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INSTITUTO DE BIOCÊNCIAS
PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA



Tese de Doutorado

Ecologia e Conservação do Sapinho-admirável-de-barriga-vermelha

***Melanophryniscus admirabilis* (Anura: Bufonidae)**

Michelle Abadie de Vasconcellos

Porto Alegre, abril de 2021

ECOLOGIA E CONSERVAÇÃO DO
SAPINHO-ADMIRÁVEL-DE-BARRIGA-VERMELHA
Melanophryniscus admirabilis (ANURA:BUFONIDAE)

Michelle Abadie de Vasconcellos

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*“The secret, Alice, is to surround yourself with people who make your
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Resumo

A maior parte dos anfíbios ameaçados no mundo têm distribuição inferior a 20.000 km². O que aconteceu na história recente de uma espécie para que ela seja encontrada em somente uma faixa restrita de distribuição? É uma espécie rara? É pouco abundante? Ou somente é pouco estudada? Esta tese de doutorado tem como objetivo contribuir com o conhecimento sobre história natural e abundância de um anuro microendêmico, ao mesmo tempo que reavalia seu risco de extinção e sugere ações de conservação. Além da introdução geral (em português), a tese é composta por três capítulos, cada um referente a um artigo científico (em inglês). No primeiro capítulo, abordamos “tudo que você gostaria de saber sobre *Melanophryniscus admirabilis*”, o sapinho-admirável-de-barriga-vermelha. Descrevemos o girino, o canto de anúncio, o padrão temporal reprodutivo, as estratégias de reprodução e de defesa, tempo de eclosão do girino e de metamorfose, além do dimorfismo sexual, fidelidade de sítio e longevidade. No segundo capítulo, estimamos o tamanho da única população conhecida da espécie, e sua variação ao longo de oito anos, utilizando modelos de marcação e recaptura aliados à fotoidentificação. No último capítulo, descrevemos a distribuição microendêmica da espécie, além de identificar as ameaças e ordená-las por relevância. Utilizando essas informações e o tamanho populacional, nós reavaliamos o status de conservação da espécie sob os critérios B, C e D da Lista Vermelha da IUCN. Adicionalmente, discutimos ações prioritárias para a conservação do sapinho-admirável e como elas podem reduzir o risco de extinção da espécie. Demonstramos que, embora um grande esforço de ações protetivas precise ser feito, a distribuição de *M. admirabilis* é tão pequena que somente o encontro de uma nova população poderia alterar a sua condição de espécie ameaçada.

Palavras-chave: microendemismo, espécie ameaçada, *Melanophryniscus admirabilis*, história natural, tamanho populacional, distribuição geográfica

Abstract

Most threatened amphibians have a distribution smaller than 20,000 km². What happened in recent history for it to have such a restricted range? Is it a rare species? Is it not abundant? Or is it not just studied enough? This doctoral thesis aims to contribute to the body of knowledge about the natural history and abundance of a microendemic anuran while reassessing its risk of extinction and suggesting conservation actions. In addition to the general introduction (in Portuguese), the thesis consists of three chapters, each one referring to a scientific article (in English). In the first chapter, we address "everything you would like to know about *Melanophryniscus admirabilis*", the Admirable Redbelly Toad. We described the tadpole, the advertisement call, the temporal reproductive pattern, the reproductive and defensive strategies, the tadpole's hatching time and metamorphosis, in addition to sexual dimorphism, site fidelity and longevity. In the second chapter, we estimated the abundance for the only known population of the species, and its variation over eight years, using capture-recapture models with photoidentification. In the last chapter, we described the microendemic distribution of the species, besides identifying threats and ordering them by relevance. Using this information and population size, we reassessed the species' conservation status under IUCN Red List criteria B, C and D. Additionally, we discussed priority actions for the conservation of the toad and how they can reduce the extinction risk of the species. We demonstrated that, although a great effort is needed to protect this species, the distribution of *M. admirabilis* is so small that only finding a new population could remove the species from threatened categories.

Keywords: microendemism, threatened species, *Melanophryniscus admirabilis*, natural history, population size, geographic distribution

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Introdução Geral

“Raridade” é um termo amplamente utilizado na literatura científica, porém nem sempre é empregado no mesmo contexto (GASTON, 1994). Na maioria das vezes, esse termo está associado à baixa abundância ou à pequena área de distribuição de uma espécie e, em geral, abundância e distribuição não são independentes (GASTON, 1994). A relação positiva entre essas duas variáveis é um dos padrões empíricos mais gerais dentro da ecologia: espécies de distribuição restrita tendem a ser pouco abundantes (GASTON; BLACKBURN, 2007). Entretanto, uma espécie abundante na sua pequena distribuição pode ser considerada rara?

Algumas espécies tornam-se raras imediatamente antes de sua extinção, enquanto outras espécies mantêm essa raridade ao longo de toda sua existência. Embora o termo raridade ainda seja amplamente utilizado, Rabinowitz (1981) propôs um conceito mais preciso a partir de três componentes, onde uma espécie extremamente rara tem (1) pequena distribuição geográfica, (2) alta especificidade de habitat e (3) baixa abundância local. No sentido de endemismos restritos, espécies com alcance geográfico estreito e especificidade de habitat estreita são raridades clássicas e estão frequentemente ameaçadas (RABINOWITZ, 1981).

As características demográficas de espécies altamente endêmicas podem indicar um maior risco de extinção e devem ser consideradas nas iniciativas de conservação, uma vez que essas espécies são potencialmente mais sensíveis à estocasticidade ambiental e demográfica. Compreender a diferença entre uma população estável e uma população suscetível à extinção requer conhecimento sobre o tamanho da população e sua variação temporal (INCHAUSTI; HALLEY, 2003). As mudanças no número de indivíduos maduros de uma espécie, assim como a amplitude da distribuição geográfica, são

critérios-chave para a avaliação do risco de extinção pela União Internacional para Conservação da Natureza (IUCN STANDARDS AND PETITIONS COMMITTEE, 2019), cuja abordagem tende a estar fortemente relacionada à estrutura proposta por Rabinowitz sobre raridade (TOLEDO *et al.*, 2014).

Esse conhecimento sobre os aspectos demográficos das espécies é especialmente importante para os anfíbios, que são particularmente propensos à extinção (BISHOP *et al.*, 2012; STUART *et al.*, 2004). Embora cerca de 40% das espécies de anfíbios estejam provavelmente ameaçadas ('best estimate'; IUCN, 2021), parece ser o grupo de vertebrados menos conhecido: somente 12% das espécies têm informações disponíveis sobre aspectos de história de vida ou dados demográficos (CONDE *et al.*, 2019). Esse nível de informação cai para apenas 0,2% quando somente as ameaçadas são levadas em conta (CONDE *et al.*, 2019).

Na falta de informações sobre a abundância e sua tendência ao longo do tempo, a maior parte das classificações de anfíbios em alguma categoria de ameaça é baseada exclusivamente em critérios de distribuição geográfica (Critério B e D2; IUCN, 2021). Isso significa que, em escala global, cerca de 85% dos anfíbios ameaçados não dispõem de informações para serem avaliados por outros critérios (IUCN, 2021). Além disso, no mínimo 90% dos anfíbios ameaçados ou quase ameaçados ('Near Threatened' – NT) podem ser considerados endêmicos (*i.e.*, tem distribuição inferior a 20.000 km²; IUCN 2021). No Brasil, das 41 espécies de anfíbios listadas como ameaçadas, trinta e oito (93%) tiveram seu status de conservação definido com base nos critérios referentes à extensão de ocorrência (critério B1) ou à área de ocupação (critérios B2 ou D2; Portaria MMA N^o 444, 2014).

Nos Neotrópicos, a América do Sul se destaca nas taxas de endemismo de anuros (PIMM *et al.*, 2014; VILLALOBOS *et al.*, 2013). Entre esses destaca-se o gênero *Melanophryniscus* (Anura: Bufonidae), que possui uma notável riqueza de espécies endêmicas, além de ser um importante grupo para a conservação (ZANK *et al.*, 2014). Os sapinhos-de-barriga-vermelha, como são chamadas as espécies desse gênero, são representados atualmente por 29 espécies e distribuem-se exclusivamente nas regiões tropical e subtropical da América do Sul (FROST, 2021; ZANK *et al.*, 2014). Muitas das espécies desses pequenos anuros (comprimento rostro-cloacal inferior a 50 mm; GARRAFFO *et al.*, 2012) estão ameaçadas de extinção e apresentam uma distribuição muito restrita (ZANK *et al.*, 2014). Assim como para a maioria dos anfíbios anuros, os aspectos ecológicos e de história de vida dos sapinhos-de-barriga-vermelha ainda estão longe de serem conhecidos. As nove espécies de *Melanophryniscus* que se encontram listadas em alguma categoria de ameaça foram avaliadas com base apenas na sua restrita distribuição (CARREIRA; MANEYRO, 2015; IUCN, 2021; PORTARIA MMA N° 444, 2014; VAIRA *et al.*, 2012). Entre as espécies do gênero que tive oportunidade de conhecer, uma delas me chamou a atenção. Considerando as chamadas '7 formas de raridade' (RABINOWITZ, 1981), *Melanophryniscus admirabilis*, o sapinho-admirável-de-barriga-vermelha (“Admirable Redbelly Toad”, em inglês), se enquadra em dois dos três eixos de raridade: pequena amplitude de distribuição e alta especificidade de habitat. Entre os endemismos, ele é um dos mais extremos. Possivelmente, a vulnerabilidade de uma espécie tão restrita, tão particular e tão iminentemente ameaçada de desaparecer foram as motivações para meu envolvimento pessoal e acadêmico nessa história de conservação.

Melanophryniscus admirabilis foi descrito em 2006, para apenas uma localidade no extremo sul da Mata Atlântica brasileira (DI-BERNARDO; MANEYRO; GRILLO,

2006), cerca de três anos após a publicação das listas de espécies da fauna ameaçada no Brasil (INSTRUÇÃO NORMATIVA MMA Nº 3, 2003) e no Rio Grande do Sul (MARQUES *et al.*, 2002). Portanto, naquele momento, a espécie não estava legalmente protegida por qualquer política ou lei ambiental brasileira. Em 2010, quatro anos após a descrição da espécie, a Fundação Estadual do Meio Ambiente – FEPAM, instituição do estado do Rio Grande do Sul responsável pelo licenciamento ambiental do estado, concedeu a primeira de três licenças para a construção da Pequena Central Hidrelétrica (PCH) Perau de Janeiro. O projeto era localizado no rio Forqueta, a apenas 500 metros à montante do único local de ocorrência da espécie. Nosso trabalho na região foi inicialmente motivado pelo senso de urgência de conservação do sapinho-admirável-de-barriga-vermelha, já que nada se sabia a respeito da espécie que, naquele momento, corria o risco de ser extinta.

Graças a um esforço multi-institucional, a licença para a construção da PCH foi cancelada (FONTE *et al.*, 2014), e a situação serviu de gatilho para iniciarmos um projeto de monitoramento da espécie a longo prazo. Nesta tese de doutorado, apresento uma compilação (de parte) do trabalho que venho desenvolvendo com *M. admirabilis* ao longo dos últimos 10 anos, praticamente durante toda minha vida acadêmica.

