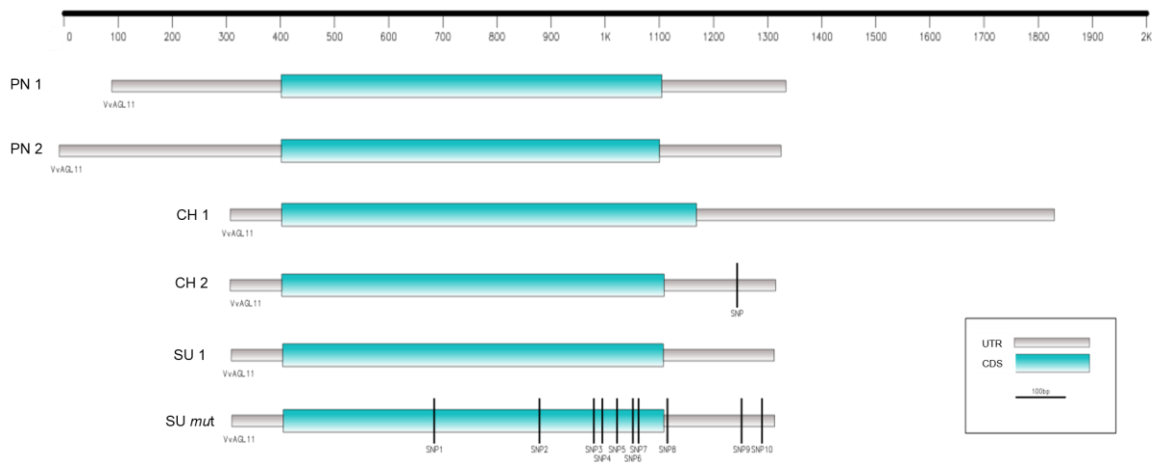


Fig. 3. Comparison between *VvAGL11* mRNA forms. PN 1 and PN 2 represent mRNA versions of *VvAGL11* from the Pinot Noir reference cultivar (VIT_218s0041g01880.1 and VIT_218s0041g01880.2, respectively). ‘Chardonnay’ allele 1 (CH1), ‘Chardonnay’ allele 2 (CH2), ‘Sultanina’ allele 1 (SU1) are compared with the ‘Sultanina’ mutant allele (SU *mut*). SNPs represented by black stripes (GenBank KM401845, KM401846, KM401847 and KM401848).

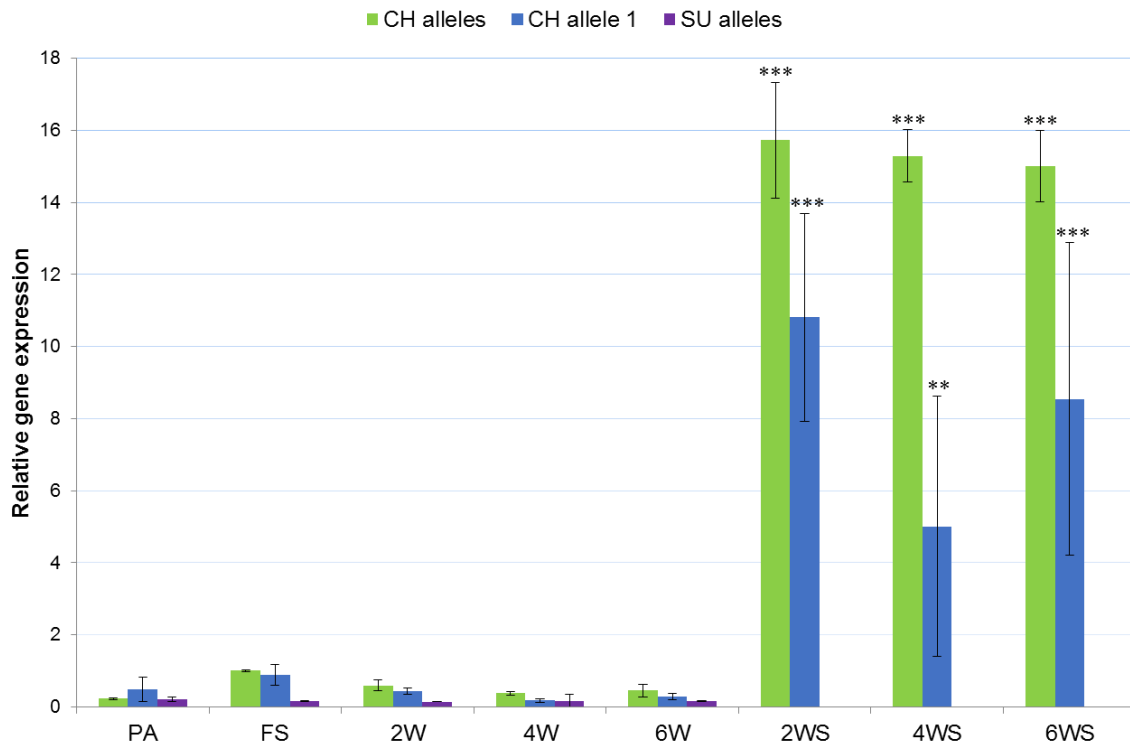


Fig. 4. Relative gene expression of *VvAGL11* alleles from ‘Chardonnay’ and ‘Sultanina’ via RT-qPCR. The developmental stages sampled are shown in the x-axis: **(PA)** pre anthesis flower; **(FS)** fruit set; **(2W-4W-6W)** fruits at 2, 4 and 6 weeks after fruit set and ‘Chardonnay’ seed stages **(2WS-4WS-6WS)**. ‘Chardonnay’ fruit-set was used as reference. The relative expression expressed in the y-axis was calculated by the Pfaffl (2001) method. Stander deviation is showed for each stage and sample. Asterisks indicate statistical significance between ‘Chardonnay’ alleles in seed stages to others stages evaluated for the ‘Chardonnay’ and ‘Sultanina’ alleles (Tukey test: ** $p < 0.01$, *** $p < 0.001$).

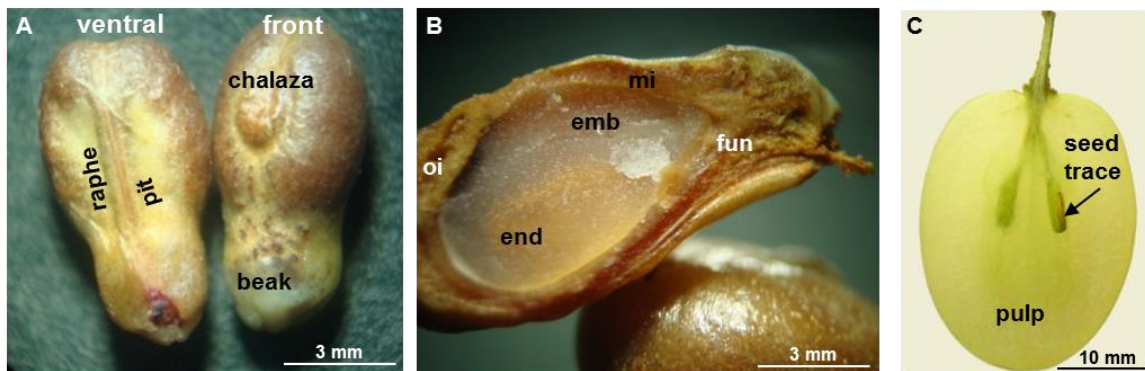


Fig. 5. Seeds and seed traces in *V. vinifera*. (A) Mature 'Chardonnay' (CH) seeds at their ventral and frontal view. Main structures are indicated. (B) CH seed cross section, with the internal view exhibiting outer integument (oi), medium integument (mi), endosperm (end), embryo (emb), and funiculus (fun). (C) Mature 'Sultanina' (SU) berry showing its seed traces.

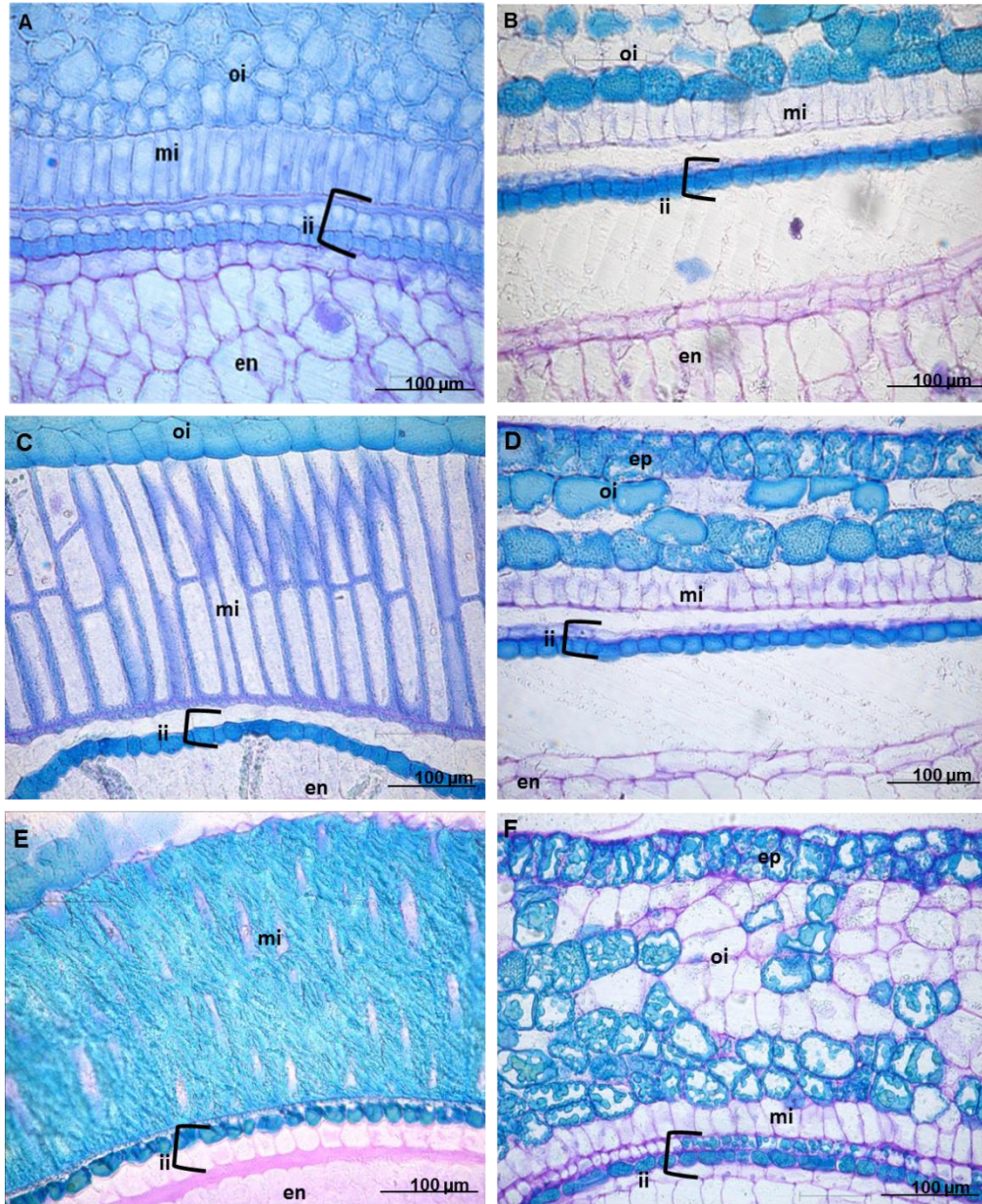


Fig. 6. Grapevine seed structure development from 2 to 6 weeks after fruit set. Light micrographs of grape seed cross sections were stained with toluidine blue O. The cross sections in the median area of the seeds of ‘Chardonnay’ and seed traces of ‘Sultanina’ enable the visualization of three defined integuments. ‘Chardonnay’ seed: 2 weeks (A), 4 weeks (C) and 6 weeks (E). ‘Sultanina’ seed trace: 2 weeks (B), 4 weeks (D) and 6 weeks (F). Endosperm (en), epidermis (ep), internal integument (ii), medium integument (mi), outer integument (oi).

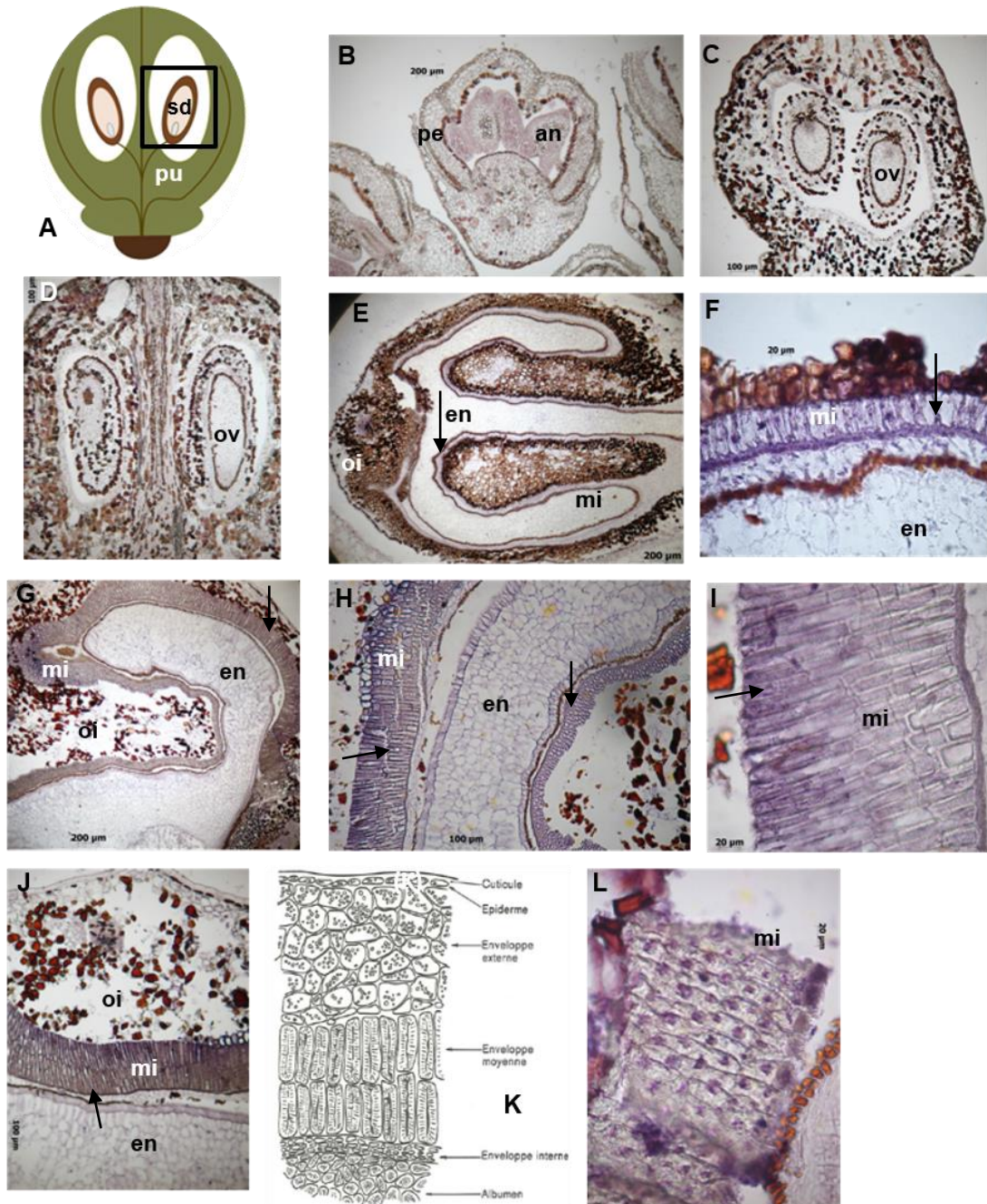


Fig. 7. *In situ* hybridization of *VvAGL11* in ‘Chardonnay’ reproductive tissues and organs, from flowers to seeds after 6 weeks of fruit development. (A) Schematic berry. No hybridization signal is visible in (B) H1 flower stage, (C) pre anthesis flower stage and (D) fruit set flower stage. *VvAGL11* expression is detectable in the medium integument of (E-F) 2 weeks seeds and (G-J) 4 weeks seeds (black arrows). No hybridization signal is visible in (L) 6 week seeds, compared to the control slides (Supplementary Fig. 2). (K) Schematic figure of the grapevine seed layers (Pratt, 1971). Anther (an), endosperm (en), medium integument (mi), outer integument (oi), petal (pe), pulp (pu), seed (sd).

