



UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
INSTITUTO DE BIOCÊNCIAS
DEPARTAMENTO DE ZOOLOGIA
LABORATÓRIO DE CARCINOLOGIA

Desempenho alimentar de *Balloniscus sellowii* (Brandt, 1833) (Crustacea: Isopoda: Oniscidea) em folhas de diferentes estágios de decomposição e sua relação com o conteúdo de fenólicos e de flavonoides

CAMILA TIMM WOOD

Monografia apresentada à banca examinadora do Instituto de Biotecnologia, Universidade Federal do Rio Grande do Sul, como requisito parcial à obtenção do grau de Bacharel em Ciências Biológicas.

Orientador: Prof^a. Dr^a. Paula B. Araujo

Co-orientador: Prof. Dr. Geraldo L. G. Soares

Artigo a ser submetido no *Journal of Crustacean Biology*

Porto Alegre, julho de 2009.

Desempenho alimentar de *Balloniscus sellowii* (Brandt, 1833) (Crustacea: Isopoda: Oniscidea) em folhas de diferentes estágios de decomposição e sua relação com o conteúdo de fenólicos e de flavonoides

Camila Timm Wood

Monografia de bacharelado aprovada em 1º de Julho de 2009.

Orientação: Prof^a. Dr^a. Paula Beatriz de Araujo

Co-orientação: Prof. Dr. Geraldo Luiz Gonçalves Soares

Banca examinadora:

Prof. Dr. José Cláudio Fonseca Moreira

Prof. Dr. Milton de Souza Mendonça Junior

AGRADECIMENTOS

À UFRGS e CNPq pela infra-estrutura e bolsa concedida para o projeto.

Aos meus mestres, pessoas que me ensinaram mais que biologia, meu sincero obrigada. Os professores do início do curso me transmitiram conhecimento de base e os do final de curso que me ajudaram a ligar essas informações e ter uma ideia muito mais integrada da biologia.

Aos meus amigos por entenderem a ausência devido ao trabalho de pesquisa que exige dedicação integral.

Aos meus pais por terem me ensinado a importância do estudo e incentivado a minha curiosidade, criando uma pesquisadora desde pequena.

Aos meus colegas do laboratório de Carcinologia, especialmente a Priscila Bugs e Carina Appel pela ajuda em coleta.

Ao meu co-orientador Geraldo Soares por me auxiliar na parte química do trabalho e a minha colaboradora Carolina Schlindwein pelas ajudas nas análises que nem sempre davam certo de primeira, mas que sempre eram divertidas.

À minha orientadora Paula por ser um exemplo de profissional, estando sempre disponível e sendo muito mais que uma orientadora, sendo também uma amiga.

SUMÁRIO

APRESENTAÇÃO	7
OBJETIVOS DO TRABALHO	9
“FEEDING PERFORMANCE OF <i>BALLONISCUS SELLOWII</i> (ISOPODA: ONISCIDEA) IN LEAF LITTER OF DIFFERENT STAGES OF DECOMPOSITION AND ITS RELATION TO THE PHENOLIC AND FLAVONOID CONTENT”	11
ABSTRACT	13
RESUMO	14
INTRODUCTION	15
MATERIAL AND METHODS	18
RESULTS	22
DISCUSSION	25
REFERENCES	31
TABLE	39
LIST OF FIGURES	41
FIGURES	43
CONSIDERAÇÕES FINAIS	55
REFERÊNCIAS BIBLIOGRÁFICAS	57
ANEXO 1	61

APRESENTAÇÃO

Os isópodos terrestres (Isopoda: Oniscidea), comumente conhecidos como “tatuzinhos”, são crustáceos que obtiveram grande sucesso na colonização do ambiente terrestre. Fazem parte da macrofauna de solo e estão fortemente envolvidos nos processos de detritivoria (Hassall et al., 1987; Sousa et al., 1998; Zimmer & Topp, 1999; Kautz & Topp, 2000). Habitam grande variedade de ambientes, sendo um dos grupos de detritívoros mais abundantes em diversas comunidades (Hassall & Dangerfield, 1990; Zimmer, 2002a). São de fácil captura e cultivo em laboratório, característica que os fazem ótimos modelos para um maior entendimento da dinâmica da formação do solo.

Nessa transição para o ambiente terrestre, esse grupo necessitou de adaptações às fontes de alimentação. As espécies de plantas terrestres possuem maior quantidade de lignina e celulose e de compostos secundários, sendo necessárias, diversas enzimas para a quebra de compostos complexos (Zimmer, 2002b). Entre as estratégias utilizadas por esses animais estão a aquisição de bactérias endossimbiontes que contribuem para a digestão de lignina e celulose (Zimmer & Topp, 1998; Zimmer et al. 2001; Zimmer, 2006) e o comportamento coprofágico para um melhor aproveitamento nutricional da fonte de alimento ingerida primariamente (Hassall & Rushton, 1982; Gunnarson & Tunid, 1986; Szlávecz & Pobozy, 1995).

Por serem detritívoros, eles servem de ligação entre produtores primários e níveis tróficos mais altos, contribuindo nos processos de decomposição por consumir grandes quantidades de serapiheira e por promover atividade microbiana (Zimmer, 2002b). Eles retornam grande quantidade de matéria na forma de fezes, o que contribui para a formação do húmus do solo e para o crescimento das florestas

(Knoepp et al., 2000; Förster et al., 2006) e aumentam a qualidade do solo (Kautz & Topp, 2000). Hassall et al. (1987), combinando dados de pesquisadores, sugeriram que cerca de 80,8% da serapilheira seria consumida pela fauna de solo.

Oniscidea inclui aproximadamente 3600 espécies de tamanho pequeno a médio (1,2-30mm) distribuídas por todo o mundo (Schmalfuss, 2003). Para o Brasil, cerca de 120 espécies são registradas (Leistikow & Wägele, 1999), sendo o gênero *Balloniscus* Budde-Lund, 1908 representado por *B. glaber* Araujo & Zardo, 1995 e *B. sellowii* (Brandt, 1833), ambas as espécies nativas.

Estudos sobre nutrição de invertebrados têm sido realizados em sua maioria com insetos (Appel, 1993; Harborne, 1993). Os estudos com isópodos, realizados na maioria na Europa (Gunnarsson, 1987; Szlávecz, 1992, 1993, Szlávecz & Pobožny, 1995, Rushton & Hassall, 1983a,b; Souza et al., 1998; Zimmer, 2002, Ihnen & Zimmer, 2008), incluem espécies de *Porcellio* Latreille, 1804 e *Armadillidium* Brandt 1833, as quais apresentam distribuição mundial. Estudos com espécies neotropicais são mais escassos, não obstante sua importância para o entendimento da dinâmica de solo, precisando estes, portanto, serem encorajados.

OBJETIVOS DO TRABALHO

Sabe-se que organismos detritívoros são de grande importância para a formação do solo. Para isso, esse trabalho foi realizado para contribuição ao conhecimento sobre a nutrição de isópodos terrestres.

Objetivos específicos:

- comparar o desempenho alimentar (taxas de consumo, de egestão e de assimilação) em folhas de diferentes estágios de decomposição;
- verificar a eficiência de assimilação dos animais nas diferentes folhas e o tempo de passagem do alimento pelo trato digestório;
- verificar o conteúdo de fenólicos totais e de flavonoides das folhas consumidas;
- estimar a quantidade de fenólicos totais e de flavonoides ingeridos pelos isópodos;
- relacionar o desempenho alimentar dos animais com o conteúdo de fenólicos totais e de flavonoides.

**“FEEDING PERFORMANCE OF *BALLONISCUS SELLOWII* (ISOPODA: ONISCIDEA) IN LEAF
LITTER OF DIFFERENT STAGES OF DECOMPOSITION AND ITS RELATION TO THE PHENOLIC AND
FLAVONOID CONTENT”**

Camila Timm Wood, Paula Beatriz Araujo, Carolina Casco Duarte Schlindwein, and
Geraldo Luiz Gonçalves Soares

(CTW, PBA) Laboratório de Carcinologia, Departamento de Zoologia, Universidade Federal
do Rio Grande do Sul, Brazil

(CCDS, GLGS) PPG Ecologia, Laboratório de Laboratório de Quimiotaxonomia e Ecologia
Química, Departamento de Botânica, Universidade Federal do Rio Grande do Sul, Brazil

(CTW, ctwood86@gmail.com; PBA, pbaraujo@portoweb.com.br, CCDS,
carolcasco@gmail.com, GLGS, glgsoares@gmail.com)

ABSTRACT

The goal of this study was to use a terrestrial isopod to compare feeding performance in leaves of different stages of decomposition according to their phenolic and flavonoid content. Leaves from the most visually abundant plants were offered to isopods collected in an urban area. The plant species that presented the highest consumption was used to verify feeding performance in different stages of decomposition. Green leaves were collected from branches and placed in litterbags, left to decompose, collected after one, two and three months. Then they were offered to animals as well as additional green leaves. The total phenolic and flavonoid content was determined for green leaves and other stages of decomposition. The consumption rate was significantly higher when animals fed on *Schinus terebinthifolius* and there was no significant difference between *Lithraea brasiliensis* and *Ricinus communis*. *Schinus terebinthifolius* was tested in four stages of decomposition, and the consumption and egestion rates were significantly higher on two months-old leaves. The assimilation rate was significantly highest in green leaves. There was a correlation between the consumption and egestion rates. The mode time of passage through gut was two hours for all treatments and the beginning of ingestion of leaves occurred after two or three days for green leaves, and in the same day for one, two and three months-old leaves. Assimilation efficiency was 43% for green leaves and about 20% for the other stages. The phenolic content was highest in green and lowest in the third month, and the flavonoid content was highest in green leaves and lowest in the second month. The estimated phenolic amount ingested by animals was significantly highest when consumption rate was also highest. The estimated flavonoid amount ingested followed the same trend as the assimilation rate, being significantly higher in green leaves. The mean flavonoid content of each decomposition stage was correlated with the assimilation efficiency. The highest consumption (optimal palatability) occurred on two months-old leaves and not when the phenolic content was lowest. The flavonoids represented a large portion of the total phenolics and the estimated amount of flavonoid consumed by animals was almost the same for leaves with one, two and three months of decomposition. It seems that animals increase consumption to maintain a minimum intake of the flavonoids in leaves with lower flavonoid content, suggesting that they might use these flavonoids as a food parameter.

KEY WORDS: leaf decomposition, Oniscidea, total phenolics, flavonoids.