Esta tese está estruturada em três capítulos, cada um correspondendo a um artigo. No primeiro capítulo, abordamos aspectos da história natural do sapinho-admirável-de-barriga-vermelha, passando pela descrição do girino, descrição do canto de anúncio, dimorfismo sexual, comportamento reprodutivo e longevidade. A abordagem utilizada vai desde observações oportunísticas de campo até marcação e recaptura aliada a técnica de fotoidentificação.

No segundo capítulo, estimamos o tamanho da única população da espécie e sua variação ao longo do tempo, abordando o uso de três estimadores: Modelo de População Fechada (OTIS *et al.*, 1978), Modelo de População Aberta – POPAN (SCHWARZ; ARNASON, 1996) e Desenho Robusto (POLLOCK, 1982). Para isso, construímos um banco de dados de marcação e recaptura ao longo de oito anos.

De maneira geral, quando se trata de espécies com distribuição restrita, espera-se que elas apresentem um risco de extinção mais elevado, pois são mais suscetíveis a efeitos estocásticos demográficos ou ambientais. Por outro lado, será possível que uma espécie microendêmica, como o sapinho-admirável-de-barriga-vermelha, não apresente um alto risco de extinção? No último capítulo, apresentamos a distribuição atual da espécie, fazemos a reavaliação do estado de conservação, ranqueamos ameaças e discutimos as ações prioritárias para tentar reduzir o grau de ameaça sobre a espécie.

Cada capítulo foi formatado conforme a norma da revista para a qual será submetido (informado no início de cada capítulo). Para facilitar a leitura, as tabelas e figuras foram inseridas no corpo de texto. Na última seção, após o terceiro capítulo, fiz uma compilação das principais conclusões de cada um dos artigos e como que eu espero que essa tese contribua para redução do risco de extinção do sapinho-admirável-de-barriga-vermelha e inspire outras(os) pesquisadoras(os) a trilhar o caminho da conservação.

Capítulo 1

Morphology, reproduction, longevity, and bioacoustics of the microendemic and threatened Admirable Redbelly Toad

Este manuscrito segue o formato da revista *Zoology*. A fim de facilitar a leitura, as figuras e tabelas foram inseridas no corpo do texto.

Morphology, reproduction, longevity, and bioacoustics of the microendemic and threatened Admirable Redbelly Toad

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1 **Abstract**

2 Although information about the natural history of a species is fundamental for
3 conservation planning, little is known about threatened species, and this is more critical
4 for amphibians. One of the most relevant aspects of amphibian natural history is its
5 reproductive behaviour, particularly for explosive breeders that are often considered to
6 be rare. *Melanophryniscus admirabilis* is a microendemic and Critically Endangered
7 toad for which the lack of such information almost resulted in its extinction. In this
8 study, we address aspects of morphology, reproduction, bioacoustics, and longevity of
9 *M. admirabilis* and provide insights on the behaviour of *Melanophryniscus* species.
10 From seven years of capture-recapture data and *in situ* observations, we found that the
11 species is a seasonal explosive breeder and we recaptured individuals seven years from
12 the first capture. The species has female-biased sexual size dimorphism, diurnal and
13 nocturnal breeding activity and site fidelity. We also describe the tadpole and the
14 advertisement call. Similar to other *Melanophryniscus* species, *M. admirabilis*
15 characteristics seem to be adapted to ephemeral and particularly risky and noisy
16 environments. Based on this study's findings, it is possible to improve the identification
17 of suitable habitats for the species, as well as facilitating in-situ protection.

18 **Keywords**

19 *Melanophryniscus admirabilis*, lifespan, capture-recapture, tadpole, advertisement call

20 **1. Introduction**

21 Knowing the natural history of a species is a fundamental step to effectively plan
22 conservation actions. When there is limited knowledge about a species, it often becomes
23 difficult to assess its conservation status. This can be of particular concern for rare or
24 poorly known species, as the lack of information can lead to a Data Deficient (DD)
25 categorization, when, in fact, it can be actually threatened (IUCN, 2021). The
26 availability of information about a species, and its consequent categorization or not in a
27 red list, can determine how it will be deemed by decision-makers.

28 Besides being the least known tetrapod group, with only 12% having any
29 demographic information (Conde et al., 2019), amphibians are the most threatened
30 vertebrate class globally (IUCN, 2021). When only threatened amphibians are taken
31 into account, the level of demographic information drops to 0.2% (Conde et al., 2019).
32 Out of the 7,212 species assessed by the IUCN (2021), 34% are currently listed in a
33 threatened category (Critically Endangered (CR), Endangered (EN) or Vulnerable
34 (VU)). While a considerable proportion of amphibian species is at risk of extinction,
35 amphibians are also the group of tetrapods with the highest proportion of Data Deficient
36 (DD) species, with 16% listed as such (IUCN, 2021). Estimates indicate that up to 50%
37 of all amphibian species could be threatened (González-del-Pliego et al., 2019; IUCN,
38 2021). Alarmingly, other studies have indicated even higher estimates (Howard and
39 Bickford, 2014).

40 Anurans are the most diverse group of amphibians, currently comprising more
41 than 7,300 species (Frost, 2021). They present the greatest diversity of reproductive
42 modes (Haddad & Prado, 2005) and their reproductive behaviour is one of the most
43 relevant aspects of their life history. This is especially true for species with explosive
44 reproduction. Explosive breeders (*sensu* Wells, 1977) usually spend much of their lives

45 in hiding, only emerging during short periods of time to reproduce. For this reason, they
46 are often considered to be rare. The breeding period can last from 2 days to a few weeks
47 (Wells, 1977) and understanding the timing of these events may optimize research and
48 management planning.

49 The South American Redbelly toads, genus *Melanophryniscus* (Anura:
50 Bufonidae), exemplify the typical knowledge shortfalls for Neotropical anurans. They
51 are small toads, explosive breeders with poor knowledge about their natural history and
52 ecology, despite great interest in their conservation (Zank et al., 2014). Out of the 29
53 valid species, more than half (16 spp.) have been described in the last 20 years (Frost,
54 2021), suggesting that the genus' species richness is still underestimated. Fourteen
55 species are endemic to the Atlantic Forest biome (Frost, 2021), a hotspot for
56 conservation (Myers et al., 2000). Out of these, 4 are considered to be threatened (CR,
57 EN, VU), 2 are listed as DD and 1 as Near Threatened (NT) in the Brazilian Red List
58 (Portaria MMA N° 444, 2014), with natural history and reproductive biology
59 information available for only a few species (Baldo et al., 2014; Caorsi et al., 2014).

60 The microendemic and Critically Endangered Admirable Redbelly Toad
61 (*Melanophryniscus admirabilis* Di-Bernardo, Maneyro, and Grillo, 2006) is an
62 emblematic example. The lack of scientific knowledge about this species almost
63 resulted in its extinction. It was described from a single site in the southern edge of the
64 Brazilian Atlantic Forest, in 2006, four years after the last national and regional red list
65 assessments, which were valid at that time (Instrução Normativa MMA n° 3, 2003;
66 Marques et al., 2002). For this reason, its extinction risk was not assessed, and therefore
67 it remained unprotected by any conservation policy or national environmental law. In
68 2010, the regional government granted a licence for the construction of a small
69 hydroelectric power plant exactly in the type locality. Even though at the time, the only

70 available information about the species was its original description, a multi-institutional
71 effort led by the authors of this study allowed for the quick assessment of its
72 conservation status (Fonte et al., 2014). Due to its extremely reduced area of occupancy
73 (AOO = 0.035 km²) and the imminent threat imposed by the construction of the power
74 plant, the species was further listed as CR at the global (IUCN SSC Amphibian
75 Specialist Group, 2013), national (Portaria MMA N° 444, 2014), and regional (Rio
76 Grande do Sul, 2014) levels, remaining in this category at all levels since then (Haddad
77 et al., 2018).

78 The licence for the construction of the hydroelectric power plant was cancelled
79 (Fonte et al., 2014), and the situation triggered the beginning of a long-term monitoring
80 project focused on the species. The main objective of this paper is to present the
81 knowledge produced over seven years of population monitoring, from capture-recapture
82 data and *in situ* observations. Here, we provide information about the population
83 (longevity and sexual dimorphism), reproduction behaviour (site fidelity, reproductive
84 strategies, temporal breeding pattern, oviposition and time to metamorphose),
85 description of the tadpole and the advertisement call, and aspects of the defensive
86 behaviour of *M. admirabilis*. Our results contribute to Neotropical anurans knowledge
87 and provide insights into the behaviour of *Melanophryniscus* species. Moreover, we
88 expect that the scientific knowledge presented here will contribute to implementing
89 efficient conservation actions to protect the only known population of the Admirable
90 Redbelly Toad.

91 2. Materials and methods

92 2.1 Study area

93 We performed the study at Perau de Janeiro (28° 51' 25.3" S; 52° 18' 12.3" W),
94 the Admirable Redbelly Toad type and only locality, at the southernmost range of the

95 Atlantic Forest biome. Located on the slopes of the southern border of the Brazilian
96 Southern Plateau, in the Brazilian state of Rio Grande do Sul, Perau de Janeiro is part of
97 a ca.5 km² forest patch, one of the largest forest remnants in the region.

98 The climate of the region is classified as Subtropical Humid, without dry season
99 and with hot summers (*Cfa*, Koeppen's climate classification; Alvares et al, 2013).
100 Precipitation is evenly distributed throughout the year (1600 mm; Minella et al., 2009)
101 and the temperature determines the seasonality of the region, with four well defined
102 seasons (Supplementary Information Figure S1; Zepner et al., 2020). The *Cfa* climate
103 type is characterised by average temperatures varying between -3 °C and 18 °C for the
104 coldest month (July) and above 22 °C for the hottest month (January). Negative
105 temperatures are common during the winter, with an average of 19 frost days per year
106 (Moreno, 1961).

107 Our study area is part of the Taquari-Antas River basin, at an elevation of
108 around 550 m above sea level. It extends along 700 m of steep forested slopes of the
109 Forqueta River. We sampled along ca. 400 m on the left margin, where most individuals
110 concentrate to breed on pools formed on volcanic rock outcrops along the river bank.
111 These flattened river bank outcrops are 1-14 m wide, from the river margin to the forest
112 edge. Even though we concentrated our efforts on this site, we also made a few registers
113 on the right margin of the river.

114 *2.2 Data collection*

115 Our final sampling effort totalled 122 days collecting capture-recapture data,
116 starting in October 2010 and finishing in December 2017, at different intervals. Over
117 the years, as we learned about the species, we adjusted our sampling design over this
118 period. Fieldwork was carried out between October 2010 and October 2011 (monthly),
119 July 2013 and August 2013 (monthly), and between August 2013 and October 2014

120 (bimonthly). In 2016 and 2017, a local resident and project partner visited the site once
121 a week and sent us information about the Admirable Toad activity. From this
122 information, we conducted surveys from July to December, whenever we detected
123 breeding activity (see below for details). Fieldwork varied from two (2010-2011) to
124 eight (2013-2014) continuous days in each survey. In 2012 and 2015 we did not survey
125 the population.