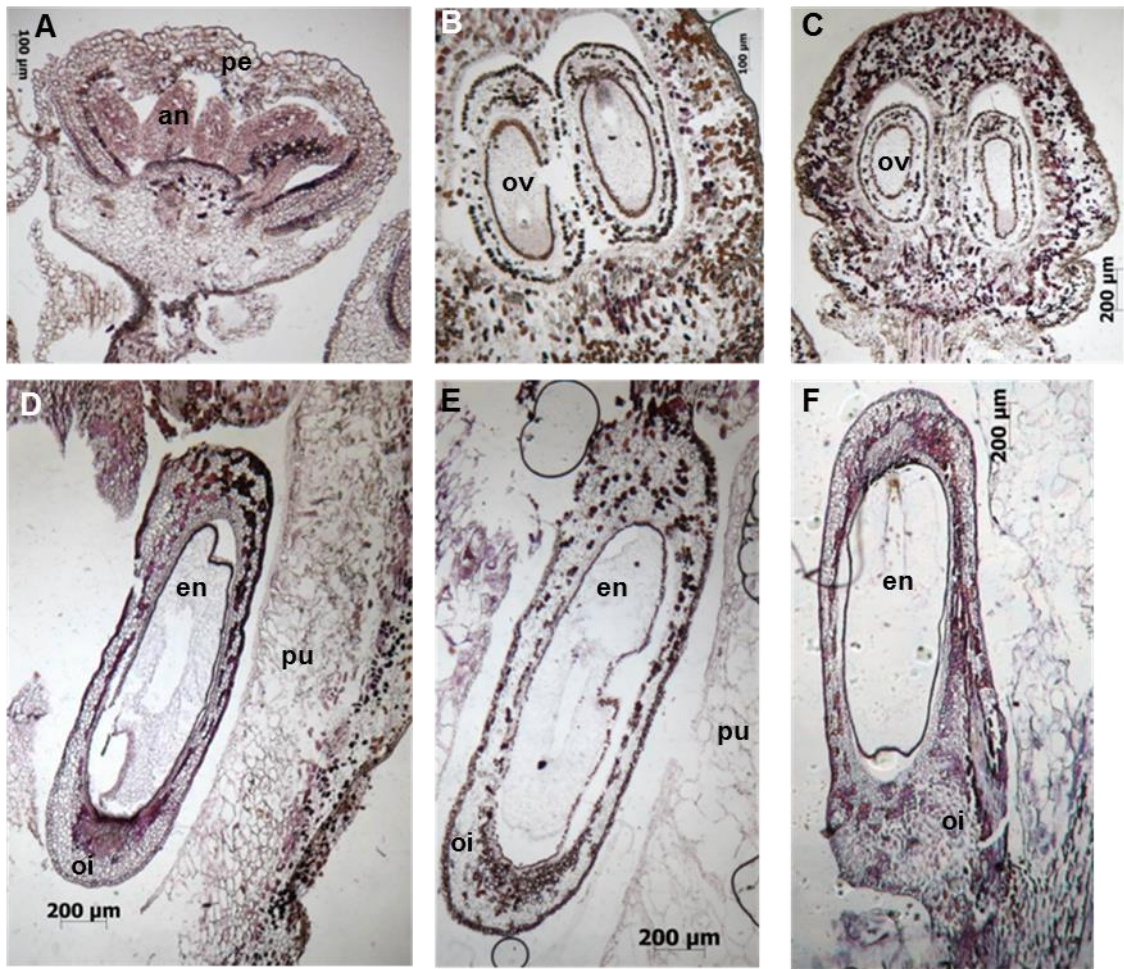


Fig. 8. *In situ* hybridization of *VvAGL11* in ‘Sultanina’ from flowers to 6 weeks of fruit development. No hybridization signal is visible in H1 flower stage (A), pre anthesis flower stage (B), fruit set flower stage (C) and in 2 (D), 4 (E) and 6 (F) weeks of fruit development. All samples compared with control slides (Supplementary Fig. 3). Anther (an), endosperm (en), medium integument (mi), outer integument (oi), petal (pe), pulp (pu).




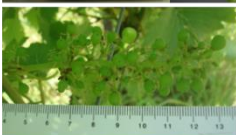



Supplementary Material

Supplementary Table 1. Primers designed for the amplification and sequencing of *VvAGL11*.

CDS+INTRON_AGL11_R_1	CAT CAG CAG ATG CCA GAT
CDS+INTRON_AGL11_F_1	CAA TAA ACC TAA CAG TGT AGT GAA C
CDS+INTRON_AGL11_R_2	GGA CAG TGG AAT ACA GAT TC
CDS+INTRON_AGL11_F_2	TAG GAA GGG ATT ACA TGG G
CDS+INTRON_AGL11_R_3	GAT GGC ATT GAG CAA ACA
CDS+INTRON_AGL11_F_3	CTA GGC AAC AAC CAG TTA G
CDS+INTRON_AGL11_R_4	ATA AGG AAG ACT TCT CCC C
CDS+INTRON_AGL11_F_4	CCC ATT ACA TCT TTG TGT GG
CDS+INTRON_AGL11_R_5	CTT TTT CTC CTA CTT TTC CTC
CDS+INTRON_AGL11_F_5	CGA TCT TAC ATG AGA TTT TAC C
CDS+INTRON_AGL11_R_6	TTG GAG CAT CGT CTT AGA G
CDS+INTRON_AGL11_F_6	GAGGAAAAGTAGGAGAAAAAG
CDS+INTRON_AGL11_R_7	AAA CCC CAC TGT GAT AGG
CDS+INTRON_AGL11_F_7	GGA TGG TGT CAT AAT GAT AGG
CDS+INTRON_AGL11_R_8	GAA ATA TCA TAG TCA TCC TCA C
CDS+INTRON_AGL11_F_8	TAA GGT GTC GAA GCC ATG
CDS+INTRON_AGL11_R_9	TGA GTT TCT TTG TGT GGA C
CDS+INTRON_AGL11_F_9	GAG AGC CTA TTT GGG ATA AC
CDS+INTRON_AGL11_R_10	GAG TTT GTA TGG AGA ATA GCA G
CDS+INTRON_AGL11_F_10	GCA AAG CAG TTA TTG AAA GC
CDS+INTRON_AGL11_R_11	GGA TTT GGA TTG ATT CAT TAA C
CDS+INTRON_AGL11_F_11	AGG AGA AAC CAG GAC AAG
CDS+INTRON_AGL11_R_12	CAA ATG CTC AGC CAG ATT AG
CDS+INTRON_AGL11_F_12	TGC CCA AGT AAG AAA ACT TC
CDS+INTRON_AGL11_R_13	CGG TAT CAA CTG TTG TGC
CDS+INTRON_AGL11_F_13	GGT AAC GAC ACC TAG ACA C
CDS+INTRON_AGL11_R_14	GCT TGA AAT GTC TCA TTA AGC
CDS+INTRON_AGL11_F_14	CGT ATA TCT CCG AAC CAA G
CDS+INTRON_AGL11_R_15	GGC GGA TAT TTT CTC CCA TC
CDS+INTRON_AGL11_F_15	CTC GGG TAC ACT CAT CTT TC
CDS+INTRON_AGL11_R_16	TGA TTA TCA ACA CCA AAC ATG GC
CDS+INTRON_AGL11_F_16	TTC AAG TTC GAC CTA ATT CCT GG
Promo_AGL11_R_1	GAT CTC GAT CTT TCC TCT CCC
Promo_AGL11_F_1	CCT CAC CCT ATT TTT CGA ACA AC
Promo_AGL11_R_2	GAG ATC TGA ATG ATG ATA GAC TGA G
Promo_AGL11_F_2	CTT GAG GGG AAA AAG CCA G
Promo_AGL11_R_3	CAT AAA CCC AGG ATA TGG ATG AG
Promo_AGL11_F_3	TTG CCT ACA TAT GAA CAT CTG C
Promo_AGL11_R_4	ATT TGG AGA GAT GGG CAC TG
Promo_AGL11_F_4	AAA GTC ACT TTC TAC ATC TGC AC
Promo_AGL11_R_5	GGA GAG ATG TGA GAC GGA TAT G

Promo_AGL11_F_5	AGA AGA AAG TTT GCA GTT TGT G
Promo_AGL11_R_6	CAA AGT CGG ATA CAA GAC ATA GTA G
Promo_AGL11_F_6	CCA CTG ATA TGG ATT GAT TTG CC

Supplementary Table 2. Stages of development of grapevine reproductive organs sampled for the evaluation of *VvAGL11* transcriptional profile (except H1) and for the *in situ* hybridization experiment. Dates of harvesting, the phenological stage according to Baggiolini (1952), grapevine cultivars and a representative picture of each sample are represented.

Dates	Phenological stage (Baggiolini, 1952)	Sample	Cultivar	
09/11/12	H1	Flowers at inicial development	Chardonnay	
08/30/12			Sultanina	
10/08/12	Pre anthesis	Flowers before anthesis	Chardonnay	
10/08/12			Sultanina	
10/22/12	Fruit-set	Flowers in anthesis	Chardonnay	
10/08/12			Sultanina	
11/05/12	2 weeks of development	Fruits (Pulp and seeds)	Chardonnay	
11/22/12			Sultanina	
11/19/12	4 weeks of development	Fruits (Pulp and seeds)	Chardonnay	
11/05/12			Sultanina	
12/03/12	6 weeks of development	Fruits (Pulp and seeds)	Chardonnay	
11/19/12			Sultanina	
01/02/13	Maturation	Fruits (Pulp and seeds)	Chardonnay	
12/17/12			Sultanina	

CLUSTAL multiple sequence alignment by MUSCLE (3.8)

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su_mut      CCACTGATATGGATTGATTTGCCTTTTAAATTGAAGACCCTTTTCTGTTGGTAACTTTGT
su_Di Genova CCACTGATATGGATTGATTTGCCTTTTAAATTGAAGACCCTTTTCTGTTGGTAACTTTGT
ch2         CCACTGATATGGATTGATTTGCCTTTTAAATTGAAGACCCTTTTCTGTTGGTAACTTTGT
VIT_218S0041G01880.2 CCACTGATATGGATTGATTTGCCTTTTAAATTGAAGACCCTTTTCTGTTGGTAACTTTGT
su_wt      CCACTGATATGGATTGATTTGCCTTTTAAATTGAAGACCCTTTTCTGTTGGTAACTTTGT
ch1        CCACTGATATGGATTGATTTGCCTTTTAAATTGAAGACCCTTTTCTGTTGGTAACTTTGT
*****

su_mut      GAAGGCATGAACGAAGTTGATTACTGAAAATATTAGCAACATTACTAGGGTTTTGGCGGA
su_Di Genova GAAGGCATGAACGAAGTTGATTACTGAARATATTAGCAACATTACTAGGGTTTTGGCGGA
ch2         GAAGGCATGAACGAAGTTGATTACTGAAGATATTAGCAACATTACTAGGGTTTT---GGA
VIT_218S0041G01880.2 GAAGGCATGAACGAAGTTGATTACTGAAGATATTAGCAACATTACTAGGGTTTT---GGA
su_wt      GAAGGCATGAACGAAGTTGATTACTGAAGATATTAGCAACATTACTAGGGTTTT---GGA
ch1        GAAGGCATGAACGAAGTTGATTACTGAAGATATTAGCAACATTACTAGGGTTTT---GGA
*****

su_mut      GATGAGACTGTAAGTTGGCTAAAACCTAGAGGCATGTTGTTTTTAATAATTGGAGAT
su_Di Genova GATGAGACTGTAAGTTGGCTAAAACCTAGAGGCATGTTGTTTTTAATAATTGGAGAT
ch2         GATGAGACTGTAAGTTGGCTAAAACCTAGAGGCATGTTGTTTTTAATAATTGGAGAT
VIT_218S0041G01880.2 GATGAGACTGTAAGTTGGCTAAAACCTAGAGGCATGTTG-TTTTTAATAATTGGAGAT
su_wt      GATGAGACTGTAAGTTGGCTAAAACCTAGAGGCATGTTG-TTTTTAATAATTGGAGAT
ch1        GATGAGACTGTAAGTTGGCTAAAACCTAGAGGCATGTTG-TTTTTAATAATTGGAGAT
*****

su_mut      ACCATATATGTATGTTGCGAGTGCAGATATACGTGGAAGCACAAAAATCCTAACCTGTT
su_Di Genova ACCATATATGTATGTTGCGAGTGCAGATATACGTGGAAGCACAAAAATCCTAACCTGTT
ch2         ACCATATATGTATGTTGCGAGTGCAGATATACGTGGAAGCACAAAAATCCTAACCTGTT
VIT_218S0041G01880.2 ACCATATATGTATGTTGCGAGTGCAGATATACGTGGAAGCACAAAAATCCTAACCTGTT
su_wt      ACCATATATGTATGTTGCGAGTGCAGATATACGTGGAAGCACAAAAATCCTAACCTGTT
ch1        ACCATATATGTATGTTGCGAGTGCAGATATACGTGGAAGCACAAAAATCCTAACCTGTT
*****

su_mut      AAACGTAAATTTGGGAAGATATTGTGTTTCTTTTACGTGTTGGATACAACATTGGGTTGT
su_Di Genova AAACGTAAATTTGGGAAGATATTGTGTTTCTTTTACGTGTTGGATACAACATTGGGTTGT
ch2         AAACGTAAATTTGGGAAGATATTGTGTTTCTTTTACGTGTTGGATACAACATTGGGTTGT
VIT_218S0041G01880.2 AAACGTAAATTTGGGAAGATATTGTGTTTCTTTTACGTGTTGGATACAACATTGGGTTGT
su_wt      AAACGTAAATTTGGGAAGATATTGTGTTTCTTTTACGTGTTGGATACAACATTGGGTTGT
ch1        AAACGTAAATTTGGGAAGATATTGTGTTTCTTTTACGTGTTGGATACAACATTGGGTTGT
*****

su_mut      GGCCTTGAAGAAAATAAAGAAGAGAAGAAAGTTTGCAGTTTATGGTGGGAATTTGGAAC
su_Di Genova GGCCTTGAAGAAAATAAAGAAGAGAAGAAAGTTTGCAGTTTATGGTGGGAATTTGGAAC
ch2         GGCCTTGAAGAAAATAAAGAAGAGAAGAAAGTTTGCAGTTTATGGTGGGAATTTGGAAC
VIT_218S0041G01880.2 GGCCTTGAAGAAAATAAAGAAGAGAAGAAAGTTTGCAGTTTATGGTGGGAATTTGGAAC
su_wt      GGCCTTGAAGAAAATAAAGAAGAGAAGAAAGTTTGCAGTTTATGGTGGGAATTTGGAAC
ch1        GGCCTTGAAGAAAATAAAGAAGAGAAGAAAGTTTGCAGTTTATGGTGGGAATTTGGAAC
*****

su_mut      GCAAAGCTGTTTACATTATAAAACTACTATGCTTGTATCCGACTTTGGGTAATAAGCCA
su_Di Genova GCAAAGCTGTTTACATTATAAAACTACTATGCTTGTATCCGACTTTGGGTAATAAGCCA
ch2         GCAAAGCTGTTTACATTATAAAACTACTATGCTTGTATCCGACTTTGGGTAATAAGCCA
VIT_218S0041G01880.2 GCAAAGCTGTTTACATTATAAAACTACTATGCTTGTATCCGACTTTGGGTAATAAGCCA
su_wt      GCAAAGCTGTTTACATTATAAAACTACTATGCTTGTATCCGACTTTGGGTAATAAGCCA
ch1        GCAAAGCTGTTTACATTATAAAACTACTATGCTTGTATCCGACTTTGGGTAATAAGCCA
*****

su_mut      TTCGGAAGTCTTTTTCTTCCTCAACCC-----CTCTCTCTCTCTCTCTCTTTGAAGAAG
su_Di Genova TTCGGAAGTCTTTTTCTTCCTCAACCC-----CTCTCTCTCTCTCTCTCTTTGAAGAAG
ch2         TTCGGAAGTCTTTTTCTTCCTCAACCC-----CTCTCTCTCTCTCTCTCTTTGAAGAAG
VIT_218S0041G01880.2 TTCGGAAGTCTTTTTCTTCCTCAACCC-----CTCTCTCTCTCTCTCTCTTTGAAGAAG
su_wt      TTCGGAAGTCTTTTTCTTCCTCAACCC-----CTCTCTCTCTCTCTCTCTTTGAAGAAG
ch1        TTCGGAAGTCTTTTTCTTCCTCAACCC-----CTCTCTCTCTCTCTCTCTTTGAAGAAG
*****
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su_mut      TTTCCATTTCTTCCTCTTTCTTTTGTGTCAGTGCCCATCTCTCCAAATTTACTTCCACCTCT
su_Di Genova TTTCCATTTCTTCCTCTTTCTTTTGTGTCAGTGCCCATCTCTCCAAATTTACTTCCACCTCT
ch2         TTTCCATTTCTTCCTCTTTCTTTTGTGTCAGTGCCCATCTCTCCAAATTTACTTCCACCTCT
VIT_218S0041G01880.2 TTTCCATTTCTTCCTCTTTCTTTTGTGTCAGTGCCCATCTCTCCAAATTTACTTCCACCTCT
su_wt      TTTCCATTTCTTCCTCTTTCTTTTGTGTCAGTGCCCATCTCTCCAAATTTACTTCCACCTCT
ch1         TTTCCATTTCTTCCTCTTTCTTTTGTGTCAGTGCCCATCTCTCCAAATTTACTTCCACCTCT
*****

su_mut      TACATTTTCTTACCATTCTTTTTAGATTTCTTGGCTTGATTTACTCTCTTCTTCTCCTGCAA
su_Di Genova TACATTTTCTTACCATTCTTTTTAGATTTCTTGGCTTGATTTACTCTCTTCTTCTCCTGCAA
ch2         TACATTTTCTTACCATTCTTTTTAGATTTCTTGGCTTGATTTACTCTCTTCTTCTCCTGCAA
VIT_218S0041G01880.2 TACATTTTCTTACCATTCTTTTTAGATTTCTTGGCTTGATTTACTCTCTTCTTCTCCTGCAA
su_wt      TACATTTTCTTACCATTCTTTTTAGATTTCTTGGCTTGATTTACTCTCTTCTTCTCCTGCAA
ch1         TACATTTTCTTACCATTCTTTTTAGATTTCTTGGCTTGATTTACTCTCTTCTTCTCCTGCAA
*****

su_mut      CACTTTACTCTTCAGTTCTTGATTCTCTTTTGCCTTCTTCATGCATTTGTTTCCATTCTT
su_Di Genova CACTTTACTCTTCAGTTCTTGATTCTCTTTTGCCTTCTTCATGCATTTGTTTCCATTCTT
ch2         CACTTTACTCTTCAGTTCTTGATTCTCTTTTGCCTTCTTCATGCATTTGTTTCCATTCTT
VIT_218S0041G01880.2 CACTTTACTCTTCAGTTCTTGATTCTCTTTTGCCTTCTTCATGCATTTGTTTCCATTCTT
su_wt      CACTTTACTCTTCAGTTCTTGATTCTCTTTTGCCTTCTTCATGCATTTGTTTCCATTCTT
ch1         CACTTTACTCTTCAGTTCTTGATTCTCTTTTGCCTTCTTCATGCATTTGTTTCCATTCTT
*****