RESUMO

O objetivo do trabalho foi utilizar um isópodo terrestre para verificar o desempenho alimentar de folhas em diferentes estágios de decomposição e a sua relação com o conteúdo de fenólicos e de flavonoides. Folhas das plantas mais abundantes do local foram oferecidas aos isópodos coletados em uma área urbana. A espécie de planta que obteve o maior consumo foi utilizada para verificar o desempenho alimentar em diferentes estágios de decomposição. Folhas verdes foram coletadas dos galhos e colocadas em *litter bags* que foram recolhidas após um, dois e três meses e oferecidas aos animais, assim como folhas verdes adicionais. O conteúdo de fenólicos totais e de flavonoides foi determinado para folhas verdes e em cada estágio de decomposição. A taxa de consumo foi significativamente maior quando os animais consumiram *Schinus terebinthifolius* e não houve diferença significativa entre *Lithraea brasiliensis* e *Ricinus communis*. *Schinus terebinthifolius* foi testada em quatro estágios de decomposição e as taxas de consumo e de egestão foram significativamente maiores em folhas de dois meses. A taxa de assimilação foi significativamente maior em folhas verdes. Houve correlação entre as taxas de consumo e de egestão. A moda do tempo de passagem pelo trato digestório foi de duas horas para todos os tratamentos e o início da ingestão das folhas ocorreu após dois ou três dias para folhas verdes, e no mesmo dia para folha de um, dois e três meses. A eficiência de assimilação foi de 43% para folhas verdes e em torno de 20% para os demais estágios. O conteúdo de fenólicos foi maior nas folhas verdes e menor no terceiro mês e o conteúdo de flavonoides foi maior nas folhas verdes e menor em folhas do segundo mês. A quantidade estimada de fenólico ingerido pelos animais foi significativamente maior quando o consumo foi maior. A quantidade estimada de flavonoides ingeridos apresentou o mesmo padrão das taxas de assimilação, sendo significativamente maior em folhas verdes. A média do conteúdo de flavonoide de cada mês foi correlacionada com a eficiência de assimilação. O maior consumo (palatabilidade ótima) ocorreu em folhas de dois meses e não quando o conteúdo de fenólicos era mais baixo. Os flavonoides representaram uma grande porção do conteúdo dos fenólicos totais e a quantidade estimada de flavonoides consumidos pelos animais foi quase a mesma com folhas de um, dois e três meses de decomposição. Parece que os animais aumentaram o consumo para ingerir uma quantidade mínima de flavonoides quando o conteúdo de flavonoides era mais baixo, sugerindo que eles possam utilizar os flavonoides como um parâmetro alimentar.

PALAVRAS CHAVE: decomposição de folhas, Oniscidea, fenóis totais, flavonoides.

INTRODUCTION

Litter dynamics is of great importance in the functioning of the ecosystems and it is influenced by many different organisms. Isopods, earthworms, lumbricids, diplopods, dipteran larvae and termites are detritivores of organic soil litter that play a major role in the cycling of nutrients. Detritivores have low assimilation efficiency (Szláveczs and Pobožny, 1995) contributing more indirectly to leaf litter decomposition by returning great amounts of the consumed litter as feces (Quadros and Araujo, 2008) which present increased surfaces that are readily colonized by microbial populations (Hassall et al., 1987, Loureiro et al., 2006).

Detritivores present feeding preference that has been related to leaf senescence (Wieser, 1984; Hassall et al., 1987; Yeates and Barmuta, 1999), nutrient content of food (Graça et al., 2001), microbial colonization (Gunnarsson, 1987; Kautz and Topp, 2000; Zimmer et al., 2003; Ilnen and Zimmer, 2008) and the presence of unpalatable or indigestible compounds (Target et al., 1986; Canhoto and Graça, 1999, Lambdon and Hassall, 2005). The isopods contribution to decomposition depends on leaf litter degradation and may be influenced by food preference (Van Wensem et al. 1993).

Changes in the chemical composition of the litter due to decomposition increase its palatability to detritivores (Cameron and LaPoint, 1978; Neuhauser and Hartenstein, 1978; Rushton and Hassall, 1983; Hassall and Rushton, 1984; Hassall et al., 1987; Wan Wensen et al., 1993). During leaf senescence, there is a reduction in the phenolic content due to action of microorganisms and leaching (Zimmer, 1999b; Zimmer, 2002a) that increases palatability for detritivores (Cameron and LaPoint, 1978; Hassall and Rushton, 1984).

Phenolics are thought to play a fundamental role in chemical defense of plants against herbivores and pathogens (Harborne, 1993; Kefeli et al., 2003; Gould and Lister, 2006) although their effects are controversial and not fully understood (Appel, 1993; Johnson and Felton, 2001). Total phenolic content varies with plant growth and abiotic factors such as temperature and radiation (Salgado et al., 2008) and may affect microbial decomposers since a bulk of phenolics remains present during leaf senescence and after death (Bärlocher and Graça, 2005). Although not well understood, the existence of a phenolic cycle in the plant-soil system has been recorded (Kefeli et al., 2003). Differences in the composition and concentration of resin acids and phenolics during leaf and needle litter senescence have been recorded (Kuiters and Sarink, 1986; Kainulainen and Holopainen, 2002). Hassall and Rushton (1984) observed a rapid degradation of some phenolics (alkaloids and terpenes) during leaf senescence and a negative correlation between isopod feeding preference and the phenol content. However, Neuhauser and Hartenstein (1978) found no relation between leaf palatability and total phenolic content, and Kasurinen et al. (2007) found a weak or inconsistent correlation between detritivore feeding performance and chemical parameters of leaf litter.

Flavonoids are phenolic compounds commonly found in plants and might interfere with feeding, molting and reproduction of insects (Oberdörster et al., 2001; Simmonds, 2001; Boué and Raina, 2003; Gould and Lister, 2006). Some flavonoids play an important role in the protection of plants from harmful UV-B levels (Gould and Lister, 2006) and several classes of flavonoids show antioxidant activity towards a variety of readily-oxidizable compounds (Gryglewski et al., 1987; Dixon and Steele, 1999; Zhishen et al., 1999).

The ability to digest phenolics such as tannins and lignin is essential in the use of litter (Zimmer, 2002*b*) and studies have demonstrated that isopods are capable of oxidizing (Stevenson, 1961; Zimmer and Topp, 1998; Zimmer, 1999*b*; Zimmer et al, 2002) or hydrolyzing ingested phenolics (Zimmer, 1999*b*; Zimmer et al, 2002). Cameron and LaPoint (1978) observed decreased mortality and increased leaf consumption after leaching of tannins in *Armadillidium vulgare* (Latreille, 1804) that was related with leaf senescence, and suggested that litter resource cannot be used immediately after leaf fall due to chemical and mechanical defenses of plants.

The goal of this study was to use the terrestrial isopod *Balloniscus sellowii* (Brandt, 1833) as a detritivorous model, to verify feeding performance (consumption, egestion, and assimilation rates and assimilation efficiency) in leaves of different stages of decomposition according to their phenolic and flavonoid content.

MATERIAL AND METHODS

Species and study site

The species *Balloniscus sellowii* (Brandt, 1833) is a common species in Southern Brazil, Uruguay and the region around Buenos Aires in Argentina (Schmalfuss, 2003).

The specimens of *B. sellowii* were collected in a urban area of Porto Alegre, Rio Grande do Sul, southern Brazil. They were kept in laboratory conditions at $20\pm 1^{\circ}\text{C}$, with 12:12 (light: dark) photoperiod. Only animals heavier than 25mg were used in the experiments, excluding ovigerous females.

The source-site consisted of an area where animals were abundant and there were trees characteristic of pioneer vegetation colonization. The three most abundant plant species in the site were *Lithraea brasiliensis* Marchand (Anacardiaceae); *Ricinus communis* Linnaeus (Euphorbiaceae) and *Schinus terebinthifolius* Raddi (Anacardiaceae).

Feeding trials

Leaves from the most abundant plants of the local were offered to isopods to verify feeding preference based on consumption rate. Green leaves from three different plant species were taken from branches and placed into litterbags for decomposition in loco for 14 days. After this period, the leaves were taken to the laboratory and circles of 18mm of diameter were cut and oven dried at 60°C for 48 hours. The discs were weighed (GIBERTINI E425-B) and remoisten before offered to animals for one week period in individual units consisted of 8cm diameter plastic containers with moist plaster of Paris in the bottom and a net over it to minimize coprophagy (Fig. 1). Animals were kept without food source for two days to empty

gut prior to the experiment. After the experiment, the remaining plant material and feces were oven dried and reweighed, and consumption rates calculated. The control consisted of units containing leaves and no animals. The mean percentage of leaf weight lost due to autogenic changes (weight lost independent from the action of consumers) was subtracted from the amount of plant consumed. The plant species that presented the highest consumption was used to verify feeding performance in different stages of leaf decomposition as well as the phenolic and flavonoid content of leaves.

Feeding performance on leaves in different stages of decomposition

Green leaves were collected from branches in the same site and placed into 20 litterbags (10x15cm) fastened to the soil with small wooden sticks. Litterbags were collected after one, two and three months of decomposition, taken to laboratory and offered to animals, as well as additional green leaves collected from branches when litter bags were placed in the soil. Oven dried leaves from each decomposition stage as well as the remains were stored under refrigeration for phenolic and flavonoid content analysis.

Consumption, egestion and assimilation rates

Experimental units as described above were used in the experiment. Two or three discs of 18mm (or approximate amount for the third month of decomposition, which presented leaves that were too friable to be cut into discs) were oven dried at 60° for 48 hours, weighted, remoisten with distilled water, and offered to animals for ten days. Animals were kept without food two days prior and after the experiment to empty gut. Remaining leaves and feces were collected from units, oven dried and weighted after the experiment to calculate feeding performance. Twenty repetitions and 20 controls were used for each stage of decomposition.

The consumption rates were calculated as mg of ingested leaves (subtracted mean percentage of autogenic losses) in dry weight (DW) per g of body in fresh weight (FW), per day. The egestion rates were calculated as mg of produced feces (DW) per g of body weight (DW) per day. The assimilation rates were calculated as mg of ingested leaves (DW) minus mg of produced feces (DW) per g of body weight (FW) per day (adapted from Souza et al, 1998; Loureiro et al., 2006).

Time of passage through gut and assimilation efficiency

To calculate the assimilation efficiency, the time food was retained in the gut needed to be tested. Animals were kept in individual units containing carrot as a food source for a week. The carrot was used as a marker provided that the fecal pellets derived from its feeding differ in color (Fig. 2). After that, ten animals were exposed to one disc of leaf litter from each decomposition stage, and monitored every two hours for 80 hours. The time that the leaf first appeared fed upon and the time the first feces from feeding on leaves were present in experimental units were recorded.