126 We arbitrarily divided the flattened river bank outcrop into 27 sectors (15 metres
127 in length). We used Visual Encounter Survey (Crump and Scott Jr., 1994) and Surveys
128 at Breeding Sites (Scott Jr. and Woodward, 1994) to search for individuals during the
129 day, starting about 9 am and stopping when new individuals were no longer detected,
130 sometimes at night (i.e. surveys were not limited by time). Captured toads were sexed,
131 measured (snout-vent-length – SVL; calliper to the nearest 0.01 mm), photographed and
132 then released at the same place of capture. We classified individuals into three
133 categories: juvenile, male (adult) or female (adult). We considered individuals as adults
134 when sex recognition was possible, by the presence of brownish nuptial pads at the base
135 of the male thumb (absent in females), by the calling behaviour of males, or by the
136 presence of eggs in females. We considered as juvenile all individuals that did not have
137 any secondary sex character (which in this study were those smaller than 28 mm).

138 We marked individuals using the Photographic Identification Method (PIM)
139 following a semi-automatic procedure with the software Wild-ID (Bolger et al., 2012).
140 We photographed the entire ventral surface of black pigmentation on each animal and
141 used its unique colour pattern resulting from the yellowish-green glands for individual
142 recognition (Caorsi et al., *in prep*). We took two to five pictures of each captured
143 individual and selected the best one, which was cropped to the area of interest, between
144 the throat and the cloaca, eliminating as much of the background as possible. The

145 software Wild-ID returned the 20 closest matches in our photo catalogue. This list was
146 subsequently examined by a researcher who attributed a new or existing individual
147 identity to the captured toad. This approach was successfully tested and applied to this
148 species (Abadie et al., 2021 - Chapter 2; Caorsi et al., *in prep*) and other South
149 American Redbelly Toads (*e.g.* Bardier et al., 2019; Caorsi et al., 2012; Elgue et al.,
150 2014). We used the capture-recapture method to address the longevity, site fidelity and
151 dispersal of individuals.

152 To infer reproductive temporal pattern and reproductive strategies, we recorded
153 the following evidence of reproductive activity: the presence of calling males,
154 amplexant pairs and recently laid clutches (indicating current breeding activity), or
155 presence of old clutches and tadpoles (indicating previous breeding activity). We
156 considered as explosive reproductive events the aggregation of adults at the breeding
157 site coupled with associated reproductive behaviour (Wells, 1977).

158 We recorded the number of eggs per clutch and time of hatching by monitoring
159 *in situ* 17 recently laid clutches until hatching, in different pools at the breeding site in
160 2011 (n= 12) and 2013 (n=5). Furthermore, in October 2014, we collected clutches to
161 conduct *ex situ* experiments for another study on the exposure to pesticides and the
162 antioxidant capacity of the species (da Silva et al., 2021), allowing us to observe the
163 development of tadpoles (control group) until metamorphosis. To perform this
164 experiment, we collected clutches in different pools along the breeding site and, soon
165 after hatching, tadpoles were randomly distributed in glass aquaria. The tadpoles were
166 kept in the laboratory, with photoperiod set to 12 h light: 12 h dark, room temperature
167 set to 23 ± 1 °C and fed once daily with commercial fish feed. For more details, see da
168 Silva et al. (2021). The newly metamorphosed juveniles were fixed in 10% formalin.

169 Vocalisations were recorded in October 2014 using a Marantz PMD 670 and a
170 Sennheiser ME 67 directional microphone at a distance of about 50-100 cm from the
171 calling individual. To obtain the best signal-to-noise ratio and avoid distortion, the
172 recording level was adjusted manually. Recording level, microphone distance and
173 orientation were kept constant during each session. Sounds were recorded using a
174 sampling rate of 48 kHz, 16-bit depth and uncompressed WAV as output format. After
175 each recording, we measured the air and water temperature of the specific individual
176 calling site using a digital thermometer (accuracy 0.1°C). One recorded individual was
177 collected (voucher) and fixed in 10% formalin.

178 To describe the tadpole, we randomly collected them in the field from different
179 pools, in October 2011. Following the standard methodology to describe tadpoles, we
180 used tadpoles between Gosner stages 32 - 37 (n= 20; Gosner, 1960). Collected tadpoles
181 were fixed in 10% formalin.

182 Notes of reproductive and defensive behaviours were described based on
183 opportunistic observations. All field procedures followed recommendations by the
184 Chico Mendes Institute for Biodiversity Conservation (ICMBio) under licences number
185 40004-5 and 10341-1 issued by the Biodiversity Information and Authorization System
186 (SISBIO), and Institutional Research and Ethics Committees on Animal Use at UFRGS
187 (Projects 19541, 25528, 25526) to Márcio Borges-Martins. All collected individuals
188 were deposited in the Coleção Herpetológica do Departamento de Zoologia da
189 Universidade Federal do Rio Grande do Sul (UFRGS), in Brazil, under the numbers
190 UFRGS XXXX (newly metamorphosed juveniles), UFRGS 7070 (vocalizing male
191 voucher) and UFRGS 6358 (tadpoles).

192 2.3 Data analyses

193 To test for sexual size dimorphism, we used only the data collected in 2013 and
194 2014 since all individuals over that period were measured by the same researcher (TM),
195 ensuring standardisation. We applied a Simple Linear Model to test whether the size is
196 associated with the individuals' sex.

197 To infer lifespan (longevity), we analysed the time between captures, using the
198 entire capture-recapture database (2010 – 2017). To infer site fidelity, we used the
199 maximum distance (in sectors) between two captures of the same individual. We
200 applied a binomial Generalised Linear Model to determine if movements were
201 influenced by sex (male or female) or maturity (adult or juvenile). Analyses were
202 performed using R version 4.0.2 and results were considered significant when $p < 0.05$.

203 To describe the annual reproductive activity pattern, we used the number of
204 captured individuals (including the zeros) in a sampling day from two data subsets:
205 2010-2011 and 2013-2014. We assumed that all adult toads at the breeding site were
206 reproductively active adults. First, we converted sampling dates into Julian days and
207 then treated them as angles (0° to 360°). To identify if the distributions of sampling and
208 frequency of captures were significantly different from a uniform distribution ($p <$
209 0.05), we performed Rayleigh's test using the "circular" R package (Agostinelli and
210 Lund, 2017). All statistical analyses were performed using R version 4.0.2.

211 The advertisement calls were analysed using Raven Pro v. 1.4 software for Mac
212 OS X (Bioacoustics Research Program 2011). We obtained temporal properties from
213 oscillograms (temporal resolution = 5.33 ms) and spectral information using Fast
214 Fourier Transformation (512 points Hann window; frequency resolution = 188 Hz). For
215 call variables, the nomenclature follows Kohler et al. (2017) and Emmrich et al. (2020),,
216 and for vocal repertoires and behaviours, Wells (2007). Since the advertisement call of
217 the *Melanophryniscus* species is composed of two distinct segments (call part A and call

218 part B), we analysed the following parameters: call duration (s), call part A duration (s),
219 number of pulses per call part A, pulse duration of call part A (s), pulse interval of call
220 part A (s), peak frequency of call part A (Hz), interval between call part A and call part
221 B (s), call part B duration, number of pulses per call part B, pulse rate in call part B
222 (pulses/sec), pulse duration of call part B (s), pulse interval of call part B (s), and peak
223 frequency of call part B (Hz).

224 For the description of the tadpole, we followed the anatomical nomenclature
225 suggested by Altig and McDiarmid (1999). We took eight measurements (mm) with the
226 aid of a stereoscope: total length (TL), body length (BL), tail length (TAL), tail
227 musculature height (TMH), maximum tail height (MTH), internarial distance (IND,
228 measured between the internal edges of narial apertures), interorbital distance (IOD, the
229 distance between interior edges of eyes), and tail musculature width (TMW). To allow
230 comparisons with other descriptions of *Melanophryniscus* spp., we took additional
231 measurements suggested by Lavilla & Scrocchi (1986): body maximum width (BMW),
232 body width at nostrils level (BWN), body width at eye level (BWE), body maximum
233 height (BMH), rostrum-spiracular distance (RSD, measured horizontally from tip of snout
234 to posterior edge of the tube), frontonasal distance (FN, from the tip of snout to anterior
235 edge of nostrils), eye diameter (E), oral disc width (OD, disc measured folded), dorsal
236 gap length (DG) and ventral gap length (VG).

237 **3. Results**

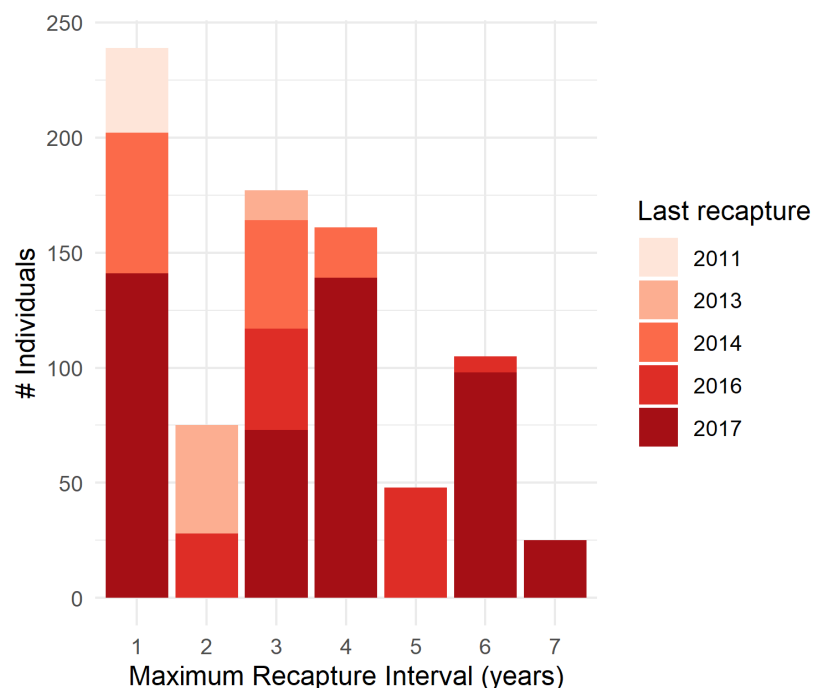
238 *3.1 Populational Data*

239 Between 2010 and 2017, we obtained 4919 captures of 2069 individuals. Of
240 these, males were more frequent (1298) than females (631) and juveniles (160). Twenty
241 individuals were first captured as juveniles and recaptured as adults in the following
242 years. The recapture rate was high, with 40% of the individuals (830 individuals)

243 captured at least twice, while 332 (16%) were recaptured three times or more in
244 different year intervals.