su_mut      TAATTAGTCATTTTCTTATTCTAAACTTTCTTTTCCCTTTTTTCGTTATTTTCAGAATGC
su_Di Genova TAATTAGTCATTTTCTTATTCTAAACTTTCTTTTCCCTTTTTTCGTTATTTTCAGAATGC
ch2         TAATTAGTCATTTTCTTATTCTAAACTTTCTTTTCCCTTTTTTCGTTATTTTCAGAATGC
VIT_218S0041G01880.2 TAATTAGTCATTTTCTTATTCTAAACTTTCTTTTCCCTTTTTTCGTTATTTTCAGAATGC
su_wt      TAATTAGTCATTTTCTTATTCTAAACTTTCTTTTCCCTTTTTTCGTTATTTTCAGAATGC
ch1         TAATTAGTCATTTTCTTATTCTAAACTTTCTTTTCCCTTTTTTCGTTATTTTCAGAATGC
*****

su_mut      AGTTTGATTATTTTGCCTTTTTTTTTCATGTTAAAACAGTATTTGTTTGGTAGTTCTCAG
su_Di Genova AGTTTGATTATTTTTC -TTTTTTTTCATGTTAAAACAGTATTTGTTTGGTAGTTCTCAG
ch2         AGTTTGATTATTTTGC -TTTTTTTTCATGTTAAAACAGTATTTGTTTGGTAGTTCTCAG
VIT_218S0041G01880.2 AGTTTGATTATTTTTC -TTTTTTTTCATGTTAAAACAGTATTTGTTTGGTAGTTCTCAG
su_wt      AGTTTGATTATTTTTC -TTTTTTTTCATGTTAAAACAGTATTTGTTTGGTAGTTCTCAG
ch1         AGTTTGATTATTTTTC -TTTTTTTTCATGTTAAAACAGTATTTGTTTGGTAGTTCTCAG
***** * *****

su_mut      TTTATTTTTTCCATAAATTTCTGTTTCAAAAACCTCTTGAAGGGAAAAAGCCAGAATTTT
su_Di Genova TTTATTTTTTCCATAAATTTCTGTTTCAAAAACCTCTTGAAGGGAAAAAGCCAGAATTTT
ch2         TTTATTTTTTCCATAAATTTCTGTTTCAAAAACCTCTTGAAGGGAAAAAGCCAGAATTTT
VIT_218S0041G01880.2 TTTATTTTTTCCATAAATTTCTGTTTCAAAAACCTCTTGAAGGGAAAAAGCCAGAATTTT
su_wt      TTTATTTTTTCCATAAATTTCTGTTTCAAAAACCTCTTGAAGGGAAAAAGCCAGAATTTT
ch1         TTTATTTTTTCCATAAATTTCTGTTTCAAAAACCTCTTGAAGGGAAAAAGCCAGAATTTT
*****

su_mut      TTTTCCTGCGTTCCTGGTCTTAAGCTTCTCATCCATATCCTGGGTTTATGGGAAATGTGTT
su_Di Genova TTTTCCTGCGTTCCTGGTCTTAAGCTTCTCATCCATATCCTGGGTTTATGGGAAATGTGTT
ch2         TTTTCCTGCGTTCCTGGTCTTAAGCTTCTCATCCATATCCTAGGTTTATGGGAAATGTGTT
VIT_218S0041G01880.2 TTTTCCTGCGTTCCTGGTCTTAAGCTTCTCATCCATATCCTGGGTTTATGGGAAATGTGTT
su_wt      TTTTCCTGCGTTCCTGGTCTTAAGCTTCTCATCCATATCCTGGGTTTATGGGAAATGTGTT
ch1         TTTTCCTGCGTTCCTGGTCTTAAGCTTCTCATCCATATCCTGGGTTTATGGGAAATGTGTT
*****

su_mut      AAGGGTCATGGGGTTTTTGAGAAATGTG----GAGAGAGAGAGAGAGAGAGAGAGAGAGA
su_Di Genova AAGGGTCATGGGGTTTTTGAGAAATGTG----GAGAGAGAGAGAGAGAGAGAGAGAGAGA
ch2         AAGGGTCATGGGGTTTTTGAGAAATGTG-----GAGAGAGAGAGAGAGAGAGAGAGAGAGA
VIT_218S0041G01880.2 AAGGGTCATGGGGTTTTTGAGAAATGTG-----GAGAGAGAGAGAGAGAGAGAGAGAGAGA
su_wt      AAGGGTCATGGGGTTTTTGAGAAATGTG-----GAGAGAGAGAGAGAGAGAGAGAGAGAGA
ch1         AAGGGTCATGGGGTTTTTGAGAAATGTG-----GAGAGAGAGAGAGAGAGAGAGAGAGAGA
***** *****

su_mut      GAGAGAGATGTGTAGACATGCACCCACTTGTAGTAGTATTAGTAGTGGAGATAGATCCTG
su_Di Genova GAGAGAGATGTGTAGACATGCACCCACTTGTAGTAGTATTAGTAGTGGAGATAGATCCTG
ch2         GAGAGAGATGTGTAGACATGCACCCACTTGTAGTAGTATTAGTAGTGGAGATAGATCCTG

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VIT_218S0041G01880.2 GAGAGAGATGTGTAGACATGCACCCACTTGTAGTAGTATTAGTAGTGGAGATAGATCCTG
su_wt GAGAGAGATGTGTAGACATGCACCCACTTGTAGTAGTATTAGTAGTGGAGATAGATCCTG
ch1 GAGAGAGATGTGTAGACATGCACCCACTTGTAGTAGTATTAGTAGTGGAGATAGATCCTG

su_mut GGTGATTTTTCAAATGGTAGATCATGTTCTTCTCTTTGTTCTCATCTCTTCATGTTCT
su_Di Genova GGTGATTTTTCAAATGGTAGATCATGTTCTTCTCTTTGTTCTCATCTCTTCATGTTCT
ch2 GGTGATTTTTCAAATGGTAGATCATGTTCTTCTCTTTGTTCTCATCTCTTCATGTTCT
VIT_218S0041G01880.2 GGTGATTTTTCAAATGGTAGATCATGTTCTTCTCTTTGTTCTCATCTCTTCATGTTCT
su_wt GGTGATTTTTCAAATGGTAGATCATGTTCTTCTCTTTGTTCTCATCTCTTCATGTTCT
ch1 GGTGATTTTTCAAATGGTAGATCATGTTCTTCTCTTTGTTCTCATCTCTTCATGTTCT

su_mut TTTTTTCTCAGTGGCTTACAGATTTTCTGTCTCCCAATCTTTGGTTTCCATCTCTTTT
su_Di Genova TTTTTTCTCAGTGGCTTACAGATTTTCTGTCTCCCAATCTTTGGTTTCCATCTCTTTT
ch2 TTTTTTCTCAGTGGCTTACAGATTTTCTGTCTCCCAATCTTTGGTTTCCATCTCTTTT
VIT_218S0041G01880.2 TTTTTTCTCAGTGGCTTACAGATTTTCTGTCTCCCAATCTTTGGTTTCCATCTCTTTT
su_wt TTTTTTCTCAGTGGCTTACAGATTTTCTGTCTCCCAATCTTTGGTTTCCATCTCTTTT
ch1 TTTTTTCTCAGTGGCTTACAGATTTTCTGTCTCCCAATCTTTGGTTTCCATCTCTTTT

su_mut CCAAGATCTCCTTATTTTCTACATTATCATCATTATTTTATTATATACACAAGCCTCAC
su_Di Genova CCAAGATCTCCTTATTTTCTACATTATCATCATTATTTTATTATATACACAAGCCTCAC
ch2 CCAAGTCTCCTTATTTTCTTACATTATCATCATTATTTTATTATATACACAAGCCTCAC
VIT_218S0041G01880.2 CCAAGATCTCCTTATTTTCTACATTATCATCATTATTTTATTATATACACAAGCCTCAC
su_wt CCAAGATCTCCTTATTTTCTTACATTATCATCATTATTTTATTATATACACAAGCCTCAC
ch1 CCAAGATCTCCTTATTTTCTTACATTATCATCATTATTTTATTATATACACAAGCCTCAC

su_mut CCTATTTTTCAAACAACATCTAACTTCTCATTTCTCAGTCTATCATCATTAGAG-----
su_Di Genova CCTATTTTTCGAACAACATCTAACTTCTCATTTCTCAGTCTATCATCATTAGAGATCTCT
ch2 CCTATTTTTCAAACAACATCTAACTTCTCATTTCTCAGTCTATCATCATTAGAGATCTCT
VIT_218S0041G01880.2 CCTATTTTTCGAACAACATCTAACTTCTCATTTCTCAGTCTATCATCATTAGAGATCTCT
su_wt CCTATTTTTCGAACAACATCTAACTTCTCATTTCTCAGTCTATCATCATTAGAGATCTCT
ch1 CCTATTTTTCGAACAACATCTAACTTCTCATTTCTCAGTCTATCATCATTAGAGATCTCT

su_mut --TCTCTCCTCTCTCCTCTTCCACCCTCTACCAAAAACACACATTTTTTCGGTAGATCTTC
su_Di Genova CCTCTCTCCTCTCTCCTCTTCCACCCTCTACCAAAAACACACATTTTTTCGGTAGATCTTC
ch2 CCTCTCTCCTCTCTCCTCTTCCACCCTCTACCAAAAACACAAATTTTTTCGGTAGATCTTC
VIT_218S0041G01880.2 CCTCTCTCCTCTCTCCTCTTCCACCCTCTACCAAAAACACACATTTTTTCGGTAGATCTTC
su_wt CCTCTCTCCTCTCTCCTCTTCCACCCTCTACCAAAAACACACATTTTTTCGGTAGATCTTC
ch1 CCTCTCTCCTCTCTCCTCTTCCACCCTCTACCAAAAACACACATTTTTTCGGTAGATCTTC

su_mut TCTCACAATTACATGCATTATTACCCTTCATATGTCACTCATACTTCAATTTT-----TT
su_Di Genova TCTCACAATTACATGCATTATTACCCTTCATATGTCACTCATACTTCAATTTT-----TT
ch2 TCTCACAATTACATGCATTATTACCCTTCATATGTCACTCATACTTCAATTTT-----TT
VIT_218S0041G01880.2 TCTCACAATTACATGCATTATTACCCTTCATATGTCACTCATACTTCAATTTT-----TT
su_wt TCTCACAATTACATGCATTATTACCCTTCATATGTCACTCATACTTCAATTTT-----TT
ch1 TCTCACAATTACATGCATTATTACCCTTCATATGTCACTCATACTTCAATTTT-----TT

su_mut TTTTTTTTTTAAAGCTTGAACAGATCTCCAACCTTTCTCTAGTATTTTGTAAATCTTTAG
su_Di Genova TTTTTTTTTTAAAGCTTGAACAGATCTCCAACCTTTCTCTAGTATTTTGTAAATCTTTAG
ch2 TTTTTTTTTTAAAGCTTGAACAGATCTCCAACCTTTCTCTAGTATTTTGTAAATCTTTAG
VIT_218S0041G01880.2 TTTTTTTTTTAAAGCTTGAACAGATCTCCAACCTTTCTCTAGTATTTTGTAAATCTTTAG
su_wt TTTTTTTTTTAAAGCTTGAACAGATCTCCAACCTTTCTCTAGTATTTTGTAAATCTTTAG
ch1 TTTTTTTTTTAAAGCTTGAACAGATCTCCAACCTTTCTCTAGTATTTTGTAAATCTTTAG

su_mut CAGTTTCATAAAGAGATGCTTTGGTAATTTTCTAATATTTTCAGTTGGTGATATAATA
su_Di Genova CAGTTTCATAAAGAGATGCTTTGGTAATTTTCTAATATTTTCAGTTGGTGATATAATA
ch2 CAGTTTCATAAAGAGATGCTTTGGTAATTTTCTAATATTTTCAGTTGGTGATATAATA
VIT_218S0041G01880.2 CAGTTTCATAAAGAGATGCTTTGGTAATTTTCTAATATTTTCAGTTGGTGATATAATA
su_wt CAGTTTCATAAAGAGATGCTTTGGTAATTTTCTAATATTTTCAGTTGGTGATATAATA
ch1 CAGTTTCATAAAGAGATGCTTTGGTAATTTTCTAATATTTTCAGTTGGTGATATAATA

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su_mut      AAACAATGTATAGTACTACAAAAATTTTGTTCCTTATATACCTAAGCCATTTTTTCTTTTG
su_Di Genova WAACAATGTATAGTACTACAAAAATTTTGTTCCTTATATACCTAAGCCATTTTTTCTTTTG
ch2         TAACAATGTATAGTACTACAAAAATTTTGTTCCTTATATACCTAAGCCATTTTTTCTTTTG
VIT_218S0041G01880.2 TAACAATGTATAGTACTACAAAAATTTTGTTCCTTATATACCTAAGCCATTTTTTCTTTTG
su_wt      TAACAATGTATAGTACTACAAAAATTTTGTTCCTTATATACCTAAGCCATTTTTTCTTTTG
ch1        TAACAATGTATAGTACTACAAAAATTTTGTTCCTTATATACCTAAGCCATTTTTTCTTTTG
*****

su_mut      GTGCAATAAACCTAACAGTGTAGTGAACATGGGGAGAGGAAAGATCGAGATCAAGAGGAT
su_Di Genova GTGCAATAAACCTAACAGTGTAGTGAACATGGGGAGAGGAAAGATCGAGATCAAGAGGAT
ch2         GTGCAATAAACCTAACAGTGTAGTGAACATGGGGAGAGGAAAGATCGAGATCAAGAGGAT
VIT_218S0041G01880.2 GTGCAATAAACCTAACAGTGTAGTGAACATGGGGAGAGGAAAGATCGAGATCAAGAGGAT
su_wt      GTGCAATAAACCTAACAGTGTAGTGAACATGGGGAGAGGAAAGATCGAGATCAAGAGGAT
ch1        GTGCAATAAACCTAACAGTGTAGTGAACATGGGGAGAGGAAAGATCGAGATCAAGAGGAT
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su_mut      CGAAAACACGACCAACCGTCAGGTCACATTCTGCAAGCGAAGGAATGGGCTTTTGAAGAA
su_Di Genova CGAAAACACGACCAACCGTCAGGTCACATTCTGCAAGCGAAGGAATGGGCTTTTGAAGAA
ch2         CGAAAACACGACCAACCGTCAGGTCACATTCTGCAAGCGAAGGAATGGGCTTTTGAAGAA
VIT_218S0041G01880.2 CGAAAACACGACCAACCGTCAGGTCACATTCTGCAAGCGAAGGAATGGGCTTTTGAAGAA
su_wt      CGAAAACACGACCAACCGTCAGGTCACATTCTGCAAGCGAAGGAATGGGCTTTTGAAGAA
ch1        CGAAAACACGACCAACCGTCAGGTCACATTCTGCAAGCGAAGGAATGGGCTTTTGAAGAA
*****

su_mut      GGCTTATGAATTATCAGTGCATGTGATGCAGAAGTTGCCCTCATCGTCTTCCAGCCG
su_Di Genova GGCTTATGAATTATCAGTGCATGTGATGCAGAAGTTGCCCTCATCGTCTTCCAGCCG
ch2         GGCTTATGAATTATCAGTGCATGTGATGCAGAAGTTGCCCTCATCGTCTTCCAGCCG
VIT_218S0041G01880.2 GGCTTATGAATTATCAGTGCATGTGATGCAGAAGTTGCCCTCATCGTCTTCCAGCCG
su_wt      GGCTTATGAATTATCAGTGCATGTGATGCAGAAGTTGCCCTCATCGTCTTCCAGCCG
ch1        GGCTTATGAATTATCAGTGCATGTGATGCAGAAGTTGCCCTCATCGTCTTCCAGCCG
*****

su_mut      CGGTCGAGTCTATGAGTACTCAAACAACAAGTAATAATTTTCTCCACCATTCTTCAACC
su_Di Genova CGGTCGAGTCTATGAGTACTCAAACAACAAGTAATAATTTTCTCCACCATTCTTCAACC
ch2         CGGTCGAGTCTATGAGTACTCAAACAACAAGTAATAATTTTCTCCACCATTCTTCAACC
VIT_218S0041G01880.2 CGGTCGAGTCTATGAGTACTCAAACAACAAGTAATAATTTTCTCCACCATTCTTCAACC
su_wt      CGGTCGAGTCTATGAGTACTCAAACAACAAGTAATAATTTTCTCCACCATTCTTCAACC
ch1        CGGTCGAGTCTATGAGTACTCAAACAACAAGTAATAATTTTCTCCACCATTCTTCAACC
*****

su_mut      ATCTGCTGAATTTTCTATGTTTCATCTTTCTCCATTTTGGTGCATCATACGGGCAATTAAT
su_Di Genova ATCTGCTGAATTTTCTATGTTTCATCTTTCTCCATTTTGGTGCATCATACGGGCAATTAAT
ch2         ATCTGCTGAATTTTCTATGTTTCATCTTTCTCCATTTTGGTGCATCATACGGGCAATTAAT
VIT_218S0041G01880.2 ATCTGCTGAATTTTCTATGTTTCATCTTTCTCCATTTTGGTGCATCATACGGGCAATTAAT
su_wt      ATCTGCTGAATTTTCTATGTTTCATCTTTCTCCATTTTGGTGCATCATACGGGCAATTAAT
ch1        ATCTGCTGAATTTTCTATGTTTCATCTTTCTCCATTTTGGTGCATCATACGGGCAATTAAT
*****

su_mut      GGTTTTATATTGATGAGATTAATATCATCAGAAGTTGCAGAACCCTAATGTTATTAAGC
su_Di Genova GGTTTTATATTGATGAGATTAATATCATCAGAAGTTGCAGAACCCTAATGTTATTAAGC
ch2         GGTTTTATATTGATGAGATTAATATCATCAGAAGTTGCAGAACCCTAATGTTATTAAGC
VIT_218S0041G01880.2 GGTTTTATATTGATGAGATTAATATCATCAGAAGTTGCAGAACCCTAATGTTATTAAGC
su_wt      GGTTTTATATTGATGAGATTAATATCATCAGAAGTTGCAGAACCCTAATGTTATTAAGC
ch1        GGTTTTATATTGATGAGATTAATATCATCAGAAGTTGCAGAACCCTAATGTTATTAAGC
*****

su_mut      AAAG-AAAAAAAAAAGAACAGTGAAGGAGGAGATGCAATAAATTTGAAAACTAACTC
su_Di Genova AAAG-AAAAAAAAAAGAACAGTGAAGGAGGAGATGCAATAAATTTGAAAACTAACTC
ch2         AAAGAAAAAAAAAAGAACAGTGAAGGAGGAGATGCAATAAATTTGAAAACTAACTC
VIT_218S0041G01880.2 AAAG-AAAAAAAAAAGAACAGTGAAGGAGGAGATGCAATAAATTTGAAAACTAACTC
su_wt      AAAG-AAAAAAAAAAGAACAGTGAAGGAGGAGATGCAATAAATTTGAAAACTAACTC
ch1        AAAG-AAAAAAAAAAGAACAGTGAAGGAGGAGATGCAATAAATTTGAAAACTAACTC
****

su_mut      ACTGCAGATTATGATTTTTTACGAGGGGAGTCAGATTCCTTTTGCTTTATTTAGGAAGGG
su_Di Genova ACTGCAGATTATGATTTTTTAYGAGGGGAGTCAGATTCCTTTTGCTTTATTTAGGAAGGG
ch2         ACTGCAGATTATGATTTTTTATGAGGGGAGTCAGATTCCTTTTGCTTTATTTAGGAAGGG
VIT_218S0041G01880.2 ACTGCAGATTATGATTTTTTATGAGGGGAGTCAGATTCCTTTTGCTTTATTTAGGAAGGG
su_wt      ACTGCAGATTATGATTTTTTATGAGGGGAGTCAGATTCCTTTTGCTTTATTTAGGAAGGG
ch1        ACTGCAGATTATGATTTTTTATGAGGGGAGTCAGATTCCTTTTGCTTTATTTAGGAAGGG