Animals were fed on carrots for a week and placed in units containing one leaf disc until all of the disc was eaten. Feces were collected, oven dried and weighted to calculate assimilation efficiency. Five units from each decomposition stage were monitored daily and time for the beginning of feeding start was also recorded. The assimilation efficiency was calculated as percentage using mg of ingested leaf minus mg of feces produced per mg of ingested leaf.

Secondary metabolites content

Secondary metabolites were related to content of total phenolics and flavonoids. Total phenolics were determined following Bärlocher and Graça (2005), using tannic acid as standard. Five samples of approximately 100mg of dry leaves were used for each stage of decomposition, totalizing 20 samples.

Determination of the flavonoid content was modified from Zhishen et al. (1999) using quercetin as pattern and five discs of dry leaves or approximate amount ($\approx 150\text{mg}$) for each stage of decomposition, also totalizing 20 samples.

The mean concentration of phenolics and flavonoids were multiplied by the consumption rates and compared.

Statistical analysis

All data was tested for normality using Kolmogorov-Smirnov test. The consumption, egestion and assimilation rates for each stage of decomposition were compared through one-way Analysis of Variance (ANOVA) followed by Tukey test. Pearson correlations were used to verify association between consumption and egestion rates among treatments. The statistical analyses were performed using InStat 3.01 software.

RESULTS

Feeding preference

The consumption rate was significantly higher when animals fed on *S. terebinthifolius* ($52.891 \pm 9.000 \text{ mg g}^{-1} \text{ day}^{-1}$) ($F_{2,26}=9.395$; $p < 0.001$) and no significant difference was recorded when animals fed on *L. brasiliensis* ($31.749 \pm 3.959 \text{ mg g}^{-1} \text{ day}^{-1}$) and *R. communis* ($15.453 \pm 2.867 \text{ mg g}^{-1} \text{ day}^{-1}$) (Fig 3). The egestion and assimilation rates for *S. terebinthifolius* were $46.290 \pm 9.761 \text{ (mg g}^{-1} \text{ day}^{-1})$ and $11.230 \pm 1.948 \text{ (mg g}^{-1} \text{ day}^{-1})$, respectively. For *L. brasiliensis*, the rates were 21.130 ± 2.595 and $14.845 \pm 1.623 \text{ (mg g}^{-1} \text{ day}^{-1})$ respectively, and for *R. communis* the amount of fecal pellets were too low to be weighted and therefore egestion rate could not be calculated, and the assimilation rate was $15.453 \pm 2.867 \text{ (mg g}^{-1} \text{ day}^{-1})$.

Schinus terebinthifolius in different decomposition stages

The consumption rate was significantly higher on two months-old leaves ($F_{3,58}=8.59$; $p < 0.001$) and there were no significant differences between green, one month-old and three months-old leaves. The egestion rate also presented significant difference, being highest with two months-old leaves ($F_{3,58}=14.171$; $p < 0.05$). There were no significant difference, neither between green and one month-old leaves, nor between one month-old and three months-old leaves. The assimilation rate was significantly highest when the consumption rate was lowest (green leaves) and presented no significant difference between one, two and three months-old leaves ($F_{3,58}=5.302$; $p < 0.005$) (Table 1).

There were significant correlations between consumption and egestion rates for all decomposition stages. The correlation was stronger for three months-old leaves ($r^2=0.9258$; $p < 0.0001$) followed by one month-old leaves ($r^2=0.8959$;

$p < 0.0001$), two months-old leaves ($r^2 = 0.8346$; $p < 0.0001$) and green leaves being the weakest ($r^2 = 0.7241$; $p < 0.0002$) (Fig. 4).

The time of passage through gut did not present difference among stages of decomposition. In all stages, the mode time for appearance of feces from leaf consumption was two hours. However, the beginning of ingestion of leaves occurred in the same day as the beginning of the experiment for most units from one, two and three months of decomposition, whereas the beginning of ingestion of green leaves occurred after two or three days. For three out of the ten units containing green leaves, the consumption was not noticed after 80h of experiment.

The calculated assimilation efficiency was $43.06 \pm 3.84\%$ for green leaves ($n=6$), $19.74 \pm 2.39\%$ for one month-old leaves ($n=4$), 20.33 ± 1.55 for two months-old leaves ($n=5$) and $19.51 \pm 2.78\%$ for three months-old leaves ($n=2$).

The phenolic content was highest in green leaves (0.0336 ± 0.0019 mg of tannic acid equivalent per mg of dry leaf) and very low after three months of decomposition (0.0035 ± 0.0002 mg mg^{-1}) ($F_{3,15} = 35.259$; $p < 0.0001$). There was no significant difference between one and two months-old leaves (0.0226 ± 0.00144 and 0.0202 ± 0.0043 mg mg^{-1} respectively). The flavonoid content was significantly higher in green leaves and lowest in the second month ($F_{3,16} = 37.31$; $p < 0.001$). There was no significant difference between the content of the first and third month of decomposition (Fig. 5).

The estimated phenolic amount ingested by animals was significantly different ($F_{3,58} = 28.934$; $p < 0.0001$), being highest when consumption rate was also highest. The estimated flavonoid amount ingested by the animals presented the same trend as the assimilation rate among the stages of decomposition, being significantly

higher in green leaves but not differing between one, two, and three months of decomposition ($F_{3,58}=10.252$; $p<0.0001$) (Fig. 6).

As the flavonoid content was not tested for every experimental unit, the mean average of the content of each month was tested for correlation with the mean average of the assimilation rate for each stage of decomposition, resulting in a high correlation ($r^2=0.9907$; $p=0.0046$, $n=4$).

DISCUSSION

Numerous studies analyze the effects of secondary metabolites in herbivores, but few studies have been conducted to understand the role of these compounds in detritivore and decomposer organisms, besides the presence of unpalatable or indigestible compounds (Cameron and LaPoint, 1978; Hassall and Rushton, 1984; Target et al., 1986; Canhoto and Graça, 1999) and their feeding preference related to leaf senescence (Cameron and LaPoint, 1978; Yeates and Barmuta, 1999). Our study site presented plant species characteristic of a successional stage that do not present mechanical structures to avoid herbivores other than lignin, suggesting that chemical defenses are key for plant protection. Tropical plants inhabiting resource-poor environments seem to invest heavily in chemical defenses such as various phenolics (Agrawal, 2006). *Ricinus communis* presented the lowest consumption rate and very little amount of fecal pellets egested. This is a plant that has been associated with a large amount of secondary metabolites, being gallic acid, quercetin, and rutin some of the major phenolic compounds responsible for the antioxidant activity of its dry leaves (Singh et al., 2009) and its decomposition after two weeks resulted in viscous leaves.

Lithraea brasiliensis also presents a large amount of secondary metabolites (Correia et al., 2006) and it is an easily found leaf in the leaf litter in Brazil although its consumption by *B. sellowii* was significantly lower than *S. terebinthifolius* and did not differ in consumption when compared to *R. communis*.

Schinus terebinthifolius is known as a source of terpenoids, simple phenolic derivatives and flavonols and the anti-oxidant activity of its aerial parts extract has been described (Velázquez et al., 2003). Leaf extracts contain triterpene acids

(Campelo and Marsaioli, 1974) and the ethanolic extract of the leaves is a source of simple phenolics, several flavonoids (flavones flavanones leucoanthocyanidins), xanthenes and free steroids (Lima et al., 2006). After the beginning of decomposition, leaves start to curl and might be used by animals for shelter as well as for feeding.

There has been controversy on results of assimilation efficiency on high and low quality food. Authors have hypothesized that some foods might present high assimilation efficiency due to a slower passage through the gut, which decreases fecal production (Souza et al., 1998; Loureiro et al., 2006). In this experiment, the assimilation efficiency in the experimental units were also high for green leaves, which would be considered a low quality food, and there was no difference in the time for beginning of appearance of feces from leaves in this treatments. However, calculating the assimilation efficiency by total consumption and egestion of a leaf disc with a known mass, the results were more plausible. The latter food source makes the animal empty gut from the former ingested food source. The assimilation efficiency was still higher when animals fed on green leaves, but the value was under 80%. Leaves with one, two and three months of decomposition presented similar assimilations efficiency, as well as assimilation rates. The assimilation rates calculated from total consumption of a disc of leaf with a known mass did not differ as much from the rates calculated in the units of the first experiment. The assimilation efficiency should reduce over time (Rushton and Hassall, 1983, Hassall and Rushton, 1984) due to lower content of metabolites in leaves making them more palatable (Johnson and Feldon, 2001)

Green leaves presented high phenolic and flavonoid concentration taking a long time for the animals to begin feeding, suggesting that they might present

substances that are feeding deterrents and therefore reduce palatability. These phenolic substances are probably lost in the beginning of the decomposition process due to leaching, once the same time for the beginning of feeding was not observed in the other decomposition stages. If substances that cause feeding deterrence and inhibit feeding are lost right after the start of leaf senescence, the faster they will be consumed by detritivores returning to the soil as nutrient and therefore being also an adaptive advantage to the plants. Cameron and LaPoint (1978) observed a rapid leach of tannins that was associated with food inhibition from litter bags in the first week of decomposition.

The green leaves used in this study were oven dried before offered to animals, possibly changing properties and making them more palatable to animals. When given a choice, animals avoid green leaves that present high amounts of secondary compounds (Cameron and LaPoint, 1978). Ingesting decayed leaves is a behavioral strategy used to cope with chemically defended food. This way, the concentration of defensive chemicals is reduced and animals may use this strategy to increase their tolerance to chemically defended food (Glendinning, 2007) as recorded by Roy and Bergeron (1990) observing a small rodent cutting leaves from branches and waiting for some decomposition before ingesting it.

Switching frequently between different types of food and regulating intake of specific defensive chemicals are also behavioral mechanisms to handle chemically defended food (Glendinning 2007). Isopods ingest decayed plant material, and ingest different types of plants in their diet when given a chance (Wieser, 1984). However, experiments of feeding preference between more than two food sources are difficult to analyze (Wieser, 1984; Peterson and Renaud, 1989), and therefore preference was not tested here.

During leaf senescence, the content of secondary metabolites tends to diminish due to leaching (Kuiters and Sarink, 1986). Phenolics and flavonoids in plants has mostly been related to defense against pathogens and herbivores (Dixon and Steele, 1999; Boué and Raina, 2003) and leaves presenting a lower content of total phenolics would be less toxic and more used by isopods once ingested food does not change much in the digestory tract. Hassall and Rushton (1984) predicted that less heavily defended species would reach optimal palatability earlier than climax species that usually present more secondary metabolites for defense. However, what determines detritivore consumption is thought to be phenolic signature rather than total phenolic content (Zimmer et al., 2005), and the optimal palatability occurred on two months-old leaves and not when the phenolic content was lowest.