245 Females measured between 31.45 and 41.34 mm (36.26 ± 2.06 mm, $n = 302$),
246 while males measured between 29.00 and 39.19 mm (33.18 ± 1.49 mm, $n = 760$).
247 Although there is considerable overlap between measurements of both sexes, we
248 observed sexual dimorphism in size (F -value = 730.54, $p < 0.01$, $df = 1060$;
249 Supplementary Figure S2).

250 Considering the maximum interval between the first and last captures of
251 individuals, the maximum recorded lifespan was 7 years: 25 from 169 adult individuals
252 which however could live longer, since all of them were recaptured in 2017, the last
253 year of this monitoring (Figure 1). We do not have surveys from 2012 or 2015 and,
254 consequently, certain combinations between first and last captures were not possible
255 (e.g. 2010-2012, 2012-2014, 2012-2017, 2010-2015, 2013-2015, 2015-2017).



256 **Figure 1.** The maximum time interval between first and last captures of
257 *Melanophryniscus admirabilis* over the study duration (2010 – 2017).
258

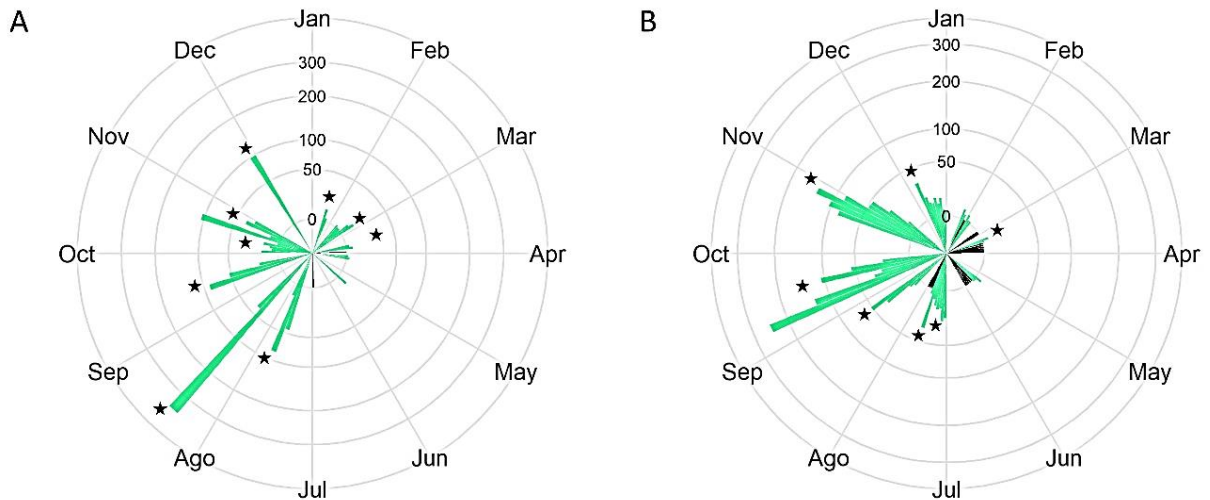
259 *3.2 Reproductive activity*

260 When toads were not breeding, they were hardly found on the riverbank, but
261 some could be found in small hollows, leaf litter or other shelters in the adjacent forest
262 (Supplementary Figure S3), but not far from the breeding site in the rocky riverbank.
263 The maximum distance from the river we found toads, into the forest, was about 50
264 metres from the river bank (a walking male).

265 Site fidelity was high, with most recaptures occurring in the same (38%, n =
266 394) or the adjacent (28%, n = 295) riverbank sector from the first capture. However, a
267 few recaptures (n = 2) correspond to individual movements of up to 300 m
268 (Supplementary Figure S4). Site fidelity was not associated with sex, but with sexual
269 maturity. Juveniles had a higher probability (62%; CI = 0.44 – 0.78) of changing sectors
270 than adults (33%; CI = 0.30 – 0.36; $p < 0.01$).

271 We recorded amplexant pairs not necessarily associated with explosive breeding
272 events. We observed some evidence of reproductive activity (current or previous) in
273 almost all months of the year, except for April, May and June (Figure 2). However, the
274 number of captures at the breeding site varied widely among the months (2010-2011: r
275 =0.7144, $p < 0.01$; 2013-2014: $r = 0.7757$, $p < 0.01$), despite the uniform distribution of
276 sampling effort over both periods considered in this analysis (2010-2011: $r = 0.1423$,
277 $p = 0.5266$; 2013-2014: $r = 0.0591$, $p = 0.8023$; Figure 2). The explosive breeding events
278 were concentrated between July and December.

279



280

281 **Figure 2.** Seasonal pattern of reproductive activity of the Admirable Redbelly Toad
 282 from 2010 to 2011 (A) and from 2013 to 2014 (B), based on the number of adults
 283 captured at the breeding site. Field trips were conducted monthly in 2010-2011 and
 284 bimonthly in 2013-2014. Each bar represents the number of captures on a sampling day.
 285 The black bar indicates no capture on a sampling day. The black star represents any
 286 evidence of breeding activity, current or previous, based on calling males, amplexant
 287 pairs, clutches, and tadpoles.

288

289 In explosive breeding events, toads could be easily found, because hundreds of
 290 individuals aggregated on the rocky outcrop along the riverbank, using small pools as
 291 vocalisation and oviposition sites (Figure 3). In these events, it was more common to
 292 hear release calls than advertisement calls. In October 2013, individuals bred at least
 293 four consecutive days, during an explosive breeding event, and in September 2014 we
 294 recorded two events with two days between them (3 and 2 days of breeding activity,
 295 respectively). The maximum number of individuals recorded on the same day was 361
 296 (August 2011). On rare occasions, some toads mated in small pools inside the forest,
 297 but always within a few meters of the river margin.

298 We observed diurnal and nocturnal reproductive activity, during explosive
 299 breeding events. Most males vocalised inside the water at or near small and shallow

300 temporary pools (<1 m² of surface and <20 cm depth, Bordignon, 2019; Figure 3A-B).
301 On occasion some males vocalised hidden under rocks or in rock crevices or underneath
302 herbaceous vegetation (at the edge between the forest and the rocky outcrop), mainly
303 during the day. The temporary pools are filled directly by rainwater and are maintained
304 by the water that drips from the riparian forest growing on the steep slopes. After heavy
305 rains, the river overflows and “washes” the rocky margins (Supplementary Figure S5),
306 sometimes carrying egg clutches and tadpoles to the river. During the austral winter
307 (June to August), the pools are filled with fallen leaves from the surrounding deciduous
308 forest. In summer (December to February), at the end of the main reproductive period,
309 we noted that the availability of water in pools is reduced, and there is an accumulation
310 of fallen fruits from the abundant Myrtaceae trees. In this period, we recorded little
311 reproductive activity restricted to pools free of rotting fruit.

312 In explosive breeding events, we recorded intense male-male combat and
313 struggles including amplexant pairs and up to three additional dislocating males (Figure
314 3D). Besides, we noticed interspecific amplexus between a male and a juvenile of
315 *Rhinella icterica* (Figure 3G), and males mating with dead individuals and inanimate
316 objects, such as algae and a fish gut (Figure 3E-F). We recorded males often embracing
317 other males. When this happens, the embraced male emits a release call, causing the
318 clasping male to release him.



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Figure 3. *Melanophryniscus admirabilis* breeding activity and behaviour at the breeding site: (A) Some individuals at a small temporary pool in the rocky river bank outcrop; (B) a male calling; (C) a pair in axillary amplexus in the water; (D) struggle between three males and an amplexant pair; (E) necrophiliac amplexus; (F) a male embracing an alga; (G) an interspecific amplexus with a juvenile of *Rhinella icterica*;

325 (H) a pair spawning in a pool with other clutches from other pairs; (I) a newly released
326 spawning with about 19 eggs; (J) three clutches released at different times; (K) tadpoles
327 almost hatching from clutch; (L) tadpoles at Gosner (1960) stage 27-29; (M) tadpoles at
328 Gosner (1960) stages 30-31, showing evident golden iridophores. Photos: Rômulo
329 Silveira (A), Valentina Caorsi (B), Simone Leonardi (H), Michelle Abadie (C–G, I–M).
330

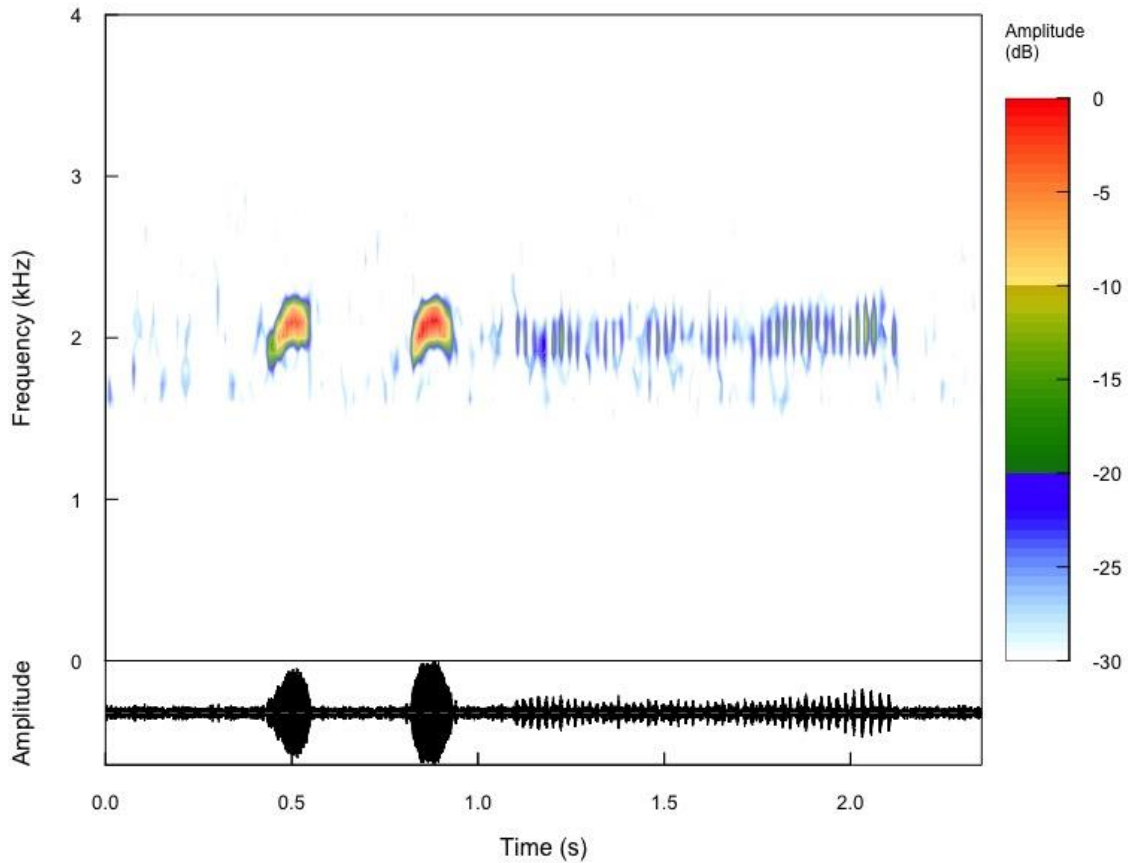
331 We followed two amplexus until the female finished oviposition, both events
332 during the day (January 2011 and October 2016). In the first one, we encountered the
333 amplexant pair in a small shaded pool. While in amplexus, the female remained still
334 with only her head sticking out of the water. Her feet and hands remained resting on the
335 pool's surface. The male contracted his abdomen repeatedly (about one contraction per
336 second). At a certain point, the female dived, taking the male into the water. The male
337 then started rubbing his feet, collecting the eggs from the female's cloaca, and leading to
338 his cloaca. Following this, the male left the eggs in submerged substrate (about 20
339 eggs). The female, still carrying the same male, left this pool and the process began
340 again in another one. After they deposited the second egg clutch, we collected this
341 amplexant pair in a plastic bag with water and they deposited two more clutches inside
342 the bag, one with 31 eggs and another with 20 eggs. We recorded the second amplexant
343 pair (Supplementary Video S6) when the female was spawning, and the male was
344 collecting submerged leaves in the water with his hind feet and leading them to his
345 cloaca to deposit the eggs. Even after laying the eggs, the female continued to do wavy
346 abdominal movements until leaving the pool, carrying the male.