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su_mut      ATTACATGGGTACCTGCTTTGATATTTATGAT-ATTTCTTTTCATCAAAATTTAATTAAT
su_Di Genova ATTACATGGGTACCTGCTTTGATATTTATGAT-ATTTCTTTTCATCAAAATTTAATTAAT
ch2         ATTACATGGGTACCTGCTTTGATATTTATGAT-ATTTCTTTTCATCAAAATTTAATTAAT
VIT_218S0041G01880.2 ATTACATGGGTACCTGCTTTGATATTTATGATAATTTCTTTTATCAAAATTTA---AAT
su_wt       ATTACATGGGTACCTGCTTTGATATTTATGATAATTTCTTTTATCAAAATTTA---AAT
ch1         ATTACATGGGTACCTGCTTTGATATTTATGATAATTTCTTTTATCAAAATTTA---AAT
*****

su_mut      GAAGGAAAATGCAGATAAATATTCTGACATTTTCATCAGGCATCTGCTGATGAATCTGAGAA
su_Di Genova GAAGGAAAATGCAGATAAATATTCTGACATTTTCATCTGGCATCTGCTGATGAATCTGAGAA
ch2         GAAGGAAAATGCAGATAAATATTCTGACATTTTCATCAGGCATCTGCTGATGAATCTGAGAA
VIT_218S0041G01880.2 GAAGGAAAATGCAGATAAATATTCTGACATTTTCATCTGGCATCTGCTGATGAATCTGAGAA
su_wt       GAAGGAAAATGCAGATAAATATTCTGACATTTTCATCTGGCATCTGCTGATGAATCTGAGAA
ch1         GAAGGAAAATGCAGATAAATATTCTGACATTTTCATCTGGCATCTGCTGATGAATCTGAGAA
*****

su_mut      AACAAATGTCTTCTTCAAGATTTTTCTACTAATTACATCTTTGACTGATATAAGGAAAATAT
su_Di Genova AACAAATGTCTTCTTCAAGATTTTTCTACTAATTACATCTTTGACTGATATAAGGAAAATAT
ch2         AACAAATGTCTTCTTCAAGATTTTTCTACTAATTACATCTTTGACTGATATAAGGAAAATAT
VIT_218S0041G01880.2 AACAAATGTCTTCTTCAAGATTTTTCTACTAATTACATCTTTGACTGATATAAGGAAAATAT
su_wt       AACAAATGTCTTCTTCAAGATTTTTCTACTAATTACATCTTTGACTGATATAAGGAAAATAT
ch1         AACAAATGTCTTCTTCAAGATTTTTCTACTAATTACATCTTTGACTGATATAAGGAAAATAT
*****

su_mut      GAGAAAGAACACTATAACCTATAGTTAGAAACTTCTTCAAAGATTAACATATCAGAGGGTTC
su_Di Genova SAGAAAGAACACTATAACCTATAGTTAGAAACTTCTTCAAAGATTAACATATCAGAGGGTTC
ch2         CAGAAAGAACACTTTACCTATAGTTAGAAACTTCTTCAAAGATTAACATATCAGAGGGTTC
VIT_218S0041G01880.2 CAGAAAGAACACTATAACCTATAGTTAGAAACTTCTTCAAAGATTAACATATCAGAGGGTTC
su_wt       CAGAAAGAACACTATAACCTATAGTTAGAAACTTCTTCAAAGATTAACATATCAGAGGGTTC
ch1         CAGAAAGAACACTATAACCTATAGTTAGAAACTTCTTCAAAGATTAACATATCAGAGGGTTC
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su_mut      TTTTATTGTATGTTTGTAGTGCATTAATAA-----AAAAATTATAATGAAAAATAAAAAATA
su_Di Genova TTTTATTGTATGTTTGTAGTGCATTAATAAATAAAAAATAAAAAATTATAATGAAAAATAAAAAATA
ch2         TTTTATTGTATGTTTGTAGTGCATTAATAAATAAAAAATAAAAAATTATAATGAAAAATAAAAAATA
VIT_218S0041G01880.2 TTTTATTGTATGTTTGTAGTGCATTAATAAATAAAAAATAAAAAATTATAATGAAAAATAAAAAATA
su_wt       TTTTATTGTATGTTTGTAGTGCATTAATAAATAAAAAATAAAAAATTATAATGAAAAATAAAAAATA
ch1         TTTTATTGTATGTTTGTAGTGCATTAATAAATAAAAAATAAAAAATTATAATGAAAAATAAAAAATA
*****

su_mut      AAATA-GCAAAAAAACCAGAAAAGTTGAAATCCAACATATCAGTTACACTGGTAGTACTGA
su_Di Genova AAATAGGAAAAAATAACGAAAAAGTTGAAATCCAACATATCAGTTACACTGGTAGTACTGA
ch2         AAATAGGAAAAAATAACGAAAAAGTTGAAATCCAACATATCAGTTACACTGGTAGTACTGA
VIT_218S0041G01880.2 AAATAGGAAAAAATAACGAAAAAGTTGAAATCCAACATATCAGTTACACTGGTAGTACTGA
su_wt       AAATAGGAAAAAATAACGAAAAAGTTGAAATCCAACATATCAGTTACACTGGTAGTACTGA
ch1         AAATAGGAAAAAATAACGAAAAAGTTGAAATCCAACATATCAGTTACACTGGTAGTACTGA
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su_mut      TATGCATGCAAGGGAAGACAATCTAGGCAACAGCCAGTTAGGGTTTCTTGTCTAGCTAG
su_Di Genova TATGCATGCAAGGGAAGACAATCTAGGCAACARCCAGTTAGGGTTTCTTGTCTAGCTAG
ch2         TATGCATGCAAGGGAAGACAATCTAGGCAACAGCCAGTTAGGGTTTCTTGTCTAGCTAG
VIT_218S0041G01880.2 TATGCATGCAAGGGAAGACAATCTAGGCAACAACCAGTTAGGGTTTCTTGTCTAGCTAG
su_wt       TATGCATGCAAGGGAAGACAATCTAGGCAACAACCAGTTAGGGTTTCTTGTCTAGCTAG
ch1         TATGCATGCAAGGGAAGACAATCTAGGCAACAACCAGTTAGGGTTTCTTGTCTAGCTAG
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su_mut      TTCTTCCCATCTCTCCTTACTGTTCCCTTTCCAATAATATTTAACATTGTAATCTGTGAGA
su_Di Genova TTCTTCCCATCTCTCCTTACTGTTCCCTTTCCAATAATATTTAACATTGTAATCTGTGAGA
ch2         TTCTTCCCATCTCTCCTTACTGTTCCCTTTCCAATAATATTTAACATTGTAATCTGTGAGA
VIT_218S0041G01880.2 TTCTTCCCATCTCTCCTTACTGTTCCCTTTCCAATAATATTTAACATTGTAATCTGTGAGA
su_wt       TTCTTCCCATCTCTCCTTACTGTTCCCTTTCCAATAATATTTAACATTGTAATCTGTGAGA
ch1         TTCTTCCCATCTCTCCTTACTGTTCCCTTTCCAATAATATTTAACATTGTAATCTGTGAGA
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su_mut      ATCTGTATTCCACTGTCCACAGTACAGTATTTAAGAAAAGGGTTATGGCCCTTTTCCTCT
su_Di Genova ATCTGTATTCCACTGTCCACAGTACAGTATTTAAGAAAAGGGTTATGGCCCTTTTCCTCT
ch2         ATCTGTATTCCACTGTCCACAGTACAGTATTTAAGAAAAGGGTTATGGCCCTTTTCCTCT

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VIT_218S0041G01880.2 ATCTGTATTCCACTGTCCACAGTACAGTATTTAAGAAAAGGGTTATGGCCCTTTTCCTCT
su_wt ATCTGTATTCCACTGTCCACAGTACAGTATTTAAGAAAAGGGTTATGGCCCTTTTCCTCT
ch1 ATCTGTATTCCACTGTCCACAGTACAGTATTTAAGAAAAGGGTTATGGCCCTTTTCCTCT
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su_mut TCCACTCTTTGTTCAAGTCTCAGTTTCTCTGCTCTTCTCTTTCCATTTCAGCTTTGGGA
su_Di Genova TCCACTCTTTGTTCAAGTCTCAGTTTCTCTGCTCTTCTCTTTCCATTTCAGCTTTGGGA
ch2 TCCACTCTTTGTTCAAGTCTCAGTTTCTCTGCTCTTCTCTTTCCATTTCAGCTTTGGGA
VIT_218S0041G01880.2 TCCACTCTTTGTTCAAGTCTCAGTTTCTCTGCTCTTCTCTTTCCATTTCAGCTTTGGGA
su_wt TCCACTCTTTGTTCAAGTCTCAGTTTCTCTGCTCTTCTCTTTCCATTTCAGCTTTGGGA
ch1 TCCACTCTTTGTTCAAGTCTCAGTTTCTCTGCTCTTCTCTTTCCATTTCAGCTTTGGGA
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su_mut GGCTTGAGGCCATGTATCATAAATCCTTCCCTTGTTTTTCTCCATCTTTTTGTTTTGGG
su_Di Genova GGCTTGAGGCCATGTATCATAAATCCTTCCCTTGTTTTTCTCCATCTTTTTGTTTTGGG
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VIT_218S0041G01880.2 GGCTTGAGGCCATGTATCATAAATCCTTCCCTTGTTTTTCTCCATCTTTTTGTTTTGGG
su_wt GGCTTGAGGCCATGTATCATAAATCCTTCCCTTGTTTTTCTCCATCTTTTTGTTTTGGG
ch1 GGCTTGAGGCCATGTATCATAAATCCTTCCCTTGTTTTTCTCCATCTTTTTGTTTTGGG
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su_mut TTTCTTCAGCCTAAAAGCTGTGAACCTTTCAATGGAGTCTTTTGGTCTCTCTTCTCTTT
su_Di Genova TTTCTTCAGCCTAAAAGCTGTGAACCTTTCAATGGAGTCTTTTGGTCTCTCTTCTCTTT
ch2 TTTCTTCAGCCTAAAAGCTGTGAACCTTTCAATGGAGTCTTTTGGTCTCTCTTCTCTTT
VIT_218S0041G01880.2 TTTCTTCAGCCTAAAAGCTGTGAACCTTTCAATGGAGTCTTTTGGTCTCTCTTCTCTTT
su_wt TTTCTTCAGCCTAAAAGCTGTGAACCTTTCAATGGAGTCTTTTGGTCTCTCTTCTCTTT
ch1 TTTCTTCAGCCTAAAAGCTGTGAACCTTTCAATGGAGTCTTTTGGTCTCTCTTCTCTTT
*****

su_mut CCCATTCTCATTAAAGTGCAGTAAAGTGATCACCTTTCAGTCTCTTTGTTTTTTTCAC
su_Di Genova CCCATTCTCATTAAAGTGCAGTAAAGTGATCACCTTTCAGTCTCTTTGTTTTTTTCAC
ch2 CCCATTCTCATTAAAGTGCAGTAAAGTGATCACCTTTCAGTCTCTTTGTTTTTTTCAC
VIT_218S0041G01880.2 CCCATTCTCATTAAAGTGCAGTAAAGTGATCACCTTTCAGTCTCTTTGTTTTTTTCAC
su_wt CCCATTCTCATTAAAGTGCAGTAAAGTGATCACCTTTCAGTCTCTTTGTTTTTTTCAC
ch1 CCCATTCTCATTAAAGTGCAGTAAAGTGATCACCTTTCAGTCTCTTTGTTTTTTTCAC
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su_mut TTGGTGTGGGCAATTTGTGGATATCAAA--TCAGAAATGGGACGGTGTACATCAGCACC
su_Di Genova TTGGTGTGGGCAATTTGTGGATATCAAA--TCAGAAATGGGACGGTGTACATCAGCACC
ch2 TTGGTGTGGGCAATTTGTGGATATCAAA--TCAGAAATGGGACGGTGTACATCAGCACC
VIT_218S0041G01880.2 TTGGTGTGGGCAATTTGTGGATATCAAAATCTCAGAAATGGGACGGTGTACATCAGCACC
su_wt TTGGTGTGGGCAATTTGTGGATATCAAAATCTCAGAAATGGGACGGTGTACATCAGCACC
ch1 TTGGTGTGGGCAATTTGTGGATATCAAAATCTCAGAAATGGGACGGTGTACATCAGCACC
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su_mut ATTACATCTTTGTGTGGGTTTGAATAAATATTTTCATATCAATCATTTTCATGTTTGTCTCAA
su_Di Genova ATTACATCTTTGTGTGGGTTTGAATAAATATTTTCATATCAATCATTTTCATGTTTGMTCAA
ch2 ATTACATCTTTGTGTGGGTTTGAATAAATATTTTCATATCAATCATTTTCATGTTTGTCTCAA
VIT_218S0041G01880.2 ATTACATCTTTGTGTGGGTTTGAATAAATATTTTCATATCAATCATTTTCATGTTTGTCTCAA
su_wt ATTACATCTTTGTGTGGGTTTGAATAAATATTTTCATATCAATCATTTTCATGTTTGTCTCAA
ch1 ATTACATCTTTGTGTGGGTTTGAATAAATATTTTCATATCAATCATTTTCATGTTTGTCTCAA
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su_mut TGCCATCCAAGATTTTTTTTCTTCACTTCATGTCTCATTTTCTCCCGATTTCCTCCCTT
su_Di Genova TGCCATCCAAGATTTTTTTTCTTCACTTCATGTCTCATTTTCTCCCGATTTCCTCCCTT
ch2 TGCCATCCAAGATTTTTTTTCTTCACTTCATGTCTCATTTTCTCCCTGATTTCCTCCCTT
VIT_218S0041G01880.2 TGCCATCCAAGATTTTTTTTCTTCACTTCATGTCTCATTTTCTCCCTGATTTCCTCCCTT
su_wt TGCCATCCAAGATTTTTTTTCTTCACTTCATGTCTCATTTTCTCCCTGATTTCCTCCCTT
ch1 TGCCATCCAAGATTTTTTTTCTTCACTTCATGTCTCATTTTCTCCCTGATTTCCTCCCTT
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su_mut GTTCAAGGAAAATTGAAGATCTTCTCATAGTTGAGATAGTATTTAAAATAGGACATTGAT
su_Di Genova GTTCAAGGAAAATTGAAGATCTTCTCATAGTTGAGATAGTATTTAAAATAGGACATTGAT
ch2 GTTCAAGGAAAATTGAAGATCTTCTCATAGTTGAGATAGTATTTAAAATAGGACATTGAT
VIT_218S0041G01880.2 GTTCAAGGAAAATTGAAGATCTTCTCATAGTTGAGATAGTATTTAAAATAGGACATTGAT
su_wt GTTCAAGGAAAATTGAAGATCTTCTCATAGTTGAGATAGTATTTAAAATAGGACATTGAT
ch1 GTTCAAGGAAAATTGAAGATCTTCTCATAGTTGAGATAGTATTTAAAATAGGACATTGAT
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su_mut          CTTACATGAGATTTTACTTTTTGTTTTCAAATTTTAAATTATTTTTTATACTTTTTTAA
su_Di Genova   CTTACATGAGATTTTACTTTTTGTTTTCAAATTTTAAATTATTTTTTATACTTTTTTAA
ch2            CTTACATGAGATTTTACCTTTTTTTTTCAAATTTTAAATTATTTTTTATACTTTTTTAA
VIT_218S0041G01880.2 CTTACATGAGATTTTACCTTTTTTTTTCAAATTTTAAATTATTTTTTATACTTTTTTAA
su_wt          CTTACATGAGATTTTACCTTTTTTTTTCAAATTTTAAATTATTTTTTATACTTTTTTAA
ch1            CTTACATGAGATTTTACCTTTTTTTTTCAAATTTTAAATTATTTTTTATACTTTTTTAA
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su_mut          AAATACTTGTTTTTACCCTTAATCAATTAACAAACAAAAAATAAAATGAAAAAAAAGAC
su_Di Genova   AAATACTTGTTTTTACCCTTAATCAATTAACAAACAAAAAATAAAATGAAAAAAAAGAC
ch2            AAATACTTGTTTTTACCCTTAATCAATTAACAAACAAAAAATGA-AAAATG-AAAAAAAAGAC
VIT_218S0041G01880.2 AAATACTTGTTTTTACCCTTAATCAATTAACAAACAAAAAATGA-AAAATG-AAAAAAAAGAC
su_wt          AAATACTTGTTTTTACCCTTAATCAATTAACAAACAAAAAATGA-AAAATG-AAAAAAAAGAC
ch1            AAATACTTGTTTTTACCCTTAATCAATTAACAAACAAAAAATGA-AAAATG-AAAAAAAAGAC
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su_mut          CAATTTGAATTTTTTTTTTAATGTTGAGGGTGCA-----TGGGGGTGGGGGGG
su_Di Genova   CAATTTGAATTTTTTTTTTAATGTTGAGGGTGCA-----TGGGGGTGGGGGGG
ch2            CAATTTGAATTTTTTTTTTAATGTTGAGGGTGCA-----TGGGGGTGGGGGGTGGAGGGG
VIT_218S0041G01880.2 CAATTTGAATTTTTTTTTTAATGTTGAGGGTGCATGGGGGTGGGGGTGGGGGTGGAGGGG
su_wt          CAATTTGAATTTTTTTTTTAATGTTGAGGGTGCA-----TGGGGGTGGGGGTGGAGGGG
ch1            CAATTTGAATTTTTTTTTTAATGTTGAGGGTGCA-----TGGGGGTGGGGGTGGAGGGG
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su_mut          AGAAGTCTTCCTTATTATTTATTAATTTAATGCATTATTTTTAGTCAAATCCAATTTTA
su_Di Genova   AGAAGTCTTCCTTATTATTTATTAATTTAATGCATTATTTTTAGTCAAATCCAATTTWA
ch2            AGAAGTCTTCCTTATTATTTATTAATTTAATGCATTATTTTTAGTCAAATCCAATTTATA
VIT_218S0041G01880.2 AGAAGTCTTCCTTATTATTTATTAATTTAATGCATTATTTTTAGTCAAATCCAATTTATA
su_wt          AGAAGTCTTCCTTATTATTTATTAATTTAATGCATTATTTTTAGTCAAATCCAATTTATA
ch1            AGAAGTCTTCCTTATTATTTATTAATTTAATGCATTATTTTTAGTCAAATCCAATTTATA
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su_mut          TTGCATAAAAATTAATTAACATCATCTTTGATCATCTTTAAATCATATGGATTAAATCGTA
su_Di Genova   TTGCATAAAAATTAATTAACATCATCTTTGATCATCTTTAAATCATATGGATTAAATCGTA
ch2            TTGCATAAAAATTAATTAACATCATCTTTGATCATCTTTAAATCATATGGATTAAATCGTA
VIT_218S0041G01880.2 TTGCATAAAAATTAATTAACATCATCTTTGATCATCTTTAAATCATATGGATTAAATCGTA
su_wt          TTGCATAAAAATTAATTAACATCATCTTTGATCATCTTTAAATCATATGGATTAAATCGTA
ch1            TTGCATAAAAATTAATTAACATCATCTTTGATCATCTTTAAATCATATGGATTAAATCGTA
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su_mut          TTGTTGCTTATTTTCCTTATTCAAATAGAATAAAAAATTTTTAAGATTCTTTATTCCTTT
su_Di Genova   TTGTTGCTTATTTTCCTTATTCAAATAGAATAAAAAATTTTTAAGATTCTTTATTCCTTT
ch2            TTGTTGCTTATTTTCCTTATTCAAATAGAATAAAAAATTTTTAAGATTCTTTATTCCTTT
VIT_218S0041G01880.2 TTGTTGCTTATTTTCCTTATTCAAATAGAATAAAAAATTTTTAAGATTCTTTATTCCTTT
su_wt          TTGTTGCTTATTTTCCTTATTCAAATAGAATAAAAAATTTTTAAGATTCTTTATTCCTTT
ch1            TTGTTGCTTATTTTCCTTATTCAAATAGAATAAAAAATTTTTAAGATTCTTTATTCCTTT
*****