The flavonoid represented a large portion of the total phenolics in the green, one month-old and two months-old leaves. However, the amount of flavonoids in the third month exceeded the amount of total phenolics. That might have happened due to decomposition of the substance during its extraction or due to sampling. Phenolics can be converted into lignin that is broken down by microorganisms; these fragments contribute to the mineralization of soil and humus formation (Kefeli et al., 2003). Lignin phenolics cannot be isolated from lignocellulose without partial denaturation (Breznak and Brune, 1994, Zimmer, 1999a) while flavonoids are non-lignin phenolics (Brisson et al., 1986) and might not have suffered denaturation during analyses, and therefore presented a higher content than the total phenolic content in the third month.

Leaching of substances also occurs in other materials in the litter, and might increase the content of a specific constituent originated from the litter itself (from

absorption) or from the action of microorganisms. In this experiment, the phenolic content decreased throughout the months of the experiment, but the flavonoid content increased in the last month instead of decreasing in relation to the leaves of the previous month.

Although the flavonoid content differed between treatments, the estimated amount of flavonoid consumed by animals was almost the same for leaves with one, two and three months of decomposition. There was a strong correlation between the flavonoid content and the assimilation rate but no significant correlation with the total phenolic content. Neuhauser and Hartenstein (1978) did not find any correlation between phenolic content of leaves and feeding preference. However, it seems that animals increase consumption in order to ingest a minimum amount of flavonoid, suggesting that they might use these flavonoids as a food parameter. The content of phenolics and flavonoids in plants has been mostly related to defense against pathogens and herbivores and only a few times possible benefits for organisms ingesting those substances have been mentioned. Johnson and Felton (2001) using herbivores, also observed a lower consumption and digestibility on plants that were overexpressing phenolics, but no significant reduction on growth and no indications of oxidative stress as a causal factor, suggesting that a beneficial antioxidant property for herbivores. These researchers obtained the same results as this study and suggested that the animals might be using the flavonoids as an antioxidant agent.

Even though the use of flavonoids by herbivore invertebrates is not well documented, this paper suggests that the isopods may also use and discriminate concentrations of flavonoids at some degree since they seem to have increased consumption to maintain a minimum intake of flavonoids in leaves with lower

flavonoid content. To verify the plausibility of this hypothesis, experiment must be conducted in order to understand whether or not animals can discriminate different concentrations of flavonoids and if the true ingestion of this substance is equivalent to the estimate presented in this study.

ACKNOWLEDGEMENTS

The authors wish to thank to CNPq for granting of the scholarship to CTW and research fellowship to PBA. We are also grateful to Priscila Bugs and Carina Appel for helping in the field.

REFERENCES

- Agrawal, A. A. 2006. Macroevolution of plant defense strategies. *Trends in Ecology and Evolution* 22(2): 103-109.
- Appel, H. M. 1993. Phenolics in ecological interactions: the importance of oxidation. *Journal of Chemical Ecology* 19(7): 1521-1552.
- Bärlocher, F., and Graça, M. A. S. 2005. Total phenolics, pp. 97-100. In, Graça, M.A.S., Bärlocher, F., and Gessner, M. O. (eds.), *Methods to Study Litter Decomposition: A Practical Guide*, Dordrecht.
- Boué, S. M., and Raina, A. K. 2003. Effects of plant flavonoids on fecundity, survival, and feeding of the Formosan subterranean termite. *Journal of Chemical Ecology* 29(11): 2575-2584.
- Brandt, I. 1833. *Conspectus Monographiae Crustaceorum Oniscodorum Latreillii*. – *Byulleten moskovskogo Obshchestva Ispýtatelei Prirodÿ* 6: 171-193 and plate 4.
- Breznak, J. A., and Brune, A. 1994. Role of microorganisms in the digestion of lignocellulose by termites. *Annual Review of Entomology* 39: 453–487
- Brisson, L., Vacha, W. E. K., and Ibrahim, R. K. 1986. Localization of partially methylated flavonol glucosides in *Chrysosplenium americanum* – II. Immunofluorescence. *Plant Science* 44: 175–181
- Cameron, G. N., and LaPoint, T. W. 1978. Effects of tannins on the decomposition of Chinese tallow leaves by terrestrial and aquatic invertebrates. *Oecologia* 32: 349-366.
- Campelo, J., and Marsaioli, A. J. 1974. Triterpenes of *Schinus terebinthifolius*. *Phytochemistry* 13: 659–660.

- Canhoto, C. M. and Graça, M. A. S. 1999. Leaf barriers to fungal colonization and shredders (*Tipula lateralis*) consumption of decomposing *Eucalyptus globules*. *Microbial Ecology* 37: 163-172.
- Correia, S. J., David, J. P., and David, J. M. 2006. Metabólitos secundários de espécies de Anacardiaceae. *Química Nova* 29(6):1287-1300.
- Dixon, R. A., and Steele, C. L. 1999. Flavonoids and isoflavonoids – a gold mine for metabolic engineering. *Trends in plant science* 4(10): 394-400.
- Glendinning, J. I. 2007. How do predators cope with chemically defended foods? *Biological Bulletin* 213: 252-266.
- Gould, K. S., and Lister, C. 2006. Flavonoid function in plants, pp.397-441. In, Andersen, Ø. M., and Markham, K. R. (eds), *Flavonoids: Chemistry, biochemistry and applications*. Taylor & Francis Group, Boca Raton.
- Graça, M. A. S.; Cressa, C.; Gessner, M. O.; Feio, M. J.; Callies, K. A., and Barrios, C. 2001. Food quality, feeding preferences, survival and growth of shredders from temperate and tropical streams. *Freshwater Biology* 46: 947-957.
- Gryglewski, R. J., Korbut, R., Robak, J., and Świąć, J. 1987. On the mechanism of antithrombotic action of flavonoids. *Biochemical Pharmacology* 36(3): 317-322.
- Gunnarsson, T. 1987. Selective feeding on a maple leaf by *Oniscus asellus* (Isopoda). *Pedobiologia* 30: 161-165.
- Harborne, J. B. 1993. *Introduction to ecological biochemistry*. Fourth edition. University Press. 318p.
- Hassall, M., and Rushton, S. P. 1984. Feeding behaviour of terrestrial isopods in relation to plant defences and microbial activity. *Symposium of the Zoological Society of London* 53: 487-505.

- Hassall, M.; Turner, J. G. and Rands, M. R. W. 1987. Effects of terrestrial isopods on the decomposition of woodland leaf litter. *Oecologia* 72: 597-604.
- Ihnen, K., and Zimmer, M. 2008. Selective consumption and digestion of litter microbes by *Porcellio scaber* (Isopoda: Oniscidea). *Pedobiologia* 51: 335-342.
- Johnson, K. S., and Felton, G. W. 2001. Plant phenolics as dietary antioxidants for herbivores insects: a test with genetically modified Tobacco. *Journal of Chemical Ecology*, 27(12): 2579-2597.
- Kainulainen, P. and Holopainen, J. K. 2002. Concentration of secondary compounds in Scots pine needles at different stages of decomposition. *Soil Biology and Biochemistry* 34: 37-42.
- Kasurinen, A., Peltonen, P. A., Julkunen-Tiitto, R., Vapaavuori, E., Nuutinen, V., Holopainen, T., and Holopainen, J. K. 2007. Effects of elevated CO₂ and O₃ on leaf litter phenolics and subsequent performance of litter-feeding soil macrofauna. *Plant Soil* 292: 25-43.
- Kefeli, V. I., Kalevitch, M. V., and Borsari, B. 2003. Phenolic cycle in plants and environment. *Journal of Cell and Molecular Biology* 2: 13-18.
- Kuiters, A. T., and Sarink, H. M. 1986. Leaching of phenolic compounds from leaf and needle litter of several deciduous and coniferous trees. *Soil Biology and Biochemistry* 18(5): 475-480.
- Lambdon, P. W., and Hassall, M. 2005. How should toxic secondary metabolites be distributed between the leaves of a fast-growing plant to minimize the impact of herbivory? *Functional Ecology* 19: 299-305.
- Latreille, P. 1804. Histoire naturelle, générale et particulière, des crustacés et des insectes. *Cloportides*, 7: 25-49. Paris.

- Lima, M. R. F., Luna, J. S., Santos, A. F.; Andrade, M. C. C., Sant'Ana, A. E. G., Genet, J.-P., Márquez, B., Neuville, L., and Moreaub, N. 2006. Anti-bacterial activity of some Brazilian medicinal plants. *Journal of Ethnopharmacology* 105:137–147.
- Loureiro, S., Sampaio, A., Brandão, A., Nogueira, J. A., and Soares, A. M. V. M. 2006. Feeding behaviour of the terrestrial isopod *Porcellionides pruinosus* Brandt, 1833 (Crustacea, Isopoda) in response to changes in food quality and contamination. *Science of the Total Environment* 369: 119-128.
- Neuhauser, E. E., and Hartenstein, R. 1978. Phenolic content and palatability of leaves and wood to soil isopods and diplopods. *Pedobiologia* 18(8): 99-109.
- Oberdörster, E., Clay, M. A., Cottam, D., M. Wil mot, F. A., McLachlan, J. A., and Milner, M. 2001. Common phytochemical are acdysteroid agonists and antagonists: a possible evolutionary link between vertebrate and invertebrate steroid hormones. *Journal of Steroid Biochemistry & Molecular Biology* 77: 229-238.
- Peterson, C. H., and Renaud, P. E. 1989. Analysis of feeding preference experiments. *Oecologia* 80:82-86.
- Quadros, A. F., and Araujo, P. B. 2008. An assemblage of terrestrial isopods (Crustacea) in southern Brazil and their contribution to leaf litter processing. *Revista Brasileira de Zoologia* 25(1): 58-66.
- Roy, J., and Bergeron, J. M. 1990. Branch cutting behavior by the vole (*Microtus pennsylvanicus*): a mechanism to increase toxicity of secondary metabolites in conifers. *Journal of Chemical Ecology* 16: 735-741.
- Rushton, S. P., and Hassall, M. 1983. Food and feeding ratios of the terrestrial isopod *Armadillidium vulgare* (Latreille). *Oecologia* 57: 415-419.