347 *Melanophryniscus admirabilis*' egg clutch is transparent, formed by gelatinous
348 egg masses which are hatched inside the small, shallow pools on the volcanic flattened
349 river bank outcrop, frequently adhered to vegetation, fallen leaves or just floating on the
350 water. Immediately after oviposition, eggs were very close to each other (Figure 3I). A
351 couple of hours later, they swelled to a point where they could be distinguished by eye.

352 We counted *in situ* five eggs as the minimum and 36 eggs as the maximum in each
353 clutch ($n = 17$; mean = 15.4 ± 6.6). After about four or five days the tadpoles had begun
354 to hatch from the eggs (Figure 3L). In the lab, we observed that on the 10th day from the
355 hatching, the hind limbs had begun to appear (Gosner stage 26-30), and the forelimbs
356 three days after. The tail resorption began on the 14th day from the hatching and was
357 completed on the 15th day. On the 16th day, the metamorphosis was complete in all
358 individuals, making a total of 20-21 days for the full development (egg-to-juvenile).

359 3.3 Advertisement Call

360 Call recordings were carried out along the breeding site under air temperatures
361 ranging between 17.5 and 23.9 °C. The advertisement call of *M. admirabilis*, based on
362 four individuals (total of nine calls), has a duration of 21 ± 1.5 seconds and it is
363 comprised of segments A and B (Figure 4; Supplementary Table S7). Part A has 2–19
364 single frequency modulated notes (0.1 seconds each), each one comprising one pulse
365 separated by intervals of 2 ± 2.4 seconds. Notes in part A present frequency modulation
366 increasing from 1.92 ± 0.13 kHz to 2.04 ± 0.17 kHz, reaching the peak frequency on the
367 second third of the note, then decreasing to 1.98 ± 0.17 kHz. Call part B is a train of
368 unmodulated pulses emitted at a rate of 50.8 ± 2.0 pulses per second, with short time
369 intervals (0.014 ± 0.002), and lasting from 1 to 6.26 (4.1 ± 1.7) seconds. The interval
370 between call part A and B is on average 1.5 ± 4.1 seconds. The peak frequency is on
371 average 1.9 ± 0.38 kHz in part A and 1.7 ± 0.22 kHz in part B.



372

373 **Figure 4.** Advertisement call of *Melanophryniscus admirabilis*: spectrogram and
 374 oscillogram of two notes from the first segment (call part A) and the second segment
 375 (call part B).

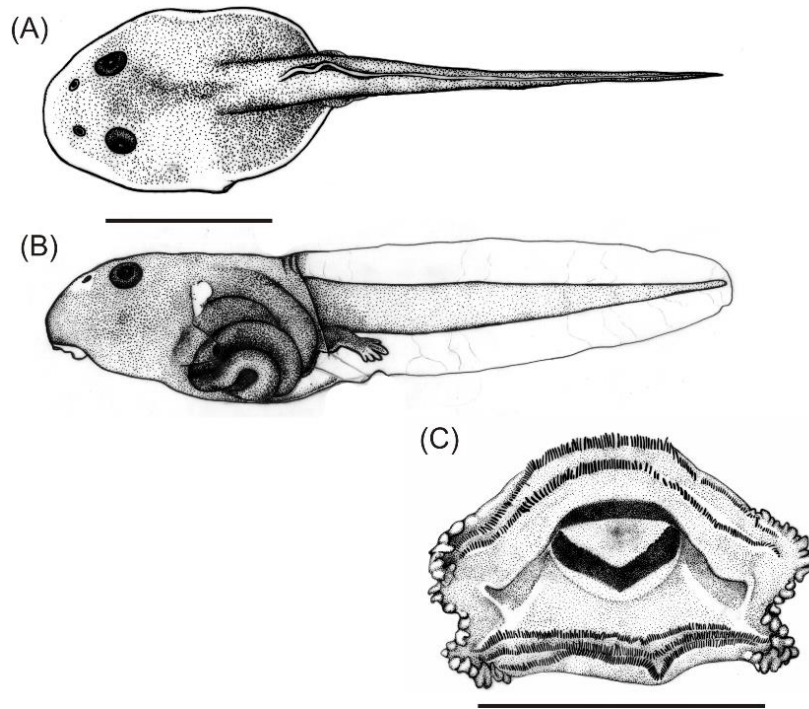
376

377 3.4 Tadpoles

378 The total length of *M. admirabilis* tadpoles (Figure 5) at stages 32 – 37 was
 379 between 13.9 and 17.6 mm (Supplementary Table S8). Body shape slightly depressed
 380 (BMH/BMW= 0.85 ± 0.06 mm) and oval in dorsal view; body length approximately
 381 40% of the total length, body width approximately 72% of the body length, and body
 382 height approximately 61% of the body length; the maximum body width was just
 383 behind the eyes. The snout was rounded in dorsal and lateral views, and the gular region
 384 was flat. Nostrils rounded, positioned dorsally with internarial distance about 34% of
 385 the body width, visible in frontal, dorsal and lateral views, opening anterolaterally, with

386 a small projection on the marginal rim all around. Eyes were medium ($E/BWE = 0.19 \pm$
387 0.02 mm) and placed dorsolaterally (interorbital distance approximately 49% of the
388 body width). The pineal end-organ, present as an unpigmented spot between the anterior
389 edges of the eyes, was almost imperceptible. The spiracle was sinistral, single, very
390 short, opening at the end of the middle third of the body ($RSD/BL = 0.63 \pm 0.04$ mm),
391 at the lateral midline, and directed posterodorsally, with the inner fused to the body
392 wall. The gut loop (“point de rebroussement”, *sensu* Hourdry & Beaumont, 1985) is
393 placed at the left of the abdomen. The vent tube was conical, short, running along its
394 right side and associated with the ventral fin; the vent opening was dextral. The tail was
395 straight, medium-sized ($TAL/TL = 0.60 \pm 0.02$ mm) and slightly lower than the body
396 ($MTH/BMH = 0.82 \pm 0.07$ mm); myomeres were evident through the whole caudal
397 musculature extension, narrowing gradually toward the end of the tail, not reaching the
398 rounded tip of the tail; the dorsal fin was originated before the body-tail junction and
399 reached its maximum height at the middle region, staying almost constant until
400 converging with the ventral fin, at the end of the tail; the ventral fin was originated on
401 the left side of the vent tube and its height was almost uniform until the tip.

402 The oral disc was anteroventral, not visible dorsally, with almost 50% of the
403 body width and laterally emarginate (Figure 5, Supplementary Table S8). The marginal
404 papillae were arranged in a single row, with wide rostral and medium-sized ventral gaps
405 (66% and 50% of the oral disc, respectively). Labial tooth row formula (LTRF) was 2/3
406 in all analysed specimens ($n=20$). The upper jaw sheath was slightly curved, the lower
407 sheath was smaller and V-shaped, both serrated.



408

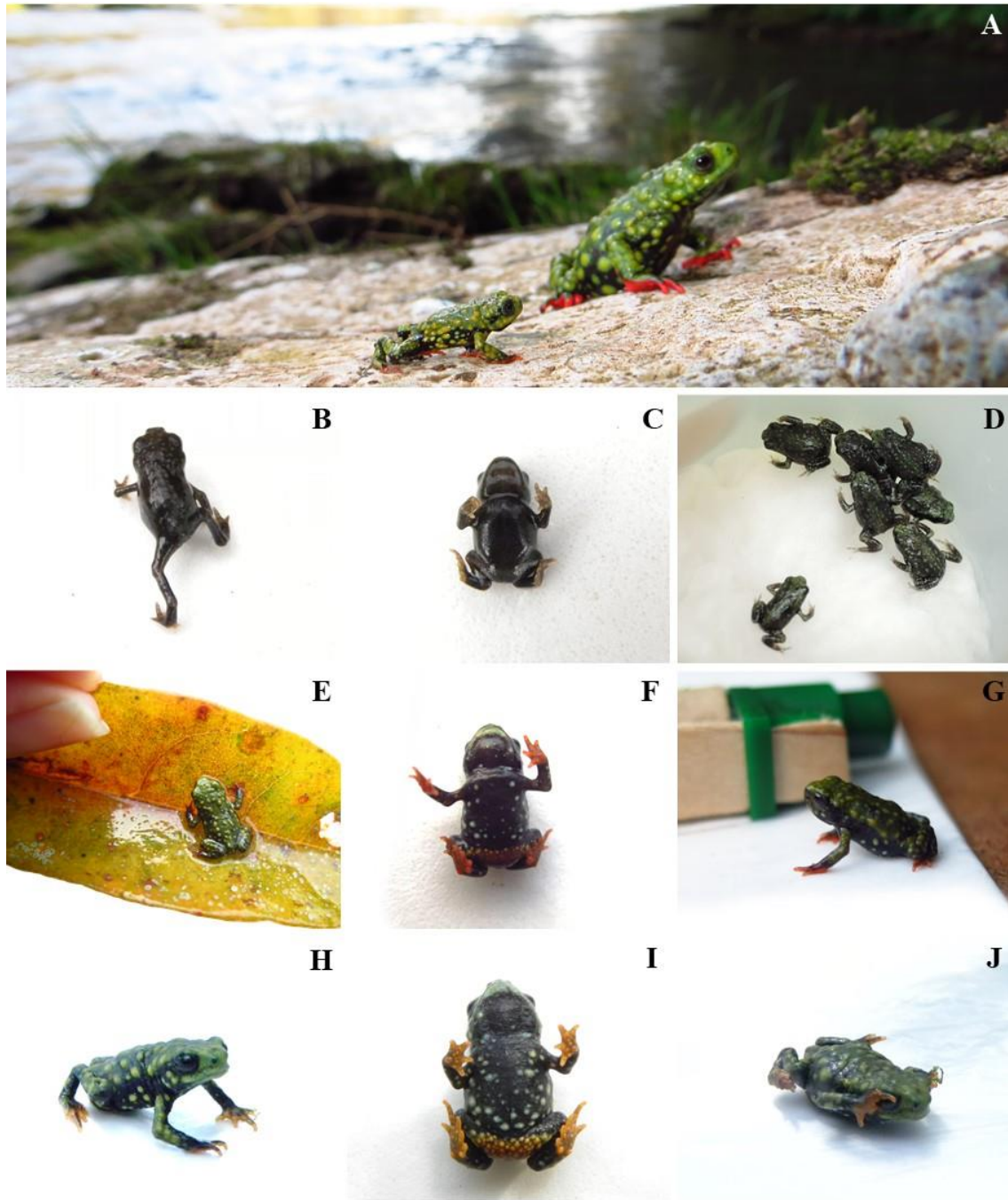
409 **Figure 5.** External morphology and oral disc illustration of the tadpole of
 410 *Melanophryniscus admirabilis*. Gosner (1960) Stage 37. (A) Dorsal view and (B) lateral
 411 view (scale bar = 5 mm). (C) Oral disc (scale bar = 1 mm). *Drawing by Marcelo Costa.*