su_mut          ATATATGAGAGAAAAATAAAAGAAATAAATTTGATAAAAATATGTGAAAAAATTTAAGT
su_Di Genova   ATATATGAGAGAAAAATAAAAGAAATAAATTTGATAAAAATATGTGAAAAAATTTAAGT
ch2            ATATATGAGAGAAAAATAAAAGAAATAAATTTGATAAAAATATGTGAAAAAATTTAAGT
VIT_218S0041G01880.2 ATATATGAGAGAAAAATAGAAGAAATAAATTTGATAAAAATATGTGAAAAAATTTAAGT
su_wt          ATATATGAGAGAAAAATAAAAGAAATAAATTTGATAAAAATATGTGAAAAAATTTAAGT
ch1            ATATATGAGAGAAAAATAAAAGAAATAAATTTGATAAAAATATGTGAAAAAATTTAAGT
*****

su_mut          TCAAGCTTTTTTATTTATTTTTATTTTTTTCATCATTTTTTTTTTGTTCTTTTTTTTTC
su_Di Genova   TCAAGCTTTTTTATTTATTTTTATTTTTTTCATCATTTTTTTTTTGTTCTTTTTTTTTC
ch2            TCAAGCTTTTTTATTTATTTTTATTTTTTTCATCA-TTTTTTTTTTGTTCTTTTTTTTTC
VIT_218S0041G01880.2 TCAAGCTTTTTTATTTATTTTTA-CTTTTCATCA-TTTTTTTTTTGTTCTTTTTTTTTC
su_wt          TCAAGCTTTTTTATTTATTTTTATTTTTTTCATCA-TTTTTTTTTTGTTCTTTTTTTTTC
ch1            TCAAGCTTTTTTATTTATTTTTATTTTTTTCATCA-TTTTTTTTTTGTTCTTTTTTTTTC
*****

su_mut          TCTCTCTAAATTTTTGAAGATTTAAATGCAACCCCTACACTTTACACCTAAGAGAGTTT
su_Di Genova   TCTCTCTAAATTTTTGAAGATTTAAATGCAACCCCTACACTTTACACCTAAGAGAGTTT
ch2            TCTCTCTAAATTTTTGAAGATTTAAATGCAACCCCTACACTTTACACCTAAGAGAGTTT
VIT_218S0041G01880.2 TCTCTCTAAATTTTTCGAAGATTTAAATGCAACCCCTACACTTTACACCTAAGAGAGTTT
su_wt          TCTCTCTAAATTTTTGAAGATTTAAATGCAACCCCTACACTTTACACCTAAGAGAGTTT
ch1            TCTCTCTAAATTTTTGAAGATTTAAATGCAACCCCTACACTTTACACCTAAGAGAGTTT

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su_mut      GAGGAAAAGTAGGAGAAAAAGAAAACAAAGAAGAAAAATGAAAGAAAAGAAAAAGAAAAAT
su_Di Genova GAGGAAAAGTAGGAGAAAAAGAAAACAAAGAAGAAAAATGAAAGAAAAGAAAAAGAAAAAT
ch2         GAGGAAAAGTAGGAGAAAAAGAAAACAAAGAAGAAAAATGAAAGAAAAGAAAAAGAAAAAT
VIT_218S0041G01880.2 GAGGAAAAGTAGGAGAAAAAGAAAACAAAGAAGAAAAATGAAAGAAAAGAAAAAGAAAAAT
su_wt      GAGGAAAAGTAGGAGAAAAAGAAAACAAAGAAGAAAAATGAAAGAAAAGAAAAAGAAAAAT
ch1        GAGGAAAAGTAGGAGAAAAAGAAAACAAAGAAGAAAAATGAAAGAAAAGAAAAAGAAAAAT
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su_mut      AAAGAAATATGTTTAAATTCAATAAATGTTATTACTTATTTTTCTAAAATTATTTTTAT
su_Di Genova AAAGAAATATGTTTAAATTCAATAAATGTTATTACTTATTTTTCTAAAATTATTTTTAT
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VIT_218S0041G01880.2 AAAGAAATATGTTTAAATTCAATAAATGTTATTACTTATTTTTCTAAAATTATTTTTAT
su_wt      AAAGAAATATGTTTAAATTCAATAAATGTTATTACTTATTTTTCTAAAATTATTTTTAT
ch1        AAAGAAATATGTTTAAATTCAATAAATGTTATTACTTATTTTTCTAAAATTATTTTTAT
*****

su_mut      TCTTTTCTCTCATATACAACCAAAACAAAATTTCAAAAATGTAAAATTTTTGATATAGTTT
su_Di Genova TCTTTTCTCTCATATACAACCAAAACAAAATTTCAAAAATGTAAAATTTTTGATATAGTTT
ch2         TCTTTTCTCTCATATACAACCAAAACAAAATTTCAAAAATGTAAAATTTTTGATATAGTTT
VIT_218S0041G01880.2 TCTTTTCTCTCATATACAACCAAAACAAAATTTCAAAAATGTAAAATTTTTGATATAGTTT
su_wt      TCTTTTCTCTCATATACAACCAAAACAAAATTTCAAAAATGTAAAATTTTTGATATAGTTT
ch1        TCTTTTCTCTCATATACAACCAAAACAAAATTTCAAAAATGTAAAATTTTTGATATAGTTT
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su_mut      TCATTAGATTTGATTTTTTCATACTTTCCATAATAATCCAACAAGATCAATTAATAAACC
su_Di Genova TCATTAGATTTGATTTTTTCATACTTTCCATAATAATCCAACAAGATCAATTAATAAACY
ch2         TCATTAGATTTGATTTTTTCATACTTTCCATAATAATCCAACAAGATCAATTAATAAACC
VIT_218S0041G01880.2 TCATTAGATTTGATTTTTTCATACTTTCCATAATAATCCAACAAGATCAATTAATAAACC
su_wt      TCATTAGATTTGATTTTTTCATACTTTCCATAATAATCCAACAAGATCAATTAATAAACC
ch1        TCATTAGATTTGATTTTTTCATACTTTCCATAATAATCCAACAAGATCAATTAATAAACC
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su_mut      GAAACAAAACCTACAATGACTTCATAATAACGGAAGTTTAAAGGAACTAAGGATGAATTG
su_Di Genova GAAACAAAACCTACAATGACTTCATAATAACGGAAGTTTAAAGGAACTAAGGATGAATTG
ch2         GAAACAAAACCTACAATGACTTCATAATAACGGAAGTTTAAAGGAACTAAGGATGAATTG
VIT_218S0041G01880.2 GAAACAAAACCTACAATGACTTCATAATAACGGAAGTTTAAAGGAACTAAGGATGAATTG
su_wt      GAAACAAAACCTACAATGACTTCATAATAACGGAAGTTTAAAGGAACTAAGGATGAATTG
ch1        GAAACAAAACCTACAATGACTTCATAATAACGGAAGTTTAAAGGAACTAAGGATGAATTG
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su_mut      AGATAGATTCGAGGATGGTGTGTCATAATGATAGGAGTCCAAATTTGGAGGTAAGAATAAAC
su_Di Genova AGATAGATTCGAGGATGGTGTGTCATAATGATAGGAGTCCAAATTTGGAGGTAAGAATAAAC
ch2         AGATAGATTCGAGGATGGTGTGTCATAATGATAGGAGTCCAAATTTGGAGGTAAGAATAAAC
VIT_218S0041G01880.2 AGATAGATTCGAGGATGGTGTGTCATAATGATAGGAGTCCAAATTTGGAGGTAAGAATAAAC
su_wt      AGATAGATTCGAGGATGGTGTGTCATAATGATAGGAGTCCAAATTTGGAGGTAAGAATAAAC
ch1        AGATAGATTCGAGGATGGTGTGTCATAATGATAGGAGTCCAAATTTGGAGGTAAGAATAAAC
*****

su_mut      TTAATAAACTCTAAGATGATGCTCCAATGATGAATTTCTTTTTTAAAAATATTTGAGTAG
su_Di Genova TTAATAAACTCTAAGATGATGCTCCAATGATGAATTTCTTTTTTAAAAATATTTGAGTAG
ch2         TTAATAAACTCTAAGACGATGCTCCAATGATGAATTTCTTTTTTAAAAATATTTGAGTAG
VIT_218S0041G01880.2 TTAATAAACTCTAAGACGATGCTCCAATGATGAATTTCTTTTTTAAAAATATTTGAGTAG
su_wt      TTAATAAACTCTAAGACGATGCTCCAATGATGAATTTCTTTTTTAAAAATATTTGAGTAG
ch1        TTAATAAACTCTAAGACGATGCTCCAATGATGAATTTCTTTTTTAAAAATATTTGAGTAG
*****

su_mut      ACATTTGTCATTAATTTTTCTAACTACAAATAGATTAACAATAAATGATCATTATCCCT
su_Di Genova GCATTTGTCATTAATTTTTCTAACTACAAATAGATTAACAATAAATGATCATAATCCCT
ch2         GCATTTGTCATTAATTTTTCTAACTACAAATAGATTAACAATAAATGATCATAATCCCT
VIT_218S0041G01880.2 GCATTTGTCATTAATTTTTCTAACTACAAATAGATTAACAATAAATGATCATAATCCCT
su_wt      GCATTTGTCATTAATTTTTCTAACTACAAATAGATTAACAATAAATGATCATAATCCCT
ch1        GCATTTGTCATTAATTTTTCTAACTACAAATAGATTAACAATAAATGATCATAATCCCT
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su_mut      TTCTACAGAACTTGTGCGATTGTTGATGTTTTTTATGATATTTTTATGTAACATGTTAA
su_Di Genova TTCAACCGAACTTGTGCGTTGTTGATGTTTTTTATGATATTTTTATGTAACATGTTAA
ch2         TTCAACCGAACTTGTGCGTTGTTGATGTTTTTTATGATATTTTTATGTAACATGTTAA

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VIT_218S0041G01880.2 TTCAACCGAACTTGTGCGGTTGTTGTATTGTTTTTTATGATATTTTTATGTAACATGTTAA
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ch1 TTCAACCGAACTTGTGCGGTTGTTGTATTGTTTTTTATGATATTTTTATGTAACATGTTAA
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su_mut ATTATAAAATAAATAAATAAGTTATAAGAAGAAAGAAAAACCAATTCAATTCCAATAAA
su_Di Genova ATTATAAAATAAATAAATAAGTTATAAGAAGAAAGAAAAACCAATTCAATTCCAATAAA
ch2 ATTATAAAATAAATAAATAAGTTATAAGAAGAAAGAAAAACCAATTCAATTCCAATAAA
VIT_218S0041G01880.2 ATTATAAAATAAATAAATAAGTTATAAGAAGAAAGAAAAACCAATTCAATTCCAATAAA
su_wt ATTATAAAATAAATAAATAAGTTATAAGAAGAAAGAAAAACCAATTCAATTCCAATAAA
ch1 ATTATAAAATAAATAAATAAGTTATAAGAAGAAAGAAAAACCAATTCAATTCCAATAAA
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su_mut AACCTTTGAAATTAATAAATGGTGGGATATGTG--AGAGGTGCAAAGTCTTGGGTTTGAT
su_Di Genova AACCTTTGAAATTAATAAATGGTGGGATATGTGATAGAGGTGCAAAGTCTTGGGTTTGAT
ch2 AACCTTTGAAATTAATAAATGGTGGGATATGTGATAGAGGTGCAAAGTCTTGGGTTTCAAT
VIT_218S0041G01880.2 AACCTTTGAAATTAATAAATGGTGGGATATGTGATAGAGGTGCAAAGTCTTGGGTTTCAAT
su_wt AACCTTTGAAATTAATAAATGGTGGGATATGTGATAGAGGTGCAAAGTCTTGGGTTTCAAT
ch1 AACCTTTGAAATTAATAAATGGTGGGATATGTGATAGAGGTGCAAAGTCTTGGGTTTCAAT
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su_mut TACCATCATTAAAGAATCCCTGGATTACTCGGAGTTGATTCTAATGGGTGTCGTTGGAATC
su_Di Genova TACCATCATTAAAGAATCCCTGGATTACTCGGAGTTGATTCTAATGGGTGTCGTTGGAATC
ch2 TACCATCATTAAAGAATCCCTGGATTACTCGGAGTTGATTCTAATGGGTGTCGTTGGAATC
VIT_218S0041G01880.2 TACCATCATTAAAGAATCCCTGGATTACTCGGAGTTGATTCTAATGGGTGTCGTTGGAATC
su_wt TACCATCATTAAAGAATCCCTGGATTACTCGGAGTTGATTCTAATGGGTGTCGTTGGAATC
ch1 TACCATCATTAAAGAATCCCTGGATTACTCGGAGTTGATTCTAATGGGTGTCGTTGGAATC
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su_mut CCTAAGGTGTCGAAGCCATGGGTGGATTTGAAAGGCCCTATCACAGTGGGGTTTCCGGGT
su_Di Genova CCTAAGGTGTCGAAGCCATGGGTGGATTTGAAAGGCCCTATCACAGTGGGGTTTCCGGGT
ch2 CCTAAGGTGTCGAAGCCATGGGTGGATTTGAAAGGCCCTATCACAGTGGGGTTTCCGGGT
VIT_218S0041G01880.2 CCTAAGGTGTCGAAGCCATGGGTGGATTTGAAAGGCCCTATCACAGTGGGGTTTCCGGGT
su_wt CCTAAGGTGTCGAAGCCATGGGTGGATTTGAAAGGCCCTATCACAGTGGGGTTTCCGGGT
ch1 CCTAAGGTGTCGAAGCCATGGGTGGATTTGAAAGGCCCTATCACAGTGGGGTTTCCGGGT
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su_mut TATCAAAAAAAAAAACACACACACACAATTTTCATAATAGAAGGTAGAAAAAATTGAG
su_Di Genova TATCAAAAAAAAAAACACACACACACAATTTTCATAATAGAAGGTAGAAAAAATTGAG
ch2 TAT----AAAAAACACACACACACACAATTTTCATAATAGAAGGTAGAAAAAATTGAG
VIT_218S0041G01880.2 TAT--AAAAAACACACACACACACAATTTTCATAATAGAAGGTAGAAAAAATTGAG
su_wt TAT-----AAAAAACACACACACACACAATTTTCATAATAGAAGGTAGAAAAAATTGAG
ch1 TAT--AAAAAACACACACACACACAATTTTCATAATAGAAGGTAGAAAAAATTGAG
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su_mut AATAAGGTAGAGAAGATGTTTTTTCC-----TAGGTACTCTAGGGTTTTGTGATAATAG
su_Di Genova AATAAGGTAGAGAAGATGTTTTTTCC TAGCTATAGGTACTCTAGGGTTTTGTGATAATAG
ch2 AATAAGGTAGAGAAGATGTTTTTTCC TAGCTATAGGTACTCTAGGGTTTTGTGATAATAG
VIT_218S0041G01880.2 AATAAGGTAGAGAAGATGTTTTTTCC TAGCTATAGGTACTCTAGGGTTTTGTGATAATAG
su_wt AATAAGGTAGAGAAGATGTTTTTTCC TAGCTATAGGTACTCTAGGGTTTTGTGATAATAG
ch1 AATAAGGTAGAGAAGATGTTTTTTCC TAGCTATAGGTACTCTAGGGTTTTGTGATAATAG
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su_mut ATTTGGATTTTTTTCCCAATAAATCAAATTATTAATTCAATTATAATAAATAAAAA
su_Di Genova ATTTGGATTTTTTTCCCAATAAATCAAATTATTAATTCAATTATAATAAATAAAAA
ch2 ATTTGGATTTTTTT-CCCAATAAATAAAATTATTAATTCAATTATAATAAATAAAAA
VIT_218S0041G01880.2 ATTTGGATTTTTTT-CCCAATAAATAAAATTATTAATTCAATTATAATAAATAAAAA
su_wt ATTTGGATTTTTTT-CCCAATAAATAAAATTATTAATTCAATTATAATAAATAAAAA
ch1 ATTTGGATTTTTTT-CCCAATAAATAAAATTATTAATTCAATTATAATAAATAAAAA
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su_mut TCAATGAGAAATCCATGGAAACCCAAATCAAATGTTGCAATATAGATCAAGATTAGCA
su_Di Genova TCAATGAGAAATCCATGGAAACCCAAATCAAATGTTGCAATATAGATCAAGATTAGCA
ch2 TCAATGAGAAATCCATGG-AAACCCAAATCAAATGTTGCAATATAGATCAAGATTAGCA
VIT_218S0041G01880.2 TCAATGAGAAATCCATGG-AAACCCAAATCAAATGTTGCAATATAGATCAAGATTAGCA
su_wt TCAATGAGAAATCCATGG-AAACCCAAATCAAATGTTGCAATATAGATCAAGATTAGCA
ch1 TCAATGAGAAATCCATGG-AAACCCAAATCAAATGTTGCAATATAGATCAAGATTAGCA
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su_mut      TTTACTTATATGCATGCATGTTAATTAGTTTGGACCTTAAAGATCTTGTAGTACTGGTTCC
su_Di Genova TTTACTTATATGCATGCATGTTAATTAGTTTGGACCTTAAAGATCTTGTAGTACTGGTTCC
ch2         TTTACTTATATGCATGCATGTTAATTAGTTTGGACCTTAAAGATCTTGTAGTACTGGTTCC
VIT_218S0041G01880.2 TTTACTTATATGCATGCATGTTAATTAGTTTGGACCTTAAAGATCTTGTAGTACTGGTTCC
su_wt      TTTACTTATATGCATGCATGTTAATTAGTTTGGACCTTAAAGATCTTGTAGTACTGGTTCC
ch1         TTTACTTATATGCATGCATGTTAATTAGTTTGGACCTTAAAGATCTTGTAGTACTGGTTCC
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su_mut      CAATATAAAAAATGAGAGCCTATTTGGGATAACTTTTTAACTCCTGCATCTAGTAATTAGA
su_Di Genova CAATATAAAAAATGAGAGCCTATTTGGGATAACTTTTTAACWCCTGCATCTAGTAATTAGA
ch2         CAATATAAAAAATGAGAGCCTATTTGGGATAACTTTTTAACACCTGCATCTAGTAATTAGA
VIT_218S0041G01880.2 CAATATAAAAAATGAGAGCCTATTTGGGATAACTTTTTAACACCTGCATCTAGTAATTAGA
su_wt      CAATATAAAAAATGAGAGCCTATTTGGGATAACTTTTTAACACCTGCATCTAGTAATTAGA
ch1         CAATATAAAAAATGAGAGCCTATTTGGGATAACTTTTTAACACCTGCATCTAGTAATTAGA
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su_mut      GATGAAACAGACATTATTTGTATTATGTAATCTATTTTACGACAAAATTTTAAAAATGC
su_Di Genova GATGAAACAGACATTATTTGTATTATGTAATCTATTTTACGACAAAATTTTAAAAATGC
ch2         GATGAAACATACATTATTTGTATTATGTAATCTATTTTATGACAAAATTTTAAAAATGC
VIT_218S0041G01880.2 GATGAAACATACATTATTTGTATTATGTAATCTATTTTATGACAAAATTTTAAAAATGC
su_wt      GATGAAACATACATTATTTGTATTATGTAATCTATTTTATGACAAAATTTTAAAAATGC
ch1         GATGAAACATACATTATTTGTATTATGTAATCTATTTTATGACAAAATTTTAAAAATGC
*****