- Salgado, P. R., Favarin, J. L., Leandro, R. A., and Lima Filho, O. F. 2008. Total phenol concentrations in coffee tree leaves during fruit development. *Science and Agriculture* 65(4): 354-359.
- Schmalfuss, H. 2003. World catalog of terrestrial isopods (Isopoda: Oniscidea). Serie A, Nr. 654. *Stuttgarter Beiträge zur Naturkunde*. 341p.
- Simmonds, M. S. J. 2001. Importance of flavonoids in insect-plant interaction: feeding and oviposition. *Phytochemistry* 56(3): 245-252.
- Singh, P. P., Ambika, and Chauhan, S. M. S. 2009. Activity guided isolation of antioxidants from the leaves of *Ricinus communis* L. *Food Chemistry* 114: 1069–1072.
- Sousa, J. P., Vingada, J.,V., Loureiro, S., Gama, M.,M., and Soares, A. M. V. M. 1998. Effects of introduced exotic tree species on growth, consumption and assimilation ratios of the soil detritivore *Porcellio dilatatus* (Crustacea: Isopoda). *Applied Soil Ecology* 9: 399-403.
- Stevenson, J. R. 1961. Polyphenol oxidase in the tegumental glands in relation to the molting cycle of the isopod crustacean *Armadillidium vulgare*. *Biological Bulletin* 121: 554-560.
- Szlávecz, K., and Pobožny, M. 1995. Coprophagy in isopods and diplopods: a case for indirect interaction. *Acta Zoologica Fennica* 196: 124-128.
- Target, N. M., Target, T. E., Vrolijk, N. H., and Ogden, J.C. 1986. The effect of macrophyte secondary metabolites on feeding preferences of the herbivorous parrotfish *Sparisma radians*. *Marine Biology* 92: 141-148.
- Van Wensem, J., Verhoef, H. A., and Van Straalen, N. M. 1993. Litter degradation stage as a prime factor for isopod interaction with mineralization process. *Soil Biology and Biochemistry* 25(9): 1175-1183.

- Velázquez, H. A., Tournier, P. M. B., Saavedra, G., and Schinella, G. R. 2003. Antioxidant activity of Paraguayan plant extracts. *Fitoterapia*, 74:91–97.
- Wieser, W. 1984. Ecophysiological adaptations of terrestrial isopods: a brief review. *Symposium of the Zoological Society of London* 53: 247-265.
- Yeates, L. V., and Barmuta, L. A. 1999. The effects of willow and eucalypt leaves on feeding preference and growth of some Australian aquatic macroinvertebrates. *Australian Journal of Ecology* 24: 593-598.
- Zhishen, .J, Mengcheng, T., and Jianming, W. 1999. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry* 64: 555-559.
- Zimmer, M. 1999a. Combined methods for the determination of lignin and cellulose in leaf litter. *Sciences of Soils*, 4:2
- Zimmer, M. 1999b. The fate and effects of ingested hydrolysable tannins in *Porcellio scaber*. *Journal of Chemical Ecology* 25: 611-628.
- Zimmer, M. 2002a. Is decomposition of woodland leaf litter influenced by its species richness? *Soil Biology and Biochemistry* 24: 277-284.
- Zimmer, M. 2002b. Nutrition in terrestrial isopods (Isopoda: Oniscidea): an evolutionary-ecological approach. *Biology Review* 77: 455-493.
- Zimmer, M., and Topp, W. 1998. Do woodlice (Isopoda: Oniscidea) produce endogenous cellulases? *Biology and Fertility of Soils* 26: 155-156.
- Zimmer, M., Danko, J. P., Pennings, S. C., Danford, A. R., Carefoot, T. H., Ziegler, A., and Uglow, R. F. 2002. Cellulose digestion and phenol oxidation in coastal isopods (Crustacea: Isopoda). *Marine Biology* 140: 1207-1213.

- Zimmer, M., Kautz, G., and Topp, W. 2003. Leaf litter-colonized microbiota: supplementary food source or indicator of food quality for *Porcellio scaber* (Isopoda: Oniscidea)? *European Journal of Soil Biology* 39: 209-216.
- Zimmer, M., Oliveira, R., Rodrigues, E., and Grança, M. A. S. 2005. Degradation of leaf litter phenolics by aquatic and terrestrial isopods. *Journal of Chemical Ecology* 31(8): 1933-1952.

Table 1. Feeding performance of *Balloniscus sellowii* on *Schinus terebinthifolius* in different stages of decomposition. Data are expressed as mean value and standard error. The superscript letters indicate significant difference among treatments ($p < 0.05$).

Stage of decomposition	Consumption rate (mg g ⁻¹ day ⁻¹)	Egestion rate (mg g ⁻¹ day ⁻¹)	Assimilation rate (mg g ⁻¹ day ⁻¹)
Green leaves	41.465±5.129 ^a	10.742 ± 3.196 ^a	30.722±2.936 ^a
1 month old leaves	50.185±5.396 ^a	29.136 ± 5.669 ^{a,b}	21.049±1.829 ^b
2 months old leaves	80.147±6.220 ^b	61.297 ± 6.414 ^c	18.851±2.633 ^b
3 months old leaves	53.391±5.516 ^a	33.440 ± 5.068 ^b	19.951±1.522 ^b

LIST OF FIGURES

- Figure 1.** Experimental units with moist plaster of Paris in the bottom and a net over it to minimize coprophagy containing *Balloniscus sellowii* and a disc of leave from *Schinus terebinthifolius*.....43
- Figure 2.** Fecal pellets from the terrestrial isopod *Balloniscus sellowii*. a) fecal pellet from the consumption of carrots and b) fecal pellet from the consumption of leaves.....45
- Figure 3.** Isopod feeding performance on leaves of different plant species with 14 days of decomposition in litter bags left to decompose in loco. The egestion rate for *Ricinus communis* could not be calculated due to low amount of fecal pellets. The values are mean and SE. Superscript letters indicate significant difference among treatments ($p < 0.05$).....47
- Figure 4.** Pearson correlations between consumption and egestion rates of *Balloniscus sellowii* on *Schinus terebinthifolius* leaves in different decomposition stages.....49
- Figure 5.** Total phenolic and flavonoid content in leaves of *Schinus terebinthifolius* in different stages of decomposition. The values are mg of equivalent of quercitin (flavonoid) or tannic acid (phenolic) per mg of dry leaf SE. Superscript letters indicate significant differences among treatments ($p < 0.05$).....51
- Figure 6.** Estimated amount of total phenolics and flavonoids ingested by *Balloniscus sellowii* in leaves of *Schinus terebinthifolius* in different stages of decomposition. The values are mean and SE. Superscript letters indicate significant differences among treatments ($p < 0.05$).....53

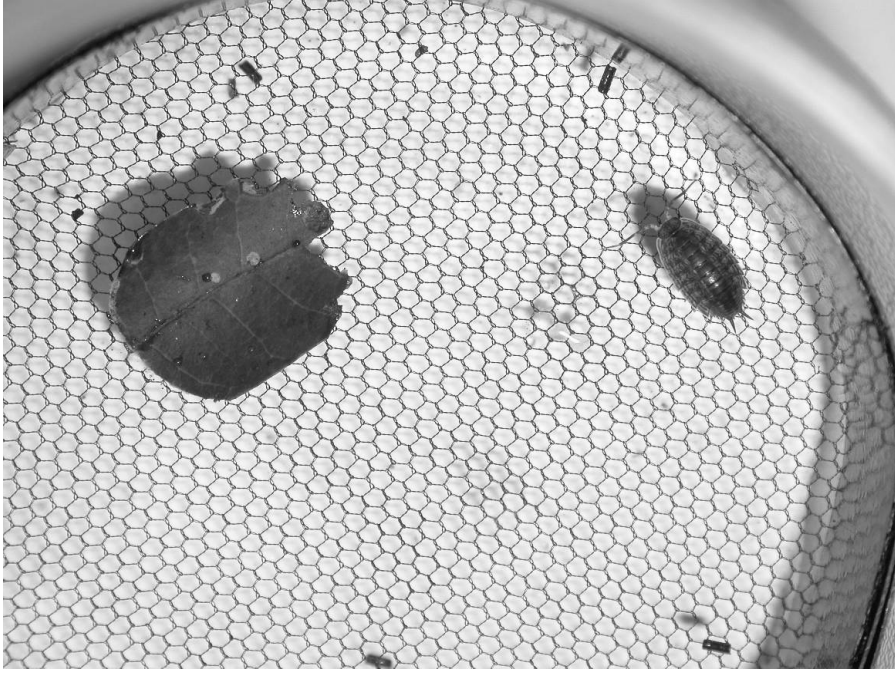


Figure 1. Experimental units with moist plaster of Paris in the bottom and a net over it to minimize coprophagy containing *Balloniscus sellowii* and a disc of leave from *Schinus terebinthifolius*.

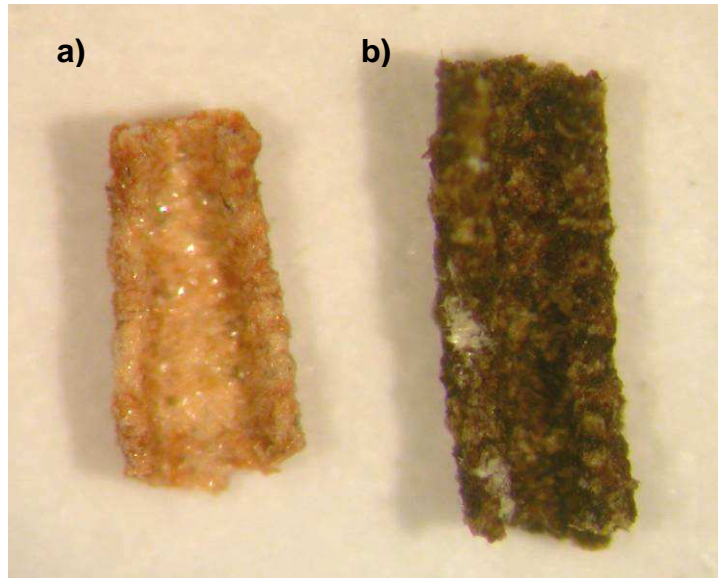


Figure 2. Fecal pellets from the terrestrial isopod *Balloniscus sellowii*. a) fecal pellet from the consumption of carrots and b) fecal pellet from the consumption of leaves.