412

413 The colouration of tadpoles in life was almost uniformly brown and opalescent
 414 immediately after hatching (Figure 3L). After stage 32, the dorsum colouration of the
 415 tadpoles was dark brown (melanophores, *sensu* Gosner 1960) with spreading small
 416 golden dots (iridophores, *sensu* Gosner 1960). The dorsal fin was translucent with
 417 scattered brown spots; the ventral fin was translucent and unpigmented. Caudal
 418 musculature with patches of brown and golden dots, especially in dorsal myosepts. The
 419 venter was translucent with scattered brown spots, except for the gular region which
 420 was almost entirely translucent. In preserved specimens, the golden dots disappeared,
 421 the skin was translucent, and the dorsal colouration was dark and opaque brown.

422 *3.5 Juvenile and adult colouration and defensive behaviour*

423 The colouration of the individuals' skin varies ontogenetically (Figure 6;
424 Supplementary Figure S9). Newly metamorphosed individuals have a dark green
425 (almost black) dorsum, translucent pale palmar and plantar surfaces and black, partially
426 translucent venter (Figure 6B-C). Two to three days after metamorphosis, we observed
427 the beginning of the development of dorsal glands and the emergence of green
428 pigmentation (Figure 6D). We do not know when, in the life of an individual, the red
429 colouration and the yellowish-green ventral spots (glands) emerge, but we do know this
430 process is gradual (Figure 6C-F-I). The palmar, plantar and pelvic surfaces remain
431 opaque (Figure 6F-I), varying from pale orange to dark red (but still translucent) until
432 adulthood, when the red colouration is bright and conspicuous (Supplementary Figure
433 S9). Even without presenting the brightness of the red aposematic colouration, young
434 individuals can perform the *unken reflex* behaviour (Figure 6J).



435 **Figure 6.** The colouration of juveniles of *Melanophryniscus admirabilis*. (A) A juvenile
 436 and an adult at the breeding site; (B–C) a newly metamorphosed individual without
 437 colouration, found at the breeding site; (D) a few individuals about four days after
 438 metamorphosis, presenting dorsal glands and some green pigmentation; (E–G) a
 439 juvenile individual, presenting conspicuous green colouration, yellowish-green dorsal
 440 glands, opaque red pigmentation on the palmar, plantar and pelvic surface and few
 441 ventral glands (green-bluish ventral dots); (H–J) a juvenile individual (11.04 mm and
 442 0.2 g), presenting more ventral glands than the previous juvenile, still opaque and

443 slightly translucent red colouration and performing the *unken reflex* behaviour. Photos:
444 Luis Fernando Marin da Fonte (A), Michelle Abadie (B–J).

445

446 The dorsal colouration of adults does not vary widely, being bright green and,
447 sometimes, presenting greyish-blue patches and/or yellow spots (more rarely red spots).
448 The ventral surface of adults is a black background with well-defined yellowish-green
449 spots (glands), sometimes with red or greyish-blue spots. There are no evident
450 differences in the colour pattern of males and females.

451 The Admirable toads are hard to see in the green vegetation, especially against
452 green mosses. Notably, it was common, even to trained collectors, to misidentify
453 mosses and fallen green leaves as toads (Figure 7A - B). *Melanophryniscus admirabilis*
454 exhibited the *unken reflex* when disturbed (Figure 7C), usually after trying to escape or
455 freezing, often leaving the eyes uncovered.



456

457 **Figure 7.** Colouration and behaviour of *Melanophryniscus admirabilis*. (A) Two
458 individuals on green moss at the breeding site; (B) an individual and a leaf with galls,
459 often encountered at the breeding site; (C) an individual performing *unken reflex*
460 behaviour, showing the red colouration on the palmar and plantar surfaces. Photos:
461 Michelle Abadie.

462

463 4. Discussion

464 In this study, we gathered all unpublished information compiled from seven
465 years and 122 days of *in situ* observation and monitoring of the only known population

466 of *M. admirabilis*. We provide the most up-to-date and comprehensive synthesis of
467 morphological, behavioural and other life-history information for the species. We
468 expect this will lead to new research on the evolutionary origin and ecological relevance
469 of the described traits for the species and stimulate comparative studies within the genus
470 *Melanophryniscus*.

471 *Melanophryniscus admirabilis* exhibits a seasonal explosive breeding pattern
472 and similar reproductive behaviour described for other *Melanophryniscus* species
473 (Baldo et al., 2014), such as aggregation of hundreds of individuals, active searching
474 and tactile identification (trial-and-error clasping behaviour), amplexus with inanimate
475 objects or anything that moves, intense male combat, struggling including non-
476 amplexant and amplexant males, diurnal and nocturnal breeding activity, breeding in
477 temporary rain pools or other ephemeral habitats (Maneyro and Carreira, 2012; Cairo et
478 al., 2013, 2008; Caldart et al., 2013; Caorsi et al., 2020, 2014; Pereira and Maneyro,
479 2018; Santos et al., 2010; Silva et al., 2009; Vaira, 2005). In this study, we recorded
480 evidence of breeding activity in almost all months of the year. Nevertheless, the
481 explosive breeding events, characterised by the high density of individuals at the
482 breeding site and the typical associated behaviour, were not uniform throughout the year
483 and were concentrated between mid-July and early December. Some geographically
484 close species of *Melanophryniscus* (*M. cambaraensis*, Santos et al., 2010; *M.*
485 *pachyrhynchus*, Caldart et al., 2013; *M. macrogranulosus*, Caorsi et al., 2014) are
486 aseasonal explosive breeders. Wells (1977) refers to the prolonged and explosive
487 breeders as two ends of a continuum, and *M. admirabilis* is an example that appears to
488 be somewhere in the middle.

489 Seasonality is reported for most *Melanophryniscus* species for which the
490 reproductive cycle was described (*M. moreirae*, Van Sluys and Guido-Castro, 2011; *M.*

491 *rubriventris*, Vaira, 2005; *M. stelzneri*, Pereyra et al., 2011) and it is always associated
492 with the seasonality of rainfall in those regions and the warm seasons in the austral
493 hemisphere. In the Perau de Janeiro region, monthly rainfall is well distributed over the
494 year (Alvares et al., 2013; Zepner et al., 2020) and the four seasons are well-defined,
495 with hot summers and cold winters, and mild springs and autumns. In these latitudes,
496 environmental seasonality is mostly determined by variation in temperature and
497 photoperiod (Both et al., 2008). We encourage future studies to simultaneously monitor
498 rainfall, temperature, photoperiod and reproductive activity to identify triggering factors
499 of the breeding activity of *M. admirabilis*.

500 Admirable toads use small shallow temporary pools in riverbanks as oviposition
501 sites, and they need to cope with a high risk of reproductive failure through desiccation
502 or flooding of pools, risking both clutches and tadpoles. Species reproducing in these
503 unpredictable and ephemeral environments have at least three not mutually exclusive
504 strategies to increase offspring survival: (1) not release all eggs in a single clutch, but
505 release them in a sequence of clutches, spreading several egg masses to different sites in
506 the same pool and different pools at the breeding site; (2) rapid larval development; and
507 (3) select more favourable pools. The first two strategies have already been reported for
508 other *Melanophryniscus* species (*M. rubriventris*, Goldberg et al., 2006; *M. stelzneri*,
509 Bustos Singer and Gutiérrez, 1997; *M. montevidensis*, Pereira and Maneyro, 2018;
510 *Melanophryniscus* sp., Cairo et al., 2008; *M. cambaraensis*, Caorsi et al., 2012).
511 Spreading eggs in several sequential clutches has the potential to increase the
512 probability that at least one spawning survives (Spieler and Linsenmair, 1997). Since
513 temperature influences the larval growth rate and development (Hayes et al., 1993;
514 Smith-Gill and Berven, 1979), direct or indirect sun warming of the water in pools may
515 help to accelerate the development of tadpoles and, consequently, reduce the risks

516 associated with water dependence. According to Goldberg et al. (2006), toads of *M.*
517 *rubriventris* preferred warmer flooded pools. On the other hand, increasing temperature
518 also increases the pool desiccation rate. Thus, we could expect that recruitment would
519 be higher in pools that have the most favourable balance between development rate and
520 mortality risk. All these aspects are open questions that should be explored in future
521 studies. They can help explain why other available rocky outcrops along the river are
522 not used for reproduction by *M. admirabilis*.

523 Even though it is the second-largest species of the genus, *M. admirabilis* does
524 not have the largest tadpoles (BL= 6.33±0.46; see Table on Baldo et al., 2014). Like
525 other *Melanophryniscus* species from the *stelzneri* group (Baldo et al., 2014), *M.*
526 *admirabilis* exhibits one of the most basal reproductive modes found in anurans, which
527 relies on lentic water bodies for spawning and tadpole development (Duellman and
528 Trueb, 1994). Its tadpole is similar to other lentic tadpoles of the genus (*M. stelzneri*
529 group; see Baldo et al., 2014), but the tail is slightly lower than body, the oral disc is
530 wide and dorsal and ventral gaps are large, as reported for *M. rubriventris* (Baldo et al.,
531 2014; Lavilla and Vaira, 1997). The tadpole of the Admirable Redbelly Toad has
532 intermediate features among phenetic groups of the genus (*sensu* Caramaschi and Cruz,
533 2002), and an analysis of the body shape through geometric morphometrics (Haad et al.,
534 2011) probably would put it closer to *M. rubriventris* (*M. stelzneri* group) or *M.*
535 *devincenzii* (*M. tumifrons* group) (Baldo et al., 2014). As concluded by Baldo et al.
536 (2014), *Melanophryniscus* tadpoles are very similar and the differences observed are
537 mainly quantitative.