su_mut      TTTAATAAAGTAGTGAGGATGACTATGATATTTCAAAAAAATTTAATTAATAAAAAATTT
su_Di Genova TTTAATAAAGTAGTGAGGATGACTATGATATTTCAAAAAAATTTAATTAATAAAAAATTT
ch2         TTTAATAAAGTAGTGAGGATGACTATGATATTTCA-----AAAAATTTT
VIT_218S0041G01880.2 TTTAATAAAGTAGTGAGGATGACTATGATATTTCA-----AAAAATTTT
su_wt      TTTAATAAAGTAGTGAGGATGACTATGATATTTCA-----AAAAATTTT
ch1         TTTAATAAAGTAGTGAGGATGACTATGATATTTCA-----AAAAATTTT
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su_mut      ACTAACAAAAATGTCGATAAATCATTTATAAGTGTTATCTACATGTCCTTAAATCACTT
su_Di Genova ACTAACAAAAATGTCGATAAATCATTTATAAGTGTTATCTAAATGCCTTAAATCACTT
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VIT_218S0041G01880.2 ACTAACAAAAATGTCGATAAATCATTTATAAGTGTTATCTAAATGCCTTAAATCACTT
su_wt      ACTAACAAAAATGTCGATAAATCATTTATAAGTGTTATCTAAATGCCTTAAATCACTT
ch1         ACTAACAAAAATGTCGATAAATCATTTATAAGTGTTATCTAAATGCCTTAAATCACTT
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su_mut      ATTTAAATAATAGAAATTAAGCTAGATTAATAATGAATGAAAAGAAAAAATAAAATTT
su_Di Genova ATTTAAATAATAGAAATTAAGCTAGATTAATAATGAATGAAAAGAAAAAATAAAATTT
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VIT_218S0041G01880.2 ATTTAAATAATAGAAATTAAGCTAGATTAATAATGAATGAAAAGAAAAAATAAAATTT
su_wt      ATTTAAATAATAGAAATTAAGCTAGATTAATAATGAATGAAAAGAAAAAATAAAATTT
ch1         ATTTAAATAATAGAAATTAAGCTAGATTAATAATGAATGAAAAGAAAAAATAAAATTT
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su_mut      GTACAAACCATCCAGTTTAGTACTGGTTCGAACCCCTTCAAGCTCAAAAGGTCATAGG
su_Di Genova GTACAAATCATCCAGTTTAGTACTGGTTCGAACCCCTTCAAGCTCAAAAGGTCATCGG
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VIT_218S0041G01880.2 GTACAAATCATCCAGTTTAGTACTGGTTCGAACCCCTTCAAGCTCAAAAGGTCATCGG
su_wt      GTACAAATCATCCAGTTTAGTACTGGTTCGAACCCCTTCAAGCTCAAAAGGTCATCGG
ch1         GTACAAATCATCCAGTTTAGTACTGGTTCGAACCCCTTCAAGCTCAAAAGGTCATCGG
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su_mut      ATTGGACTAGGATCCGGTCAATAGTTCGATTGATTAATCGATCAATT--GGTCCGATGTT
su_Di Genova ATTGGACTAGGATCTGGTCAACAGTTCGATTGATT----GATCAATTCGGTCCGATCTT
ch2         ATTGGACTAGGATCTGGTCAACAGTTCGATTGATT----GATCAATTCGGTCCGATCTT
VIT_218S0041G01880.2 ATTGGACTAGGATCTGGTCAACAGTTCGATTGATT----GATCAATTCGGTCCGATCTT
su_wt      ATTGGACTAGGATCTGGTCAACAGTTCGATTGATT----GATCAATTCGGTCCGATCTT
ch1         ATTGGACTAGGATCTGGTCAACAGTTCGATTGATT----GATCAATTCGGTCCGATCTT
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su_mut      TAAAACATTACAATAAECTTATCTTATCTGAGTTTTGATGCATCTCATACTGCAAAGTA
su_Di Genova TAAAACATTACAATAAECTTATCTTATGTGAGTTTTGGTGCATCTCATACTGCAAAGCA
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VIT_218S0041G01880.2 TAAAACATTACAATAAECTTATCTTATGTGAGTTTTGGTGCATCTCATACTGCAAAGCA
su_wt      TAAAACATTACAATAAECTTATCTTATGTGAGTTTTGGTGCATCTCATACTGCAAAGCA
ch1         TAAAACATTACAATAAECTTATCTTATGTGAGTTTTGGTGCATCTCATTATGCAAAGCA

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su_mut      GTTATTGAAAGCCAATACATATTTTCATACAAAGTCCACACAAAGAACTCAAATAAAAA
su_Di Genova GTTATTGAAAGCCAATACATATTTTCATACAAAGTCCACACAAAGAACTCAAATAAAAA
ch2         GTTATTGAAAGCCAATACATATTTTCATACAAAGTCCACACAAAGAACTCAAATAAAAA
VIT_218S0041G01880.2 GTTATTGAAAGCCAATACATATTTTCATACAAAGTCCACACAAAGAACTCAAATAAAAA
su_wt      GTTATTGAAAGCCAATACATATTTTCATACAAAGTCCACACAAAGAACTCAAATAAAAA
ch1         GTTATTGAAAGCCAATACATATTTTCATACAAAGTCCACACAAAGAACTCAAATAAAAA
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su_mut      ATAAAAAATCAGAGAATAAAATCCTCATGGTAAAGTTGCACCAATAAGACCTATACTGC
su_Di Genova ATAAAAAATCAGAGAATAAAATCCTCATGGTAAAGTTGCACCAATAAGACCTATACTGC
ch2         ATAAAAAATCAGAGAATAAAATCCTCATGGTAAAGTTGCACCAATAAGACCTATACTGC
VIT_218S0041G01880.2 ATAAAAAATCAGAGAATAAAATCCTCATGGTAAAGTTGCACCAATAAGACCTATACTGC
su_wt      ATAAAAAATCAGAGAATAAAATCCTCATGGTAAAGTTGCACCAATAAGACCTATACTGC
ch1         ATAAAAAATCAGAGAATAAAATCCTCATGGTAAAGTTGCACCAATAAGACCTATACTGC
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su_mut      ATACAGTTGCAGAGTGGCAGTGTCTCCATTGTTCTTTCTCTTGCTTTTTCAGTGGTGCAA
su_Di Genova ATACAGTTGCAGAGTGGCAGTGTCTCYCATTGTTCTTTCTCTTGCTTTTTCAGTGGTGCAA
ch2         ATACAGTTGCAGAGTGGCAGTGTCTTTCATTGTTCTTTCTCTTGCTTTTTCAGTGGTGTA
VIT_218S0041G01880.2 ATACAGTTGCAGAGTGGCAGTGTCTTTCATTGTTCTTTCTCTTGCTTTTTCAGTGGTGTA
su_wt      ATACAGTTGCAGAGTGGCAGTGTCTTTCATTGTTCTTTCTCTTGCTTTTTCAGTGGTGTA
ch1         ATACAGTTGCAGAGTGGCAGTGTCTTTCATTGTTCTTTCTCTTGCTTTTTCAGTGGTGTA
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su_mut      TCCATGAGAATCTGAACCTCTGGCAGTGTCTGAAAAAGGGTGGTCATAGCAGTACAACC
su_Di Genova TCCATGAGAATCTGAACCTCTGGCAGTGTCTGAAAAAGGGTGGTCATAGCAGTACAACC
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VIT_218S0041G01880.2 TCCATGAGAATCTGAACCATCTGGCAGTGTCTGAAAAAGGGAGGTCATAGCAGTACAACC
su_wt      TCCATGAGAATCTGAACCATCTGGCAGTGTCTGAAAAAGGGAGGTCATAGCAGTACAACC
ch1         TCCATGAGAATCTGAACCATCTGGCAGTGTCTGAAAAAGGGAGGTCATAGCAGTACAACC
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su_mut      AACCACCTCATTTTTCATCTTTCCCTTGATCTTCCCTTGTTTGCAAAATCTCAGTTTTTCT
su_Di Genova AACCACCTCATTTTTCATCTTTCCCTTGATCTTCCCTTGTTTGCAAAATCTCAGTTTTTCT
ch2         AACCACCTCATTTTTCATCTTTCCCTTGATCTTCCCTTGTTTGCAAAATCTCAGTTTTTCT
VIT_218S0041G01880.2 AACCACCTCATTTTTCATCTTTCCCTTGATCTTCCCTTGTTTGCAAAATCTCAGTTTTTCT
su_wt      AACCACCTCATTTTTCATCTTTCCCTTGATCTTCCCTTGTTTGCAAAATCTCAGTTTTTCT
ch1         AACCACCTCATTTTTCATCTTTCCCTTGATCTTCCCTTGTTTGCAAAATCTCAGTTTTTCT
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su_mut      GGTGTTTCTTTCCACTTTTGGCTTTCCGCAAACCTTGAATGGAGTCTTGGGCTTTTCT
su_Di Genova GGTGTTTCTTTCCACTTTTGGCTTTCCGCAAACCTTGAATGGAGTCTTGGGCTTTTCT
ch2         GGTGTTTCTTTCCACTTTTGGCTTTCCGCAAACCTTGAATGGAGTCTTGGGCTTTTCT
VIT_218S0041G01880.2 GGTGTTTCTTTCCACTTTTGGCTTTCCGCAAACCTTGAATGGAGTCTTGGGCTTTTCT
su_wt      GGTGTTTCTTTCCACTTTTGGCTTTCCGCAAACCTTGAATGGAGTCTTGGGCTTTTCT
ch1         GGTGTTTCTTTCCACTTTTGGCTTTCCGCAAACCTTGAATGGAGTCTTGGGCTTTTCT
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su_mut      CTTTGCTCTCAAAGCTGCAATCACTGCTCTTTTTTAGTGCTGGAGAGTAGAAACCAAGA
su_Di Genova CTTTGCTCTCAAAGCTGCAATCACTGCTCTTTTTTAGTGCTGGAGAGTAGAAACCAAGA
ch2         CTTTGCTCTCAAAGCTGCAATCACTGCTCTTTTTTAGTGCTGGAGAGGAGAAACCAAGA
VIT_218S0041G01880.2 CTTTGCTCTCAAAGCTGCAATCACTGCTCTTTTTTAGTGCTGGAGAGGAGAAACCAAGA
su_wt      CTTTGCTCTCAAAGCTGCAATCACTGCTCTTTTTTAGTGCTGGAGAGGAGAAACCAAGA
ch1         CTTTGCTCTCAAAGCTGCAATCACTGCTCTTTTTTAGTGCTGGAGAGGAGAAACCAAGA
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su_mut      CAAGACTTTTCAACTGCTATTCTCCATACAAACTCATGAACTGATGAACAATTGAGCAG
su_Di Genova CAAGACTTTTCAACTGCTATTCTCCATACAAACTCATGAACTGATGAACAATTGAGCAG
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VIT_218S0041G01880.2 CAAGACTTTTCAACTGCTATTCTCCATACAAACTCATGAACTGATGAACAATTGAGCAG
su_wt      CAAGACTTTTCAACTGCTATTCTCCATACAAACTCATGAACTGATGAACAATTGAGCAG
ch1         CAAGACTTTTCAACTGCTATTCTCCATACAAACTCATGAACTGATGAACAATTGAGCAG
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su_mut TAGGGTCACTGTGCATACTGTATAGTGATTTTAAT CATTTTTATGTCTCATAACTTGTGG
su_Di Genova TAGGGTCACTGTGYATACTGTATAGTGATTTTAAT CATTTTTATGTCTCATAACTTGTGG
ch2 TAGGGTCACTGTGTATACTGTATAGTGATTTTAAT CATTTTTATGTCTCATAACTTGTGG
VIT_218S0041G01880.2 TAGGGTCACTGTGTATACTGTATAGTGATTTTAAT CATTTTTATGTCTCATAACTTGTGG
su_wt TAGGGTCACTGTGTATACTGTATAGTGATTTTAAT CATTTTTATGTCTCATAACTTGTGG
ch1 TAGGGTCACTGTGTATACTGTATAGTGATTTTAAT CATTTTTATGTCTCATAACTTGTGG
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su_mut GTGTTTGTACTGTTACTTTCATCTTTTCTCATATCAAGTCATAGTCCTTTCACAAACTTGTG
su_Di Genova GTGTTTGTACTGTTACTTTCATCTTTTCTCATATCAAGTCATAGTCCTTTCACAAACTTGTG
ch2 GTGTTTGTACTGTTACTTTCATCTTTTCTCATATCAAGTCATAGTCCTTTCACAAACTTGTG
VIT_218S0041G01880.2 GTGTTTGTACTGTTACTTTCATCTTTTCTCATATCAAGTCATAGTCCTTTCACAAACTTGTG
su_wt GTGTTTGTACTGTTACTTTCATCTTTTCTCATATCAAGTCATAGTCCTTTCACAAACTTGTG
ch1 GTGTTTGTACTGTTACTTTCATCTTTTCTCATATCAAGTCATAGTCCTTTCACAAACTTGTG
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su_mut CTGTCATCTCCCATCATATATGGTTTCTCCTTTAGGGTTTTGCTTTCCGCCTTCACTTT
su_Di Genova CTGTCATCTCCCATCATATATGGTTTCTCCTTTAGGGTTTTGCTTTCCGCCTTCACTTT
ch2 CTGTCATCTCCCATCATATATGGTTTCTCCTTTAGGGTTTTGCTTTCCGCCTTCACTTT
VIT_218S0041G01880.2 CTGTCATCTCCCATCATATATGGTTTCTCCTTTAGGGTTTTGCTTTCCGCCTTCACTTT
su_wt CTGTCATCTCCCATCATATATGGTTTCTCCTTTAGGGTTTTGCTTTCCGCCTTCACTTT
ch1 CTGTCATCTCCCATCATATATGGTTTCTCCTTTAGGGTTTTGCTTTCCGCCTTCACTTT
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su_mut TGACTACTTTTGGAGCAATTTTCTCTCTTGAATTCTATCATATGTAATATTTCTATCCAT
su_Di Genova TGACTACTTTTGGAGCAATTTTCTCTCTTGAATTCTATCATATGTAATATTTCTATCCAT
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su_wt TGACTACTTTTGGAGCAATTTTCTCTCTTGAATTCTATGGTATGTAATATTTCAATCCAT
ch1 TGACTACTTTTGGAGCAATTTTCTCTCTTGAATTCTATGGTATGTAATATTTCAATCCAT
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su_mut TTTGCAACAATTTTTTTAATTCATTCATTTTTATATATTGGAAAACAG CATAAAATCAA
su_Di Genova TTTGCAACAATTTTTTTAATTCATTCATTTTTATATATTGGAAAACAG CATAAAATCAA
ch2 TTTGCAACAATTTTTTTAATTCATTCATTTTTATATATTGGAAAACAG CATAAAATCAA
VIT_218S0041G01880.2 TTTGCAACAATTTTTTTAATTCATTCATTTTTATATATTGGAAAACAG CATAAAATCAA
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ch1 TTTGCAACAATTTTTTTAATTCATTCATTTTTATATATTGGAAAACAG CATAAAATCAA
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su_mut CCATAGATAGGTACAAGAAGGCCAGCTCAGATAGTACAAATGGAGGCTCTACCATGGAGA
su_Di Genova CCATAGATAGGTACAAGAAGGCCAGCTCAGATAGTACAAATGGAGGCTCTACCATGGAGA
ch2 CCATAGATAGGTACAAGAAGGCCAGCTCAGATAGTACAAATGGAGGCTCTACCATGGAGA
VIT_218S0041G01880.2 CCATAGATAGGTACAAGAAGGCCAGCTCAGATAGTACAAATGGAGGCTCTACCATGGAGA
su_wt CCATAGATAGGTACAAGAAGGCCAGCTCAGATAGTACAAATGGAGGCTCTACCATGGAGA
ch1 CCATAGATAGGTACAAGAAGGCCAGCTCAGATAGTACAAATGGAGGCTCTACCATGGAGA
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su_mut TCAATGCCCAAGTAAGAAAACCTCCATTTTATGATAGTGAATGAATCAATCCAAATCCTA
su_Di Genova TCAATGCCCAAGTAAGAAAACCTCCATTTTATGATAGTGAATGAATCAATCCAAATCCTA
ch2 TCAATGCCCAAGTAAGAAAACCTCCATTTTATGATAGTGAATGAATCAATCCAAATCCTA
VIT_218S0041G01880.2 TCAATGCCCAAGTAAGAAAACCTCCATTTTATGATAGTGAATGAATCAATCCAAATCCTA
su_wt TCAATGCCCAAGTAAGAAAACCTCCATTTTATGATAGTGAATGAATCAATCCAAATCCTA
ch1 TCAATGCCCAAGTAAGAAAACCTCCATTTTATGATAGTGAATGAATCAATCCAAATCCTA
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su_mut TATGGTTTACTGATCAGATGTCTTATCCACCATGGCTGTTAGAACATCTCAATCCATTCT
su_Di Genova TATGGTTTACTGATCAGATGTCTTATCCACCATGGCTGTTAGAACATCTCAATCCATTCT
ch2 TATGGTTTACTGATCAGATGTCTTATCCACCATGGCTGTTAGAACATCTCAATCCATTCT
VIT_218S0041G01880.2 TATGGTTTACTGATCAGATGTCTTATCCACCATGGCTGTTAGAACATCTCAATCCATTCT
su_wt TATGGTTTACTGATCAGATGTCTTATCCACCATGGCTGTTAGAACATCTCAATCCATTCT
ch1 TATGGTTTACTGATCAGATGTCTTATCCACCATGGCTGTTAGAACATCTCAATCCATTCT
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su_mut GTGGATTGAAAAGTGACACATTTTTTCATTAGGCTGGCCACAAATTAACCCCTCATCATGC
su_Di Genova GTGGATTGAAAAGTGACACATTTTTTCATTAGGCTGGCCACAAATTAACCCCTCATCATGC
ch2 GTGGATTGAAAAGTGACACATTTTTTCATTAGGCTGGCCACAAATTAACCCCTCATCATGC
VIT_218S0041G01880.2 GTGGATTGAAAAGTGACACATTTTTTCATTAGGCTGGCCACAAATTAACCCCTCATCATGC
su_wt GTGGATTGAAAAGTGACACATTTTTTCATTAGGCTGGCCACAAATTAACCCCTCATCATGC
ch1 GTGGATTGAAAAGTGACACATTTTTTCATTAGGCTGGCCACAAATTAACCCCTCATCATGC