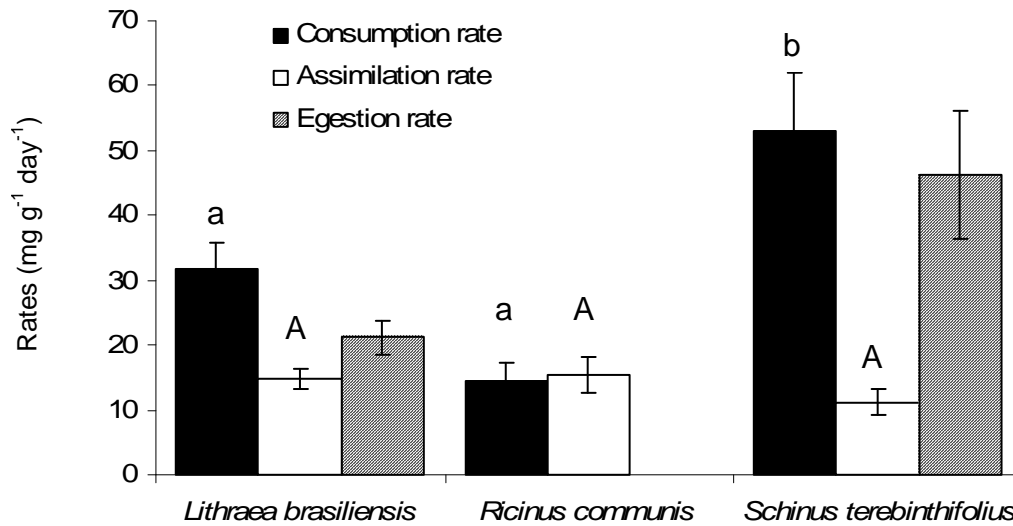


Figure 3. Isopod feeding performance on leaves of different plant species with 14 days of decomposition in litter bags left to decompose in loco. The egestion rate for *Ricinus communis* could not be calculated due to low amount of fecal pellets. The values are mean and SE. Superscript letters indicate significant difference among treatments ($p < 0.05$).

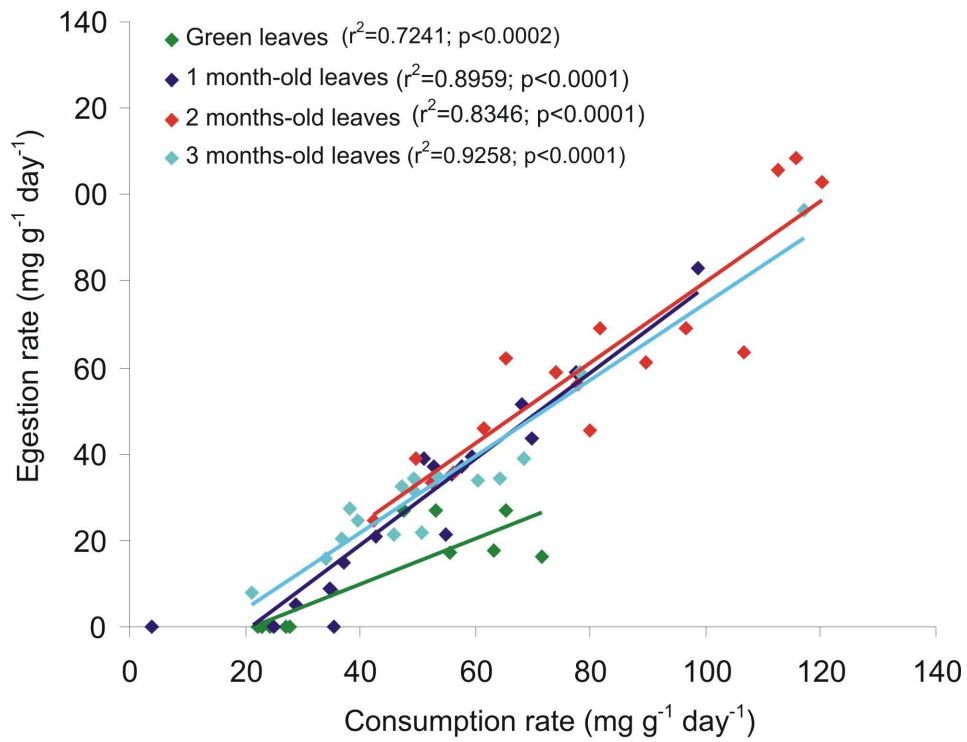


Figure 4. Pearson correlations between consumption and egestion rates of *Balloniscus sellowii* on *Schinus terebinthifolius* leaves in different decomposition stages.

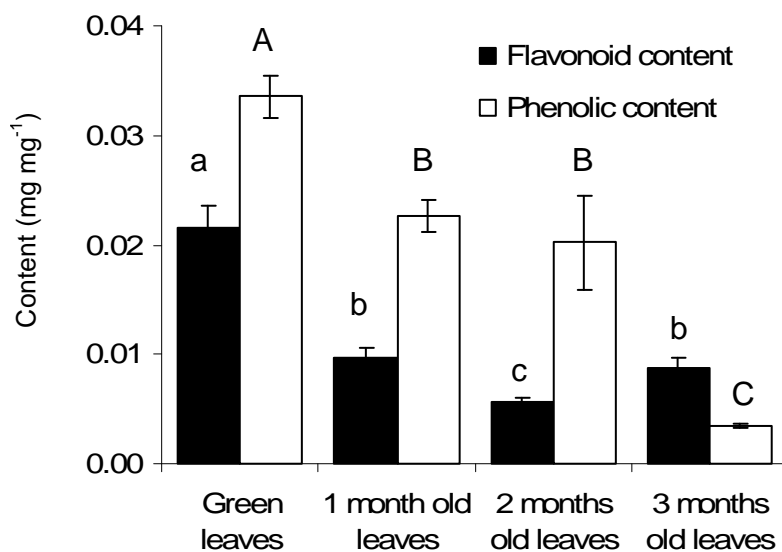


Figure 5. Total phenolic and flavonoid content in leaves of *Schinus terebinthifolius* in different stages of decomposition. The values are mg of equivalent of quercetin (flavonoid) or tannic acid (phenolic) per mg of dry leaf SE. Superscript letters indicate significant differences among treatments ($p < 0.05$).

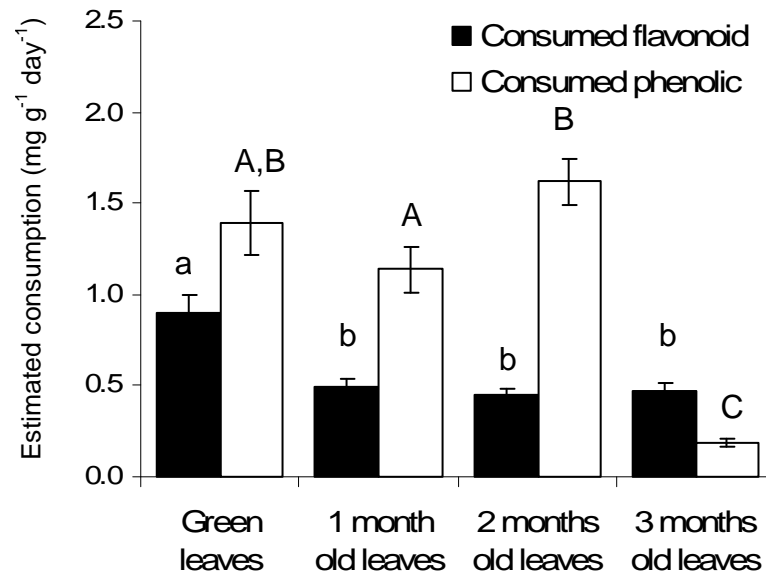


Figure 6. Estimated amount of total phenolics and flavonoids ingested by *Balloniscus sellowii* in leaves of *Schinus terebinthifolius* in different stages of decomposition. The values are mean and SE. Superscript letters indicate significant differences among treatments ($p < 0.05$).

CONSIDERAÇÕES FINAIS

Diversos trabalhos têm sido realizados utilizando isópodos terrestres como modelo para entender a nutrição de detritívoros embora não muitos discutam a relação dos metabólitos secundários.

Nesse trabalho, observou-se a relação do conteúdo de fenólicos presentes em uma planta durante sua decomposição e a sua relação com o desempenho alimentar do isópodo terrestre *Balloniscus sellowii*. Dentro dos fenólicos, o conteúdo de flavonoides teve uma relação ainda mais forte, sendo a quantidade de flavonoides ingeridos em folhas de um, dois e três meses de decomposição muito similares e sem diferença significativa. Para isso, a taxa de consumo foi mais alta quando o conteúdo de flavonoides das folhas era mais baixo. Com esse resultado, surgiu uma dúvida e uma proposta de um novo experimento a fim de testar se os animais possuem a capacidade de discriminar diferentes concentrações de flavonoides. O experimento consistirá de um disco de gelatina com diferentes concentrações de flavonoides oferecido aos animais. Desta forma, poderemos calcular mais precisamente a quantidade de flavonoides ingeridos em um período de tempo e qual a concentração ótima e inibitória de flavonoides para os animais.

Sabe-se que além do conteúdo de metabólitos secundários, diversas características das folhas são de grande importância na escolha e preferência dos animais sendo, a dureza, também importante na escolha e aproveitamento do recurso. Esta característica leva-nos a outro questionamento, relacionado ao desgaste das mandíbulas. Assim, outro experimento será conduzido, utilizando-se discos foliares dos diferentes estágios de decomposição utilizados no experimento (armazenados em refrigeração), os quais serão re-umedecidos com água destilada

e oferecidos a animais que acabaram de realizar a ecdise. Esses animais receberão discos daquele estágio de decomposição até começarem a armazenar cálcio na forma de placas de cálcio, o que indica que estão se preparando para realizar nova ecdise, e serão acondicionados em álcool 70%. Os animais serão dissecados para a retirada das mandíbulas e preparados para microscopia eletrônica de varredura (MEV). A cada ecdise, os animais trocam as peças bucais e com isso saberemos que aquele desgaste foi devido à alimentação oferecida no tratamento. As imagens das mandíbulas serão analisadas e o desgaste dos diferentes tratamentos comparados.

A análise dos dados do trabalho permitiram a proposição de experimentos que possam confirmar as hipóteses levantadas na discussão. Desta forma, a sua continuação torna-se importante para uma complementação deste trabalho.