538 Similar to all *Melanophryniscus* species, the advertisement call of the Admirable
539 Toad is structured in two parts: call A, comprised of notes of single pulses; and call B, a
540 pulsed trill. The advertisement call is complex and has modulated frequency and may be

541 assigned to Call Guild H (Emmrich et al., 2020). *Melanophryniscus admirabilis*
542 presents one of the longest known advertisement calls among the species of the genus
543 (see a comparative table in Caorsi et al., 2020), but while the long duration of the others
544 is a result of the long call B, in *M. admirabilis* it is due to long silence intervals between
545 notes of call A and, similar to species of the *M. stelzneri* group, it has a short call B.
546 Call A appears to have substantial variation in terms of the number of notes, the duration
547 of each one and the length of the gap between them. During the performance of call A,
548 we sometimes observed males alternating note emission with small movements,
549 changing position on the same site, seemingly choosing the direction of the call.
550 Moreover, we found some males calling inside burrows and rock crevices, especially
551 during the day. This behaviour may be related to sound amplification, similar to that
552 already described for other species that call in burrows (Penna, 2004; Penna and
553 Hamilton-West, 2007). The torrent water of the Forqueta river introduces background
554 noise that could mask certain frequency components of anuran calls and it could be
555 expected that these toads presented higher dominant frequencies than other non-stream-
556 dwelling *Melanophryniscus* spp. (Röhr et al., 2016 but see Vargas-Salinas and
557 Amézquita, 2014), but Admirable toads appear to have other adaptations to this noisy
558 environment. According to Goutte et al. (2018), torrent-dwelling species produced calls
559 with more pronounced frequency modulations, being positively correlated with the
560 ambient noise level. As far as we know, the notes of the call A we described here for *M.*
561 *admirabilis* have the most modulated frequency for the genus, possibly being related to
562 the very noisy environment that this species occurs. Besides, the call A of *M.*
563 *admirabilis* presented on average one out of the longest notes (0.10 s) among
564 *Melanophryniscus* species, similar to *M. atroluteus* (0.102 s; Baldo and Basso, 2004)
565 and *M. stelzneri* (0.11 s; Barrio, 1964), while it is shorted in *M. dorsalis* (0.054 s and

566 0.042 s; Kwet et al, 2005) and *M. montevidensis* (0,0313 s; Kwet et al, 2005). In
567 general, the advertisement call of *M. admirabilis* is more similar to calls of the *M.*
568 *stelzneri* group. However, given the lack of phylogenies for this genus, any hypothesis
569 about the relations and explanations of acoustic patterns for these species remain
570 inconclusive.

571 Regarding sexual dimorphism, our results showed that males are smaller than
572 females, as expected (female-biased sexual size dimorphism; Bidau et al., 2011).
573 However, the body size alone is not an efficient parameter to identify the sex, since the
574 ranges of size overlap at the extremes. Larger female body sizes are a pattern found in
575 most anurans (Shine, 1979) and several explanations have already been proposed for
576 this phenomenon, such as the capacity to produce bigger clutches or oocytes (Camargo
577 et al., 2008; Kuramoto, 1978), mainly in small-bodied and explosive breeding species
578 (Nali et al., 2014), or older age at first reproduction (Halliday and Verrell, 1988;
579 Guimarães et al., 2011). Jeckel et al. (2015) found that females are larger and older than
580 males in *M. moreirae*, suggesting that the different age structure of each sex could
581 explain the sexual size dimorphism, as proposed by Monnet and Cherry (2002).
582 However, in that study, females were still larger than males when the toads were
583 separated by age cohorts, demonstrating that there is a sexual size dimorphism-related
584 effect. Although both sexes can exhibit nuptial pads in some *Melanophryniscus* species
585 (Jeckel et al., 2019), in our study we easily differentiated males from females using the
586 presence of brownish nuptial pads at the base of the male thumb. We did not find any
587 evidence of sexual colour dimorphism by eye, but a further interesting approach could
588 be to investigate other parameters to identify gender.

589 The Admirable Redbelly Toad has changing ontogenetic colouration, and the
590 bright green and red colours are conspicuous only in adulthood. A diet rich in

591 carotenoids may result in a more saturated and brighter colour (Umbers et al., 2016).
592 Carotenoids are largely responsible for the red, orange, and yellow colours in
593 amphibians (Hoffman and Blouin, 2000) and must be acquired from the diet (Goodwin,
594 1984; Umbers et al., 2016). The red colouration in the genus *Melanophryniscus* comes
595 from the accumulation of this pigment (Bonansea et al., 2017) and it is supposedly
596 aposematic (Bonansea and Vaira, 2012, but see Bordignon et al., 2018). Together with
597 lipophilic alkaloids obtained from an arthropod-rich diet (Hantak et al., 2013; Saporito
598 et al., 2012, 2009), the aposematic colouration could be a result of the diurnal activity
599 and could provide protection against visually oriented diurnal predators (Santos and
600 Grant, 2010), especially when diurnal toads are breeding and are more exposed.
601 Moreover, the bright green colouration acquired over a juvenile's lifetime may work as
602 crypsis, reducing the animal's detection rate and protecting against visual predators as
603 well (see Dallagnol Vargas et al., 2020). The ontogenetic modification of colour has
604 already been observed in other *Melanophryniscus* species (*M. macrogranulosus*, Caorsi
605 et al., 2014; *M. klappenbachi*, Bland, 2015; *M. montevidensis*, Bardier et al., 2017), but
606 how and why this happens remains unclear.

607 We recorded the longest lifespan directly observed in the field for the genus
608 *Melanophryniscus* to date. Our results reveal that Admirable Redbelly toads can live at
609 least seven years in the wild, which represents our entire sampling window. We
610 recaptured 25 individuals in 2017, seven years after the first capture (2010), when they
611 were already adults. Additionally, in 2020 we occasionally recaptured three individuals
612 captured for the first time in 2011, when they were already adults (Michelle Abadie,
613 pers. obs.). Minimal lifespan has been estimated for another species of the genus, *M.*
614 *moreirae*, using skeletochronology. Jeckel et al. (2015) reported adult female and male
615 ages ranging from 3-6 and 2-6 years, respectively. In the same study, they recorded

616 juvenile (all females) ages ranging from 1 to 5 years. These results suggest that
617 *Melanophryniscus* individuals reach sexual maturity at least 1 year after metamorphosis,
618 possibly more. Therefore, we may assume that *M. admirabilis* individuals can
619 potentially live at least 10 years. The maximum lifespan of these two species suggests
620 that *Melanophryniscus* species can be long-lived anurans, as observed for some other
621 bufonids (e.g. La Marca, 1984; Lyapkov et al., 2020; see Carey and Judge, 2008).
622 Chemical defences can play a role in this unexpected longevity for a small, subtropical
623 and diurnal toad (Stark and Meire, 2018). Grant et al. (2012) and Jeckel et al. (2015)
624 found that the larger (and older) the redbelly toad, the greater the diversity of alkaloids
625 found in the skin and internal organs. Thus, we hypothesise that the longer they take to
626 mature, the greater their protection from predators.

627 The high rate of recaptures in the same sector (within the breeding site) suggests
628 site fidelity, with juveniles being less loyal than adults. Our results are in accordance
629 with the long-standing assumption in the amphibian literature that juveniles are more
630 likely to disperse than adults (Dole, 1971; Wells, 2010) although we cannot affirm that
631 these movements configure dispersal. Dispersal in the juvenile stage may be a response
632 to pressures such as mate availability, inbreeding avoidance, and conspecific
633 competition for resources (Greenwood, 1980; Hamilton and May, 1977; Perrin and
634 Mazalov, 2000; Dobson, 1982), as if they were looking for a breeding site to establish
635 themselves. Other studies have already demonstrated site fidelity of *Melanophryniscus*
636 species (Pereira and Maneyro, 2016, 2018). We observed fights between males while
637 they were calling at the breeding site, which may be evidence of territoriality or site
638 fidelity, but future studies are needed to confirm this. We found exceptions of adults
639 who displaced up to 300 m and we hypothesise that they might be newly recruited

640 individuals. In any case, it is the maximum distance ever recorded for a species of the
641 genus and it seems to be substantial for such a small species.

642 **5. Conclusions**

643 The Admirable Redbelly toad is a seasonal explosive breeder that lives in noisy
644 and risky conditions and seems to have adaptations to deal with this environment. In
645 this study, we described the advertisement call of the species, which the notes of call
646 part A have the most modulated frequency for the genus; the eggs are deposited in
647 several clutches in different temporary pools; the tadpole has a rapid development and is
648 one of the smallest of the genus, whereas the adults are one of the largest; the species
649 has supposedly aposematic and cryptic colouration and observed longevity extended at
650 least 9 years. Our study considerably increases knowledge about the Admirable
651 Redbelly Toad, a microendemic and Critically Endangered anuran (IUCN SSC
652 Amphibian Specialist Group, 2013). The information we present here can be used in the
653 future to evaluate the risks to the persistence of the single-known population, guide the
654 search for new populations, discuss management actions to protect the reproductive site
655 and stimulate research on the species' reproductive patterns and the mechanisms that
656 trigger them.

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675 **References**

- 676 Agostinelli, C., Lund, U., 2017. R package “circular”: Circular Statistics (version 0.4-
677 93).
- 678 Altig, R., McDiarmid, R.W., 1999. Body plan: Development and morphology, in:
679 McDiarmid, R.W., Altig, R. (Eds.), Tadpoles: The Biology of Anuran Larvae. The
680 University of Chicago Press, Chicago, Illinois, USA, pp. 24–51.
- 681 Alvares, C.A., Stape, J.L., Sentelhas, P.C., De Moraes Gonçalves, J.L., Sparovek, G.,
682 2013. Köppen’s climate classification map for Brazil. Meteorol. Zeitschrift 22,
683 711–728. <https://doi.org/10.1127/0941-2948/2013/0507>
- 684 Baldo, D., Basso, N.G., 2004. A New Species of *Melanophryniscus* (Anura:
685 Bufonidae), with Comments on the Species of the Genus Reported for Misiones,
686 Northeastern Argentina. J. Herpetol. 38(3), 393-403. [https://doi.org/10.1670/144-](https://doi.org/10.1670/144-03A)
687 03A
- 688 Baldo, D., Candiotti, F.V., Haad, B., Kolenc, F., Borteiro, C., Pereyra, M.O., Zank, C.,
689 Colombo, P., Bornschein, M.R., Sisa, F.N., Brusquetti, F., Conte, C.E., Nogueira-
690 Costa, P., Almeida-Santos, P., Pie, M.R., 2014. Comparative morphology of pond,
691 stream and phytotelm-dwelling tadpoles of the South American Redbelly Toads
692 (Anura: Bufonidae: *Melanophryniscus*). Biol. J. Linn. Soc. 112, 417–441.
693 <https://doi.org/10.1111/bij.12296>
- 694 Bardier, C., Martínez-Latorraca, N., Porley, J.L., Bortolini, S.V., Cabrera Alonzo, N.,
695 Maneyro, R., Toledo, L.F., 2019. Seasonal demography of the threatened
696 Montevideo Redbelly Toad (*Melanophryniscus montevidensis*) in a protected area
697 of Uruguay. Can. J. Zool. 97, 131–141. <https://doi.org/10.1139/cjz-2017-0362>
- 698 Bardier, C., Pereira, G., Elgue, E., Maneyro, R., Toledo, L.F., 2017. Quantitative
699 determination of the minimum body size for photo-identification of

700 *Melanophryniscus montevidensis* (Bufonidae). Herpetol. Conserv. Biol. 12, 119–
701 126.