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su_mut      TTAGCCATAAACAAATTTGTTAAAATCATTATCTACTTTCTTTTTCTTTTTGGATTAT
su_Di Genova TTAGCCATAAACAAATATGTTAAAATCATTATCCTACTTTCTTTTTCTTTTTGGATTAT
ch2         TTAGCCATAAACAAATATGTTAAAATCATTATCCTACTTTCTTTTTCTTTTTGGATTAT
VIT_218S0041G01880.2 TTAGCCATAAACAAATATGTTAAAATCATTATCCTACTTTCTTTTTCTTTTTGGATTAT
su_wt      TTAGCCATAAACAAATATGTTAAAATCATTATCCTACTTTCTTTTTCTTTTTGGATTAT
ch1         TTAGCCATAAACAAATATGTTAAAATCATTATCCTACTTTCTTTTTCTTTTTGGATTAT
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su_mut      CCCATTGAACTTTCTCGAATTTCTGTCGTTGAACATAGTATTACCAGCAAGAATCAGCAAA
su_Di Genova CCCATTGAACTTTCTCGAATTTCTGTTGTTGAACATAGTACTACCAGCAAGAATCAGCAAA
ch2         CCCATTGAACTTTCTCGAATTTCTGTTGTTGAACATAGTACTACCAGCAAGAATCAGCAAA
VIT_218S0041G01880.2 CCCATTGAACTTTCTCGAATTTCTGTTGTTGAACATAGTACTACCAGCAAGAATCAGCAAA
su_wt      CCCATTGAACTTTCTCGAATTTCTGTTGTTGAACATAGTACTACCAGCAAGAATCAGCAAA
ch1         CCCATTGAACTTTCTCGAATTTCTGTTGTTGAACATAGTACTACCAGCAAGAATCAGCAAA
*****

su_mut      GCTGCGCCAGCAAATACAGATGCTGCAGAATTCTAACAGGTACCCTTGATTCATATTTCA
su_Di Genova GCTGCGCCAGCAAATACAGATGCTGCAGAATTCTAACAGGTACCCTTGATTCATATTTCA
ch2         GCTGCGCCAGCAAATACAGATGCTGCAGAATTCTAACAGGTACCCTTGATTCATATTTCA
VIT_218S0041G01880.2 GCTGCGCCAGCAAATACAGATGCTGCAGAATTCTAACAGGTACCCTTGATTCATATTTCA
su_wt      GCTGCGCCAGCAAATACAGATGCTGCAGAATTCTAACAGGTACCCTTGATTCATATTTCA
ch1         GCTGCGCCAGCAAATACAGATGCTGCAGAATTCTAACAGGTACCCTTGATTCATATTTCA
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su_mut      TTTTCTCACATGTAATTGAGTGTATATATATATGTCAATTTGCATTTTCTTCTGCTTG
su_Di Genova TTTTCTCACATGCAGTTGAGTGTATATATATATGTCAATTTGCATTTTCTTCTGCTTG
ch2         TTTTCTCACATGCAGTTGAGTGTATATATATATGTCAATTTGCATTTTCTTCTGCTTG
VIT_218S0041G01880.2 TTTTCTCACATGCAGTTGAGTGTATATATATATGTCAATTTGCATTTTCTTCTGCTTG
su_wt      TTTTCTCACATGCAGTTGAGTGTATATATATATGTCAATTTGCATTTTCTTCTGCTTG
ch1         TTTTCTCACATGCAGTTGAGTGTATATATATATGTCAATTTGCATTTTCTTCTGCTTG
***** * *****

su_mut      TGTCTTAACTAGGTTTTTGGCATCAGTTCCCATGCAAAAATTTCAAATATTACTTAAAG
su_Di Genova TGTCTTAACTAGGTTTTTGRCATCAGTTCCCATGCAAAAATTTCAAATATTACTTAAAG
ch2         TGTCTTAACTAGGTTTTTGACATCAGTTCCCATGCAAAAATTTCAAATATTACTTAAAG
VIT_218S0041G01880.2 TGTCTTAACTAGGTTTTTGACATCAGTTCCCATGCAAAAATTTCAAATATTACTTAAAG
su_wt      TGTCTTAACTAGGTTTTTGGCATCAGTTCCCATGCAAAAATTTCAAATATTACTTAAAG
ch1         TGTCTTAACTAGGTTTTTGGCATCAGTTCCCATGCAAAAATTTCAAATATTACTTAAAG
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su_mut      ATCTTAATTAGTATGTTAAGACTTAAGAGCAGGTTATACTAAGACATTTTGTCTGTGTT
su_Di Genova ATCTTAATTAGATATGTTAAGACTTAAGATCAGGTTGACTAAGACATTTTGTCTGTGTT
ch2         ATCTTAATTAGATATGTTAAGACTTAAGATCAGGTTGACTAAGACATTTTGTCTGTGTT
VIT_218S0041G01880.2 ATCTTAATTAGATATGTTAAGACTTAAGATCAGGTTGACTAAGACATTTTGTCTGTGTT
su_wt      ATCTTAATTAGATATGTTAAGACTTAAGATCAGGTTGACTAAGACATTTTGTCTGTGTT
ch1         ATCTTAATTAGATATGTTAAGACTTAAGATCAGGTTGACTAAGACATTTTGTCTGTGTT
***** * *****

su_mut      TATAGGCACTTAATGGGTGATTCCCTGGCTTCCTTGACTGTGAAGGAGCTAAAGCAGCTC
su_Di Genova TATAGGCACTTAATGGGTGATTCCCTGGCTTCCTTGACTGTGAAGGAGCTAAAGCAGCTC
ch2         TATAGGCACTTAATGGGTGATTCCCTGGCTTCCTTGACTGTGAAGGAGCTAAAGCAGCTC
VIT_218S0041G01880.2 TATAGGCACTTAATGGGTGATTCCCTGGCTTCCTTGACTGTGAAGGAGCTAAAGCAGCTC
su_wt      TATAGGCACTTAATGGGTGATTCCCTGGCTTCCTTGACTGTGAAGGAGCTAAAGCAGCTC
ch1         TATAGGCACTTAATGGGTGATTCCCTGGCTTCCTTGACTGTGAAGGAGCTAAAGCAGCTC
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su_mut      GAGAACAGGCTTGAACGAGGCATCACAAGAATCAGGTCGAAGAAGGTAACGACAC--ATA
su_Di Genova GAGAACAGGCTTGAACGAGGCATCACAAGAATCAGGTCGAAGAAGGTAACGACAC--ATA
ch2         GAGAACAGGCTTGAACGAGGCATCACAAGAATCAGGTCGAAGAAGGTAACGACACCTAGA
VIT_218S0041G01880.2 GAGAACAGGCTTGAACGAGGCATCACAAGAATCAGGTCGAAGAAGGTAACGACACCTAGA
su_wt      GAGAACAGGCTTGAACGAGGCATCACAAGAATCAGGTCGAAGAAGGTAACGACACCTAGA
ch1         GAGAACAGGCTTGAACGAGGCATCACAAGAATCAGGTCGAAGAAGGTAACGACACCTAGA
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su_mut      CACTAATCTAATCTGGCTGGGCATTTGACTTTGTGACTTAAATGAAATTGAAAAA
su_Di Genova CACTAATCTAATCTGGCTGGGCATTTGACTTTGTGACTTAAATGAAATTGAAAAA
ch2         CACTAATCTAATCTGGCTGGGCATTTGACTTTGTGACTTAAATGAAATTGAAAAA

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VIT_218S0041G01880.2 CACTAAATCTAATCTGGCTGAGCATTTGATTTTGGAGACTTAAATGAAA-TGGAAAAAAA
su_wt CACTAAATCTAATCTGGCTGAGCATTTGATTTTGGAGACTTAAATGAAA-TGGAAAAAAA
ch1 CACTAAATCTAATCTGGCTGAGCATTTGATTTTGGAGACTTAAATGAAA-TGGAAAAAAA
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su_mut ATGGTATCATTCACCTGATTA---AATGATACATGAAGATTGAATTAATTTACGATACCTGC
su_Di Genova ATGGTATCATTCACCTGATTAATAATAATAACTTGAAGGTTGAATTAATTTAAGATACCGC
ch2 ATGGTATCATTCACCTGATTAATAATAATAACTTGAAGGTTGAATTAATTTAAGATACCGC
VIT_218S0041G01880.2 ATGGTATCATTCACCTGATTAATAATAATAACTTGAAGGTTGAATTAATTTAAGATACCGC
su_wt ATGGTATCATTCACCTGATTAATAATAATAACTTGAAGGTTGAATTAATTTAAGATACCGC
ch1 ATGGTATCATTCACCTGATTAATAATAATAACTTGAAGGTTGAATTAATTTAAGATACCGC
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su_mut TTTCACCTTGATAACTAAAGCCTGATGGTAGGAACTTCCACCCATCTAGCTTAGTTGGTGA
su_Di Genova TTTCACCTTGATAACTAAAGCCTGATGGTAGGAACTTCCACCCATCTAGCTTAGTTGGTGA
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VIT_218S0041G01880.2 TTTCACCTTGATAACTAAAGCCTGATGGTAGGAACTTCCACCCATCTAGCTTAGTTGGTGA
su_wt TTTCACCTTGATAACTAAAGCCTGATGGTAGGAACTTCCACCCATCTAGCTTAGTTGGTGA
ch1 TTTCACCTTGATAACTAAAGCCTGATGGTAGGAACTTCCACCCATCTAGCTTAGTTGGTGA
*****

su_mut TTGAGCTGAATTTTGTCTTGGAACCTTGTGAAAAATAGACAAGTGATCACTAGATCAGC
su_Di Genova TTGAGCTGAATTTTGTCTTGGAACCTTGTGAAAAATAGACAAGTGATCACTAGATCAGC
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VIT_218S0041G01880.2 TTGAGCTGAATTTTGTCTTGGAACCTTGTGAAAAATAGACAAGTGATCACTAGATCAGC
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ch1 TTGAGCTGAATTTTGTCTTGGAACCTTGTGAAAAATAGACAAGTGATCACTAGATCAGC
*****

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su_Di Genova AAAGCCTATCTTTTTTCCCTTGTCTTAATGTGCTGCTTGTCTAATGCAATAGCATGAGT
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*****

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* *****

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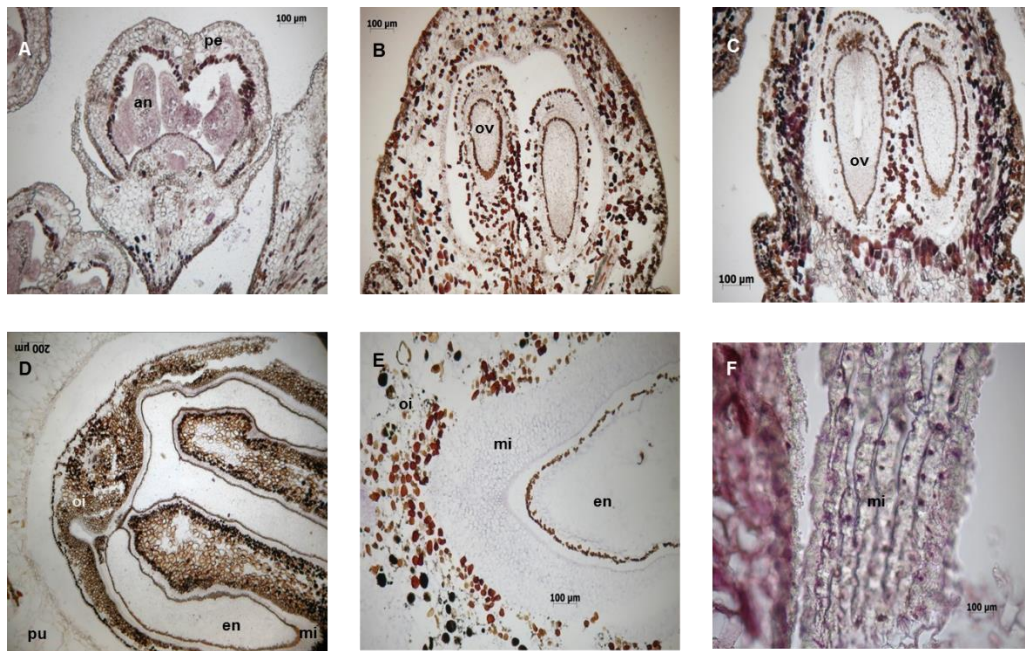
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ch2         GCCTATTAATAAT
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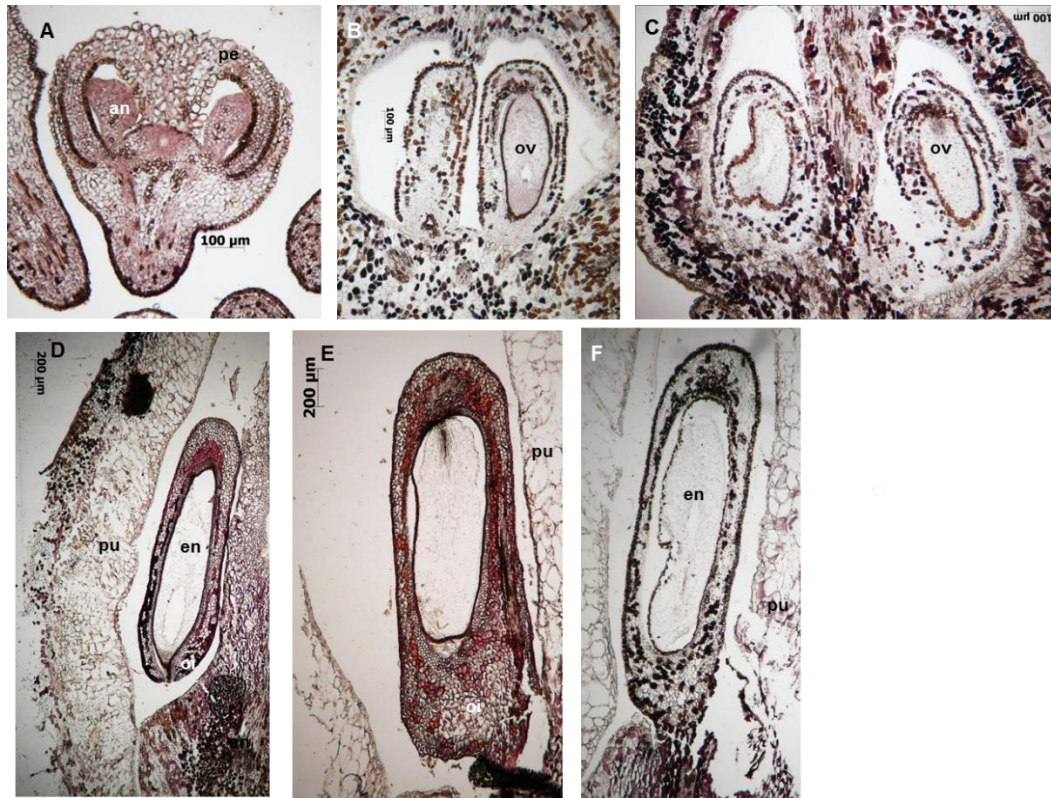
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Supplementary Fig. 1. Complete alignment of *VvAGL11* alleles. The sequences presented in the alignment are *VvAGL11* PN40024 (VIT_218S0041G01880.2), *VvAGL11*

‘Chardonnay allele 1’ (CH1), *VvAGL11* ‘Chardonnay allele 2’ (CH2), *VvAGL11* ‘Sultanina’ mutant allele (*SU mut*), *VvAGL11* ‘Sultanina wt allele’ (*SU wt*) and the available sequence from ‘Sultanina’ by Di Genova (2014). Underlined sequence represents the SSR marker VMC7F2, blue sequence marks represent UTRs, red sequences comprise exons and green enhancement are indicating putative CArG-Box sequences.