REFERÊNCIAS BIBLIOGRÁFICAS

- Appel, H. M. 1993. Phenolics in ecological interactions: the importance of oxidation. *Journal of Chemical Ecology* 19(7): 1521-1552.
- Araujo, P. B., and Zardo, M. C. L. 1995. Uma nova espécie de *Balloniscus* Budde-Lund (Crustacea, Isopoda, Balloniscidae) do sul do Brasil. *Revista Brasileira de Zoologia* 12(4): 785-790.
- Brandt, I. 1833. *Conspectus Monographiae Crustaceorum Oniscodorum Latreillii*. – *Byulleten moskovskogo Obshchestva Ispytatelei Prirody* 6: 171-193 and plate 4.
- Budde-Lund, G. 1908. Isopoda von Madagaskar und Ostafrika mit Diagnosen verwandter Arten. In, Voeltzkow, A., *Reise in Ostafrika in den Jahren 1903-1905. Wissenschaftliche Ergebnisse*, vol. 2: 265-308 and plates 12-18.
- Gunnarsson, T. 1987. Selective feeding on a maple leaf by *Oniscus asellus* (Isopoda). *Pedobiologia* 30: 161-165.
- Gunnarsson, T. and Tunlid, A. 1986. Recycling of fecal pellets in isopods: microorganisms and nitrogen compounds as potential food for *Oniscus asellus* L. *Soil Biology and Biochemistry* 18: 595-600.
- Harborne, J. B. 1993. *Introduction to ecological biochemistry*. Fourth edition. University Press. 318p.
- Hassall, M., and Dangerfield, J. M. 1990. Density-dependent processes in the population dynamics of *Armadillidium vulgare* (Isopoda: Oniscidae). *Journal of Animal Ecology* 59: 941-958.
- Hassall, M., and Rushton, S. P. 1982. The role of coprophagy in the feeding strategies of terrestrial isopods. *Oecologia* 53: 374-381.

- Hassall, M., Turner, J. G., and Rands, M. R. W. 1987. Effects of terrestrial isopods on the decomposition of woodland leaf litter. *Oecologia* 72: 597-604.
- Ihnen, K., and Zimmer, M. 2008. Selective consumption and digestion of litter microbes by *Porcellio scaber* (Isopoda: Oniscidea). *Pedobiologia* 51: 335-342.
- Kautz, G. and Topp, W. 2000. Acquisition of microbial communities and enhanced availability of soil nutrients by the isopod *Porcellio scaber* (Latr.) (Isopoda: Oniscidea). *Biology and Fertility of Soils* 31: 102-107.
- Knoepp, J. D., Coleman, D. C., Crossley-Jr., D. A., and Clarck, J. S. 2000. Biological indices of soil quality: an ecosystem case study of their use. *Forest Ecology and Management* 138: 357-368.
- Latreille, P. 1804. Histoire naturelle, générale et particulière, des crustacés et des insectes. *Cloportides*, 7: 25-49. Paris.
- Leistikow, A., and Wägele, J. W. 1999. Checklist of the terrestrial isopods of the new world (Crustacea, Isopoda, Oniscidea). *Revista Brasileira de Zoologia* 16: 1-72.
- Loureiro, S., Sampaio, A., Brandão, A., Nogueira, J. A., and Soares, A. M. V. M. 2006. Feeding behaviour of the terrestrial isopod *Porcellionides pruinosus* Brandt, 1833 (Crustacea, Isopoda) in response to changes in food quality and contamination. *Science of the Total Environment* 369: 119-128.
- Rushton, S. P., and Hassall, M. 1983a. Food and feeding ratios of the terrestrial isopod *Armadillidium vulgare* (Latreille). *Oecologia* 57: 415-419.
- Rushton, S. P., and Hassall, M. 1983b. The effects of food quality on the life history parameters of the terrestrial isopod (*Armadillidium vulgare* (Latreille)). *Oecologia* 57: 257-261.

- Schmalfuss, H. 2003. World catalog of terrestrial isopods (Isopoda: Oniscidea). Serie A, Nr. 654. Stuttgarter Beiträge zur Naturkunde. 341p.
- Sousa, J. P., Vingada, J. V., Loureiro, S., Gama, M. M., and Soares, A. M. V. M. 1998. Effects of introduced exotic tree species on growth, consumption and assimilation rates of the soil detritivore *Porcellio dilatatus* (Crustacea: Isopoda). *Applied Soil Ecology* 9: 399-403.
- Szlávecz, K. 1992. The role of terrestrial isopods (Isopoda: Oniscidea) in the decomposition of aquatic macrophyte detritus of Lake Balaton, Hungary. *Opusc Zoologica Budapest* 25: 103-112.
- Szlávecz, K. 1993. Needle litter consumption by two terrestrial isopods, *Protracheoniscus amoenus* (C.L. Koch), and *Cylisticus convexus* (de Geere) (Isopoda, Oniscidae). *Pedobiologia* 37: 57-64.
- Szlávecz, K., and Pobožny, M. 1995. Coprophagy in isopods and diplopods: a case for indirect interaction. *Acta Zoologica Fennica* 196: 124-128.
- Zimmer, M. 2002a. Is decomposition of woodland leaf litter influenced by its species richness? *Soil Biology and Biochemistry* 24: 277-284.
- Zimmer, M. 2002b. Nutrition in terrestrial isopods (Isopoda: Oniscidea): an evolutionary-ecological approach. *Biology Review* 77: 455-493.
- Zimmer, M. & W. Topp. 1998. Microorganisms and cellulose digestion in the gut of *Porcellio scaber* (Isopoda: Oniscidea). *Journal of Chemical Ecology* 24: 1397-1408.
- Zimmer, M. 2006. The role of animal-microbe interactions in isopod ecology and evolution. *Acta Biologica Benrodis* 13: 127-168.

Zimmer, M., and Topp, W. 1999. Relationships between woodlice (Isopoda: Oniscidea) and microbial density and activity in the field. *Biology and Fertility of Soils* 30: 117–123.

Zimmer, M.; J.P. Danko; S.C. Pennings; A.R. Danford; A. Ziegler; R.F. Uglow & T.H. Carefoot. 2001. Hepatopancreatic endosymbionts in coastal isopods (Crustacea: Isopoda), and their contribution to digestion. *Marine Biology* 138: 955-963.

ANEXO 1

Normas do periódico *Journal of Crustacean Biology*.

Disponível em < <http://web.vims.edu/tcs/?svr=www>>. Acesso em: 23 maio de 2009.



[Submit electronically](#) to AllenTrack

Contact Prof. [Frederick R. Schram](#), General Editor, JCB

[Manuscript tracking at JCB](#)

[JCB Abstracts Online](#)

If you are not a native English speaker and wish to submit to JCB, see below under **Submission of Manuscripts** for a recommendation by the editorial staff.

Information for Contributors

Content

The *Journal of Crustacean Biology* contains papers of broad interest dealing with any aspect of crustacean biology, biographies of notable carcinologists, notices of business transacted at meetings of The Crustacean Society, book reviews of works on Crustacea, obituary and memorial pieces, and pertinent announcements. We also can entertain submissions from time to time on other groups of marine arthropods such as pycnogonids and merostomes. Papers will be published in English only, but abstracts or summaries in French, German, Portuguese, or Spanish may be added when appropriate. Descriptions of single new species may be accepted if accompanied by significant information on zoogeography, ecology, phylogenetic relationships, or other biological concerns.

You do not have to be a member of the Society to publish in the Journal of Crustacean Biology, but there are benefits of membership that are specified below.

Publication Costs

(1) The *Journal of Crustacean Biology* accepts papers for publication on the basis of merit. Page charges are optional for society members, but members are requested to pay full page charges, if able. Authors unable to pay full charges are requested to pay as large a share of the page charges as possible. However, lack of funds for page charges will not prevent a paper from being published. Alternatively, successful authors who are not members of the society must pay for all charges in full.

(2) The cost of printing color plates is in addition to the regular page charges and is very expensive. The author must pay for the printing of color. The Crustacean Society cannot cover that additional cost. If an author is unable to pay for color, the figure will be printed in black and white.

(3) The cost of reprints is in addition to the page charges and is billed separately to the author by the printer. The Crustacean Society and the journal are not involved in the production or sale of reprints. If your paper is published, the printer will send you a reprint order form that shows the cost of reprints, and you will pay the printer directly.

(4) If you are a member of The Crustacean Society, the society will send you a free PDF file of your paper, regardless of whether you purchase printed reprints. The PDF file should not be posted on a public website for two years, but it can be distributed like a reprint to colleagues.

Current Policies Concerning Author Costs

1. PAGE CHARGES

Nominal page charges requested from authors are US\$75.00 per printed page, which is less than the true cost to the society to publish a page. Page charges are optional for members of The Crustacean Society, but members are requested to pay full charges if able to do so. Authors unable to pay full charges are requested to pay as large a share of the page charges as possible. If an author is without funds and unable to pay page charges, the Society will, to the best of its ability, cover the cost, but this may result in a delay of the publication date. Alternatively, successful authors who are not members of the society must pay for all page charges in full. However, it is easy to become a member, and a membership form is downloadable from elsewhere on this site.

2. CHARGES FOR AUTHOR'S ALTERATIONS MADE IN PROOF

Authors must pay for each alteration to text made in proof, not to include corrections of printer's errors or editor's errors, and not to include the updating of references listed as "in press" that have since been published or updates to an author's address. The charge is US\$5.00 per alteration. The formula used by our printer to count the number of alterations will be used. This charge is not waived for authors who are members of the society.

3. MANDATORY CHARGES FOR PRINTING COLOR FIGURES

The cost of printing color figures is in addition to all other charges. Authors must pay the full cost of printing color figures after the manuscript has been accepted but before it is published. For recent issues of the journal, the costs of printing color figures has ranged from US\$692.50 to US\$832.50 for **each signature in the final compiled journal**. The cost varies depending on the size and composition of the figures and whether or not they are submitted as hard copy or as electronic image files.

4. BILLING

Authors will be billed for all charges in paragraphs 1-3 on a single invoice, which will be sent immediately after publication of the paper (payments for color plates, see below).

Papers with color illustrations will not be published with the illustrations in color before the full cost is paid by the author. An invoice for printing color figures will be sent to the author when the manuscript is accepted but before it has been sent to the printer. Failure to pay for color illustrations in advance will result in printing the illustrations in black and white.

5. PENALTIES

Authors who fail to pay a promised page charge (full or partial) or the mandatory charges (figures, tables, alterations in proof) will not have other papers published in JCB until the deficit is paid. **JCB will not publish papers from authors who will not pay the nominal page charges but offer to pay for color plates or excess illustrations or tables.** Authors who pay no page charges or pay only partial page charges must be aware that their manuscript may not be published as quickly as those from authors paying in full.

Submission of Manuscripts

Before you submit a manuscript, *and if you are not fully fluent or a native speaker of English*, we recommend the following. American Journal Experts (AJE) provides professional language editing services to

authors around the globe who wish to publish in scientific, technical, medical, and humanities journals. We urge authors who are not well versed in the English language to use this service to improve a paper's English and, therefore, its overall quality. Seeking this assistance is suggested before an article is submitted to JCB for peer review and certainly before it is finally accepted for publication.