Supplementary Fig. 2. *In situ* hybridization of *VvAGL11* sense probe (control) in ‘Chardonnay’. H1 flower stage (A), pre anthesis flower stage (B), fruit set flower stage (C), 2 weeks (D), 4 weeks (E) and 6 weeks (F) of fruit development. Anther (an), endosperm (en), medium integument (mi), outer integument (oi), ovule (ov), petal (pe), pulp (pu).



Supplementary Fig. 3. *In situ* hybridization of *VvAGL11* sense probe (control) in ‘Sultanina’. H1 flower stage (**A**), pre anthesis flower stage (**B**), fruit set flower stage (**C**), 2 weeks (**D**), 4 weeks (**E**) and 6 weeks (**F**) of fruit development. Anther (an), endosperm (en), medium integument (mi), outer integument (oi), ovule (ov), petal (pe), pulp (pu).

4 CONCLUSÕES

A utilização da técnica de sequenciamento alelo-específica mostrou-se eficiente na identificação das variações alélicas de *VvAGL11* em ‘Chardonnay’ e ‘Sultanina’. Foram identificados conjuntos de *SNPs* e *INDELS* que permitem diferenciar os alelos avaliados. Com isso, foi possível discernir dois alelos para ‘Chardonnay’ e dois para ‘Sultanina’. Um dos alelos de ‘Sultanina’, o alelo *SU mut*, é o único que possui mutações pontuais em seu RNA mensageiro, estas compostas por 10 *SNPs*, dois deles realizando modificações de aminoácidos na região codificadora. O sequenciamento por *RACE* permitiu a identificação dos *mRNAs* de *VvAGL11* de cada cultivar, mostrando que os quatro alelos são transcritos, porém ‘Chardonnay’ possui um alelo, CH1, com uma região codificante 60 nucleotídeos maior do que os outros alelos avaliados.

A quantificação de transcritos por meio da técnica de *RT-qPCR* permitiu a avaliação das isoformas de transcritos de *VvAGL11* previamente identificadas por meio de *RACE*. A expressão gênica relativa de cada isoforma foi avaliada em amostras de ambas as cultivares nos estádios fenológicos de flores e frutos. As isoformas de *VvAGL11* identificadas em ‘Sultanina’ são igualmente pouco expressas em todos os estádios analisados, enquanto que as isoformas de ‘Chardonnay’ também são pouco expressas em flores e polpa. Em contraste, os alelos de CH1 e CH2 possuem uma expressão 15-65 vezes maior em sementes em comparação com flores e frutos tanto de ‘Chardonnay’ como de ‘Sultanina’.

Para a investigação de diferenças morfológicas entre as sementes de ‘Chardonnay’ e o traço de semente de ‘Sultanina’, o emblocamento dos tecidos em resina plástica foi realizado. Isto permitiu a observação da provável perda de identidade da camada de integumento médio do traço de semente, o qual não se alongou nem se duplicou como

esperado em quatro semanas, mantendo sua forma original indiferenciada durante todo o desenvolvimento do fruto.

A caracterização da expressão tecido-específica de *VvAGL11* foi realizada com sucesso por meio da investigação do padrão de expressão espaço temporal do gene pela técnica de hibridização *in situ*. Ambas as cultivares e os estádios de desenvolvimento de flor e fruto utilizadas nos ensaios de *RT-qPCR* foram analisados. O acúmulo dos transcritos de *VvAGL11* foi detectado especificamente na camada de integumento médio da semente de ‘Chardonnay’, a qual é responsável pelo alongamento da casca em quatro semanas. Esta é justamente a camada que apresentou atraso de desenvolvimento na cultivar estenoespermocárpica Sultanina. Todavia, não foi detectada hibridização do gene na cultivar apirênica ‘Sultanina’, nem mesmo no traço de semente analisado. Os resultados são consistentes com a hipótese que a expressão de *VvAGL11* é necessária para a formação da semente e a análise dos dados permite sugerir que a diminuição de sua expressão pode estar relacionada à perda de identidade da camada de integumento médio da semente o que afeta seu tamanho final e leva ao fenótipo de apirenia.

5 PERSPECTIVAS

- * Caracterizar a funcionalidade de *VvAGL11* por meio de complementação em modelo heterólogo (*A. thaliana*);
- * Avaliar a ortologia de *VvAGL11* e *AtAGL11* por meio da complementação de mutante *stk* (*seedstick*);
- * Caracterizar funcionalmente *VvAGL11* em videiras por meio da expressão do gene em cultivares apirênicas e do seu silenciamento em cultivares pirênicas;
- * Caracterizar as interações da proteína VvAGL11 com seus genes-alvo e os elementos *cis* (CArG-box) críticos às interações;
- * Identificar as proteínas que constituem complexos quaternários de transcrição juntamente com VvAGL11;
- * Identificar um haplótipo associado com potencial para seleção assistida da apirenia em videira;
- * Desenvolver e testar um marcador definitivo para apirenia a partir de SNPs do alelo *VvAGL11* de Sultanina mutado, almejando a geração de patente sobre essas marcas;
- * Avaliar cinco populações segregantes para o fenótipo de apirenia (1.000 indivíduos) com os haplótipos de marcadores microssatélites e com o marcador definitivo.

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VITTI, A. Análise da competitividade das exportações brasileiras de frutas selecionadas no mercado internacional. Universidade de São Paulo. Escola superior de agricultura Luis de Queiroz. Dissertação de mestrado. 2009.

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YUN, P. Y.; KIM, S. Y.; OCHIAI, T.; FUKUDA, T.; ITO, T.; KANNO, A. AVAG2 is a putative D-class gene from an ornamental asparagus. *Sexual Plant Reproduction*, 17: 107–116. 2004.

7 CURRICULUM VITAE

DADOS PESSOAIS

Nome: Jaiana Malabarba

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FORMAÇÃO

2013 – 2014

Mestrado em Biologia Celular e Molecular, PPGBCM, Universidade Federal do Rio Grande do Sul, UFRGS, Brasil. Estrutura do gene *VvAG111* e análise da expressão durante a morfogênese da semente de videira. Orientador: Dr. Giancarlo Pasquali.

2009 – 2013

Graduação em Ciências Biológicas. Universidade do Vale do Rio dos Sinos, UNISINOS, São Leopoldo, RS, Brasil. Autoecologia De *Aulacoseira Granulata* Ehrenberg (Bacillariophyta) Em Ambientes Lóticos Subtropicais Com Diferentes Graus De Preservação. Dr^a Luciane Oliveira Crossetti.

FORMAÇÃO COMPLEMENTAR

2011 – 2013

Estágio de Iniciação Científica no **Laboratório de Genética Molecular Vegetal**. EMBRAPA Uva e Vinho, Bento Gonçalves/RS.

Orientação: Dr. Luís Fernando Revers.

Pesquisa inserida no projeto: “Uvas do Brasil: Programa de Melhoramento Genético”.

2010 – 2012

Bolsista de Iniciação Científica Petrobras no **Laboratório de Ecofisiologia e Cultura Vegetal**. UNISINOS, São Leopoldo/RS.

Orientação: Dr^a. Luciane Oliveira Crossetti.

Atuando nos seguintes temas: Estudo das comunidades fitoplânctônicas de diferentes arroios da Bacia do Rio dos Sinos através da avaliação da influência da composição de mata ciliar.

2010 – 2011

Estágio de Iniciação Científica no **Laboratório de Enoquímica**. EMBRAPA Uva e Vinho, Bento Gonçalves/RS.

Orientação: Dr. Celito Crivellaro Guerra.

Atuando nos seguintes temas: Acompanhamento e análises da maturação fenólica, com determinação de taninos e antocianinas presentes em cascas e sementes de diversas variedades de uva provenientes de regiões produtoras dos estados do Rio Grande do Sul e de Santa Catarina no projeto “Novas Regiões Vitivinícolas”.

PRÊMIOS E TÍTULOS

2013

1º lugar da Pós-graduação com o trabalho 'Padrão de expressão espaço-temporal do gene *VvAGL11* em tecidos reprodutivos de videira por hibridização *in situ*', Embrapa Uva e Vinho.

2013

Proficiência em leitura da Língua Inglesa, Universidade Federal do Rio Grande do Sul.

2012

Laureada como Jovem Cientista em Fruticultura Brasileira, Sociedade Brasileira de Fruticultura.

2012

Distinção em Trabalho de Conclusão de Curso, Universidade do Vale do Rio dos Sinos.

2012

Menção honrosa com o trabalho "Avaliação De Marcadores SSR Com Potencial Para Seleção Assistida De Videiras Apirênicas e Resistentes Ao Míldio" no 2º Prêmio Jovem Cientista em Fruticultura Brasileira, Sociedade Brasileira de Fruticultura.

2011

Destaque em Mostra de Iniciação Científica, Universidade do Vale do Rio dos Sinos.

RESUMOS PUBLICADOS EM ANAIS DE CONGRESSOS

- * MALABARBA, J.; BUFFON, V.; PASQUALI, G.; REVERS, L. F. Estudo estrutural e funcional do gene *VvAGL11* e seu papel na morfogênese de sementes de *Vitis vinifera*. In: 12º Encontro de Iniciação Científica e 8º Encontro de Pós-Graduandos da Embrapa Uva e Vinho, Bento Gonçalves. 2014.
- * MALABARBA, J.; BUFFON, V.; WAIRICH, A.; MARIATH, J.; GAETA, M.; DORNELAS, M. C.; REVERS, L. F.; PASQUALI, G. Padrão de Expressão Espaço-temporal do Gene *VvAGL11* em Tecidos Reprodutivos de Videira por Hibridização *In situ*. In: X Reunião Anual do Programa de Pós-Graduação em Biologia Celular e Molecular. Porto Alegre: UFRGS, 2013.
- * MALABARBA, J.; BUFFON, V.; WAIRICH, A.; MARIATH, J.; GAETA, M.; DORNELAS, M. C.; REVERS, L. F. Gene expression analysis of *VvAGL11* in grapevine reproductive tissues by *In situ* hybridization. In: IV Simpósio Brasileiro de Genética Molecular de Plantas, Bento Gonçalves, RS. 2013.
- * WAIRICH, A.; MALABARBA, J.; BUFFON, V.; REVERS, L. F. Evaluation of resistance to *Plasmopara viticola* in a self-fertilized segregating population from Villard Blanc (*Vitis* spp.). In: IV Simpósio Brasileiro de Genética Molecular de Plantas, Bento Gonçalves, RS. 2013.

- * MALABARBA, J.; BUFFON, V.; WAIRICH, A.; MARIATH, J.; GAETA, M.; DORNELAS, M. C.; PASQUALI, G.; REVERS, L. F. Padrão de expressão espaço-temporal do gene *VvAGL11* em tecidos reprodutivos de videira por hibridização *In situ*. *In: 11º Encontro de Iniciação Científica e 7º Encontro de Pós-Graduandos da Embrapa Uva e Vinho, Bento Gonçalves. 2013.*

- * WAIRICH, A.; MALABARBA, J.; BUFFON, V.; REVERS, L. F.; WELTER, L. J. Avaliação da resistência ao *Plasmopara viticola* em uma população segregante para a resistência auto-fertilizada de 'Villard Blanc' (*Vitis* spp.). *In: 11º Encontro de Iniciação Científica e 5º Encontro de Pós-Graduandos da Embrapa Uva e Vinho, 2012, Bento Gonçalves. 2013.*

- * MALABARBA, J.; WAIRICH, A.; BUFFON, V.; CZERMAINSKI, A.; REVERS, L. F. Avaliação de marcadores SSR com potencial para seleção assistida de videiras apirênicas e resistentes ao míldio. *In: XXII Congresso Brasileiro de Fruticultura, 2012, Bento Gonçalves. XXII CBF, 2012.*

- * WAIRICH, A.; MALABARBA, J.; BUFFON, V.; CZERMAINSKI, A.; REVERS, L. F. Análise do perfil transcricional de genes candidatos associados à resistência ao míldio em videira, presentes no locus Rpv3. *In: XXII Congresso Brasileiro de Fruticultura, 2012, Bento Gonçalves. XXII CBF, 2012.*

- * WAIRICH, A.; MALABARBA, J.; BUFFON, V. ; PORTO, D. D. ; REVERS, L. F. Análise do perfil transcricional de genes candidatos associados à resistência ao míldio em videira, presentes no locus Rpv3. *In: 10º Encontro de Iniciação Científica e 6º Encontro de Pós-Graduandos da Embrapa Uva e Vinho, Bento Gonçalves. 2012.*

- * MALABARBA, J.; WAIRICH, A.; BUFFON, V.; CZERMAINSKI, A.; REVERS, L. F. Avaliação de marcadores SSR com potencial para seleção assistida de videiras apirênicas e resistentes ao míldio. *In: 10º Encontro de Iniciação Científica e 6º*

Encontro de Pós-Graduandos da Embrapa Uva e Vinho, 2012, Bento Gonçalves. 2012.

- * MALABARBA, J.; BUFFON, V.; CZERMAINSKI, A.; REVERS, L. F. Caracterização do perfil transcricional de um gene associado à estenoespermocarpia em videira (*Vitis vinifera* L.). III Simpósio de Biodiversidade, UFSM, Santa Maria, RS. 2011.
- * MALABARBA, J.; BUFFON, V.; CZERMAINSKI, A.; REVERS, L. F. Caracterização do perfil transcricional de genes associados à estenoespermocarpia em videira (*Vitis vinifera* L.). In: 9º Encontro de Iniciação Científica e 5º Encontro de Pós-Graduandos da Embrapa Uva e Vinho, Bento Gonçalves. 2011.

PARTICIPAÇÃO EM CURSOS

- * I Workshop Interno De Biologia Molecular De Plantas. Universidade Federal do Rio Grande do Sul. 2014.
- * Theoretical and practical aspects of gene silencing in plants. Peter Waterhouse & Rogério Margis. PPGBCM, Universidade Federal do Rio Grande do Sul. 2013.
- * Sequenciamento e Análise de Fragmentos através da Eletroforese Capilar na Plataforma 310. Embrapa Uva e Vinho, Bento Gonçalves. Life Technologies Brasil. 2013.
- * REVERS, L.F.; PORTO, D.D.; FALAVIGNA, V.S.; BUFFON, V.; MALABARBA, J. Genética Molecular Vegetal: mapeamento genético, identificação de QTLs, genes candidatos e suas aplicações tecnológicas. 2012. (Curso de curta duração ministrado/Extensão).

- * Manejo e uso de recursos genéticos de uva. Embrapa Uva e Vinho, Bento Gonçalves. 2011.
- * PCR em Tempo Real - Princípios e Aplicações. Life Technologies Brasil. 2011.

ATIVIDADES DESENVOLVIDAS NO PROGRAMA DE PÓS-GRADUAÇÃO EM BIOLOGIA CELULAR E MOLECULAR - UFRGS

Período Letivo	Atividade de Ensino	Créditos	Conceito
2014/1	Atividade Didática	3	A
2014/1	Redação Científica Manuscrito	1	A
2014/1	Seminário do Centro de Biotecnologia	2	A
2014/1	Seminários de Dados Experimentais Apresentação	1	A
2014/1	Seminários de Dados Experimentais Frequência	1	A
2014/1	Tópicos Avançados	3	A
2013/2	Biologia Molecular	4	B
2013/2	Estrutura e Estratégias de Análise Genômica Vegetal	3	A
2013/1	Biologia Celular	3	A
2013/1	Curso- Theoretical and practical aspects of gene silencing in plants	2	A
2013/1	Fundamentos de Análise de Proteínas	2	A