AJE has over 500 editors from Harvard, Stanford, MIT, Berkeley, and Duke; these editors are native English speakers and subject-matter experts in a wide variety of fields. They will check your manuscripts not only for terminology and language specific to your field but also for proper English usage, grammar, punctuation, spelling, verb tense, and phrasing. In addition, AJE's professional editors will make sure the text reads naturally and the sentences are well constructed. Visit AJE's website for more information, or to submit a document for their scientific proofreading service use this link: www.JournalExperts.com?rcode=JCB1

When you are secure about your text, manuscripts should be submitted on line to www.jcb.allentrack2.net

While embedded figures can suffice for the review process, by the final revision process, separate text and TIF-figure files will be needed saved at the appropriate resolution.

Form of Manuscripts

The printed manuscript must be typed double-spaced leaving margins of at least 2.54 cm (one inch) all around. Use triple spacing above primary headings. Number pages consecutively in the upper right corner.

Sequence of material should be as follows: Running head, including author name and brief title; Title; Author(s) name(s); Mailing and e-mail address(es) of author(s) (see journal for format); Abstract; Key words; Article body; Acknowledgements; References; Appendix; Tables; Figure captions; Figures (each numbered and identified).

The title page should give the running head; the title of the paper, typed in all capital letters; and the author name(s), followed by affiliation(s) including e-mail address(es). In the case of multiple authorship, list all authors' names first, then each author's initials should be enclosed in parentheses before the appropriate mailing address; the e-mail address should be within parentheses immediately following the mailing address. The running head should contain the first author's surname, a colon, and an abbreviated title in the case of a single author. When there are two authors, include both authors' surnames before the colon. In the case of three or more authors, use only the first author's surname followed by - et al. The running head should not exceed about 52 characters.

The abstract should not exceed one double-spaced page. It should contain a summary of significant findings and note the implication of those findings. The section title "ABSTRACT" should be in all capital letters and centered within the margins.

In the text, the *Journal of Crustacean Biology* follows the style of the most recent of the journal. When in doubt, consult the editor. JCB allows both American and British spelling, but spelling format should be consistent within an article. Metric units of measurement prevail.

Tables and figures should be self-explanatory, not requiring reference to the text. Each table should start on a separate page and must be double-spaced throughout, even if it extends onto multiple pages. Headings and format must be consistent with the style used in *Journal of Crustacean Biology*; see previous issues for format. Vertical rules and excessive horizontal rules should not be used. For treatment of large tables of gene sequences, see below.

Figures must be proportioned to fit nicely within the journal's margins when reduced. Figures will be reduced to either 176 mm width (full page) or 84 mm width (column width) and a maximum height of 237 mm (including the space for caption beneath). For figures with multiple parts (A, B, C, etc.), all of the parts must be together on one figure, not spread over multiple separate photographs. For electronic submission of figures, see below.

All papers referred to in the text should be listed in the "References" section alphabetically by the authors' surnames, then chronologically for multiple papers by the same author(s), e.g., Smith, 1999, followed by

Smith and Brown, 1998, followed by Smith and Jones, 1996, followed by Smith and Jones, 1997. Use only the authors' surnames and initials in the References; place a space between an author's initials. Names of periodicals should be written out in full and should not be italicized. Do not use issue numbers of continuously paginated volumes. Use a hanging indent for multiple lines within one citation (see below for more about hanging indents). All citations in the article must be in the References, INCLUDING the authors of taxonomic names.

A sample citation of an article by a single author in a serial journal follows:

Smith, J. Q. 1981. The distribution of swimming crabs. *Journal of Crustacean Biology* 1: 105-119.

A sample citation of an article by two authors follows:

Martin, G., and P. Juchault. 1999. Androgenic hormone specificity in ten species of lobsters. *Journal of Crustacean Biology* 19: 684-689.

A sample citation of an article in an edited work follows:

Garth, J. S. 1991. Taxonomy, distribution, and ecology of Galapagos Brachyura, pp. 123-125. In, M. J. James (ed.), *Galapagos Marine Invertebrates*. Plenum Publishing Company, New York.

Some Things To Do or Not To Do When Preparing the Manuscript

Do not use a type size smaller than 12 points. We prefer a Times New Roman font.

Do not use boldface type anywhere except to highlight a new taxon.

Do not justify the right margin.

Use italics only for the scientific names of genera and lower categories; do not use underlining.

Do not italicize "et al." and fully spelled Latin words such as "sensu" or "ad libitum", but make no other typesetting indications. Do not underline them.

Do not use running headers on each page of the manuscript; however, do number every page in the upper right corner.

Do not use line numbers in the submitted version. These will be added by the AllenTrack system automatically when it converts your source files into reviewable PDFs.

Center primary and secondary section headings within the margins; primary headings should be in small capital letters, and secondary headings should be in uppercase and lowercase letters. Tertiary (third-level) headings should be even with the left margin, upper and lower case letters, and followed by a period and an em-dash (or three hyphens). Quaternary (fourth-level) headings should be avoided, but if required they should be indented from the left margin, upper and lower case Roman letters, and followed by a period and an em-dash (or three hyphens).

Do not begin a sentence with an abbreviation, especially of a genus-level Latin name of an organism.

Do not use an ampersand (&) for "and", even between authors' names in text citations.

Do not quadruple-space text between paragraphs.

Indent the first line of each paragraph except for the first paragraph after a first- or second-level heading or when a third-level heading is used, which is set flush left on the margin (see above). Do not use the space bar to indent the line; either use the tab key (not preferred) or set a first-line-indent distance in paragraph formatting with the word processor settings (preferred).

In synonymies used in taxonomic papers, use a hanging indent if the references listed after a scientific name extends onto multiple lines (see below for more about hanging indents).

One can use United States Postal Service initials for states and territories of the United States. European Union country abbreviations can be included in postal code formulae, e.g., NL-1092 AD.

Start the text of your paper with an Introduction section.

Place a space between the author's initials in the References section (see above).

Do not use issue numbers of continuously paginated volumes in the References section (see above).

Shorten the title of your paper by eliminating the author and date of a taxon name (they will appear in the text) and by not using "Crustacea: " within parentheses when giving the classification of the organism (this is the *Journal of Crustacean Biology*; abstract services will know to place your article in the subject Crustacea).

N.B.: Include the author and date of publication of each species-level name the first time it appears in the body of the text or in the tables (but not in the title or abstract). In addition, these citations also must appear in the Reference section.

In References, place two spaces between the components of the citation: Author(s). date. Title. Journal with volume and pages. Separate the ranges of the pages with a simple hyphen. (**Do not italicize journal titles.**)

In the References section, do not create a hanging indent by pressing the "enter" or "return" key at the end of each line and the "tab" key at the start of indented lines. Use the software's paragraph formatting feature to make a hanging indent. Otherwise, when the printer translates the file, the line breaks and tabs end up in arbitrary places.

Do not try to create the look of certain characters by creating your own symbol. For example, a superscript letter "o" will not translate as a degrees symbol; it will translate as a superscript "o", which is unacceptable for degrees. Similarly, an apostrophe will not translate as a minute or prime symbol; a prime mark should be used for minutes in latitude-longitude coordinates.

Do not create plus-minus signs by underlining a plus sign. Use the plus-minus symbol instead.

Provide long tables or appendices of character-state data used in phylogenetic analyses as an electronic image file (not text file), because they will be treated as figures in order to avoid retyping them and introducing errors. Provide the table title for such a table on a separate page, because it will be typeset.

Do not submit long tables or appendices of gene sequences. Submit sequence data to an appropriate repository, e.g., GenBank, and publish only the accession numbers.

Do not use unusual fonts. Use the Symbol, Times/Times New Roman, or Palatino fonts. **Serif fonts are preferred over sans-serif fonts for ease of reading.**

Be consistent in the representation of each symbol throughout the document. The correct representation of special symbols is critical if the printer is to use the file.

Distinguish between similar-looking but disparate symbols, such as with the letter "x", a multiplication symbol, and the Greek letter Chi. Use the proper symbol, not the similar-looking letter, i.e., do not use "X" when you mean it to be a multiplication symbol.

Proofs

The printer will send galley proofs to the corresponding author for correction and approval. Changes other than correcting printer's or editor's errors or for updating references listed as "in press" to add the

publication date or to update an author's address will be charged to authors. Printed reprints, available by purchase from the printer only (see Publication costs, above), may be ordered at the time proofs are returned. Separate artwork is discouraged since all images should be submitted as TIF files at the proper resolution of 600 d.p.i. for photographs and 1200 d.p.i. for line art, so separate artwork will be returned only upon request.

Read and return your proofs PROMPTLY to the editor, including also the separate figure proofs. If you will be away from your office around the time proofs are scheduled (around 3 months before the scheduled publication date) make provisions for someone to open and correct the proofs. Delays in returning proofs will cause delays in printing the issue.

Final Submission

Upon final submission, not all parts of the document should be in one electronic file, as they could be upon initial submission. Parts of the document that should be included in a single electronic file are as follows: Running head; Title; Author(s) name(s) and affiliation(s)/ addresses; Key words; Abstract; Article body starting with Introduction; Acknowledgements; References; Non-tabular appendices; and Figure captions.

If the electronic version of your table is to be used, you must create them using the word processor's "Table" command or use a single tab (not space bar) between columns and a hard return between rows. Use no rules except those created by the word processor's Table-menu commands, i.e., do not use underlining to create a rule. Use no borders or shading and no text boxes.

For guidelines on the submission of figures electronically, see below.

Guidelines for Submitting Electronic Art Files to JCB

Separate electronic art files along with the text files must be submitted at the time final revisions are completed.

Line art must be scanned at 1200 dots per inch (472 dots per centimeter). **Halftones must be scanned at 600 dpi** (236 dpc). Combination halftones, i.e., continuous grade plus line art, must be scanned at 600 dpi. Color must be scanned at 350 dpi.

Only Tagged Image File Format (TIFF) [or possibly Encapsulated Postscript (EPS)] formats for both Macintosh and Windows platforms are acceptable. Indicate the file format of the image in the file name, examples: "fig1.tif" or "fig2.eps". [Images in JPEG or GIF format will be 72 dpi and are UNACCEPTABLE for the printing process. Powerpoint, Excel, DeltaGraph, and other similar files are **UNACCEPTABLE** for the printing process.]

The screen and printer font files for any text added to the figure electronically must be included on the disk. Only Adobe Postscript fonts for either Macintosh or PC platforms are acceptable. Do NOT use True-Type or system "bitmap" fonts. In EPS files, type can be converted to outlines to avoid the necessity of submitting font files.

Do not send multiple versions of the same figure.

[Submit electronically](#)

If you have any questions regarding these instructions, contact the JCB Editor at: jcb@whidbey.com.

Last updated: 16 April 2009.