702 Barrio, A. 1964. Peculiaridades del canto nupcial de *Melanophryniscus stelzneri*
703 (Weyenbergh) (Anura: Brachicephalidae). Physis, 24, 435–437.

704 Bidau, C.J., Martí, D.A., Baldo, D., 2011. Inter- and Intraspecific Geographic Variation
705 of Body Size in South American Redbelly Toads of the Genus *Melanophryniscus*
706 Gallardo, 1961 (Anura: Bufonidae). J. Herpetol. 45, 66. <https://doi.org/10.1670/09-202.1>

707

708 Bioacoustics Research Program. 2011. Raven Pro: Interactive Sound Analysis Software
709 (Version 1.4) [Computer software]. The Cornell Lab of Ornithology, Ithaca, NY.
710 Available from: <http://www.birds.cornell.edu/raven> (accessed January 2021)

711 Bland, A.W., 2015. A record of captive reproduction in the Red bellied toad
712 *Melanophryniscus klappenbachi* with notes on the use of a short-term brumation
713 period. Herpetol. Bull. 131, 15–18.

714 Bolger, D.T., Morrison, T.A., Vance, B., Lee, D., Farid, H., 2012. A computer-assisted
715 system for photographic mark-recapture analysis. Methods Ecol. Evol. 3, 813–822.
716 <https://doi.org/10.1111/j.2041-210X.2012.00212.x>

717 Bonansea, M.I., Vaira, M., 2012. Geographic and intrapopulation variation in colour
718 and patterns of an aposematic toad, *Melanophryniscus rubriventris* 33, 11–24.
719 <https://doi.org/10.1163/156853811X619754>

720 Bonansea, M.I., Heit, C., Vaira, M., 2017. Pigment composition of the bright skin in the
721 poison toad *Melanophryniscus rubriventris* (Anura: Bufonidae) from Argentina.
722 Salamandra.

723 Bordignon, D.W., Caorsi, V.Z., Colombo, P., Abadie, M., Brack, I.V., Dasoler, B.T.,
724 Borges-Martins, M., 2018. Are the unken reflex and the aposematic colouration of
725 red-bellied toads efficient against bird predation? PLoS One 13, e0193551.
726 <https://doi.org/10.1371/journal.pone.0193551>

727 Bordignon, D.W. 2019. Por que aqui e não ali? Seleção do Habitat Reprodutivo pelo
728 Sapinho-admirável-de-barriga-vermelha, uma Espécie Microendêmica e
729 Criticamente Ameaçada. MSc Dissertation, Universidade Federal do Rio Grande
730 do Sul, Porto Alegre/RS, 57 pp.

731 Both, C., Kaefer, Í.L., Santos, T.G., Cechin, S.Z., 2008. An austral anuran assemblage
732 in the Neotropics: seasonal occurrence correlated with photoperiod. J. Nat. Hist.
733 <https://doi.org/10.1080/00222930701847923>

734 Bustos Singer, R., Gutiérrez, M., 1997. Reproducción y desarrollo larval del sapo enano
735 *Melanophryniscus stelzneri stelzneri* (Weyenberg, 1875) (Anura: Bufonidae).
736 Cuad. Herpetol. 11, 21–30.

737 Cairo, S.L., Zalba, S.M., Úbeda, C.A., 2013. Reproductive pattern in the southernmost
738 populations of South American redbelly toads. J. Nat. Hist. 47, 2125–2134.
739 <https://doi.org/10.1080/00222933.2013.769644>

740 Cairo, S.L., Zalba, S.M., Úbeda, C.A., 2008. Reproductive behaviour of
741 *Melanophryniscus sp.* from Sierra de la Ventana (Buenos Aires, Argentina). South
742 Am. J. Herpetol. 3, 10–14. [https://doi.org/10.2994/1808-9798\(2008\)3\[10:RBOMSF\]2.0.CO;2](https://doi.org/10.2994/1808-9798(2008)3[10:RBOMSF]2.0.CO;2)

743

744 Caldart, V.M., dos Santos, T.G., Maneyr, R., 2013. The advertisement and release calls
745 of *Melanophryniscus pachyrhynus* (Miranda-Ribeiro, 1920) from the central region
746 of Rio Grande do Sul, southern Brazil. Acta Herpetol. 8, 115–122.
747 https://doi.org/10.13128/Acta_Herpetol-12680

748 Camargo, A., Sarroca, M., Maneyro, R., 2008. Reproductive effort and the egg number
749 vs. size trade-off in *Physalaemus* frogs (Anura: Leiuperidae). Acta Oecologica 34,

750 163–171. <https://doi.org/10.1016/j.actao.2008.05.003>

751 Caorsi, V., Abadie, M., Santos, R. F., Colombo, P., Borges-Martins, M., (*in prep*).
752 Long-term colour pattern study in South American Red-Bellied Toads (Genus
753 *Melanophryniscus*). Herpetol. J.

754 Caorsi, V., Bordignon, D.W., Márquez, R., Borges-Martins, M., 2020. Advertisement
755 call of two threatened red-bellied-toads, *Melanophryniscus cambaraensis* and *M.*
756 *macrogranulosus* (Anura: Bufonidae), from the Atlantic Rainforest, southern
757 Brazil. Zootaxa 4894, 206–220. <https://doi.org/10.11646/zootaxa.4894.2.2>

758 Caorsi, V., Colombo, P., Freire, M.D., Amaral, I.B., Zank, C., Borges-Martins, M.,
759 Grant, T., 2014. Natural history , coloration pattern and conservation status of the
760 threatened South Brazilian red bellied toad, *Melanophryniscus macrogranulosus*
761 Braun , 1973. Herpetol. Notes 7, 585–598.

762 Caorsi, V.Z., Santos, R.R., Grant, T., 2012. Clip or Snap? An Evaluation of Toe-
763 Clipping and Photo-Identification Methods for Identifying Individual Southern
764 Red-Bellied Toads, *Melanophryniscus cambaraensis*. South Am. J. Herpetol. 7,
765 79–84. <https://doi.org/10.2994/057.007.0210>

766 Caramaschi, U., Cruz, C.A., 2002. Taxonomic status of *Atelopus pachyrhynus*
767 Miranda-Ribeiro, 1920, redescription of *Melanophryniscus tumifrons* (Boulenger,
768 1905), and descriptions of two new species of *Melanophryniscus* from the state of
769 Santa Catarina, Brazil (Amphibia, Anura, Bufonidae). Arq. Mus. Nac. 61, 195–
770 202.

771 Carey, J.R., Judge, D.S., 2008. Longevity Records: Life Spans of Mammals, Birds,
772 Amphibians, Reptiles, and Fish, Monographs on Population Aging, 8. Odense
773 University Press. Available in <https://www.demogr.mpg.de/longevityrecords/>.
774 Accessed in: March, 2021.

775 Conde, D.A., Staerk, J., Colchero, F., da Silva, R., Schöley, J., Baden, H.M., Jouvett, L.,
776 Fa, J.E., Syed, H., Jongejans, E., Meiri, S., Gaillard, J.-M., Chamberlain, S.,
777 Wilcken, J., Jones, O.R., Dahlgren, J.P., Steiner, U.K., Bland, L.M., Gomez-
778 Mestre, I., Lebreton, J.-D., González Vargas, J., Flesness, N., Canudas-Romo, V.,
779 Salguero-Gómez, R., Byers, O., Berg, T.B., Scheuerlein, A., Devillard, S., Schigel,
780 D.S., Ryder, O.A., Possingham, H.P., Baudisch, A., Vaupel, J.W., 2019. Data gaps
781 and opportunities for comparative and conservation biology. Proc. Natl. Acad. Sci.
782 116, 9658–9664. <https://doi.org/10.1073/pnas.1816367116>

783 Crump, M.L., Scott Jr., N.J., 1994. Standard techniques for inventory and monitoring:
784 Visual Encounter Surveys., in: Heyer, W.R., Donnelly, M.A., McDiarmid, R.W.,
785 Hayek, L.C., Foster, M.S. (Eds.), Measuring and Monitoring Biological Diversity.
786 Standard Methods for Amphibians. Smithsonian Institution Press, Washington DC,
787 pp. 84–92.

788 da Silva, P.R., Borges-Martins, M., Oliveira, G.T., 2021. *Melanophryniscus admirabilis*
789 tadpoles’ responses to sulfentrazone and glyphosate-based herbicides: an approach
790 on metabolism and antioxidant defenses. Environ. Sci. Pollut. Res. 28, 4156–4172.
791 <https://doi.org/10.1007/s11356-020-10654-x>

792 Dallagnol Vargas, N., Guimarães, M., Caorsi, V., Wolff Bordignon, D., Borges-
793 Martins, M., 2020. An experimental assessment of the antipredatory function of
794 green dorsal coloration in poisonous Neotropical red-bellied toads. J. Zool. 310,
795 171–179. <https://doi.org/10.1111/jzo.12740>

796 Dole, J.W., 1971. Dispersal of Recently Metamorphosed Leopard Frogs, *Rana pipiens*.
797 Copeia 1971, 221. <https://doi.org/10.2307/1442821>

798 Duellman, W.E., Trueb, L., 1994. Biology of Amphibians, Reprint ed. ed. Johns
799 Hopkins University Press. <https://doi.org/10.2307/1445022>

800 Elgue, E., Pereira, G., Achaval-Coppes, F., Maneyro, R., 2014. Validity of photo-
801 identification technique to analyze natural markings in *Melanophryniscus*
802 *montevideensis* (Anura: Bufonidae). *Phyllomedusa* 13, 59–66.
803 <https://doi.org/10.11606/issn.2316-9079.v13i1p59-66>

804 Emmrich, M., Vences, M., Ernst, R., Köhler, J., Barej, M.F., Glaw, F., Jansen, M.,
805 Rödel, M.-O., 2020. A guild classification system proposed for anuran
806 advertisement calls. *Zoosystematics Evol.* 96, 515–525.
807 <https://doi.org/10.3897/zse.96.38770>

808 Fonte, L.F.M. da, Abadie, M., Mendes, T., Zank, C., Borges-Martins, M., 2014. The
809 Times they are a-Changing: How a MultiInstitutional Effort Stopped the
810 Construction of a Hydroelectric Power Plant that Threatened a Critically
811 Endangered Red-Belly Toad in Southern Brazil. *FrogLog* 22, 18–21.

812 Frost, D.R., 2021. Amphibian Species of the World: an Online Reference [WWW
813 Document]. Version 6.1. <https://doi.org/doi.org/10.5531/db.vz.0001>

814 Goldberg, F.J., Quinzio, S., Vaira, M., 2006. Oviposition-site selection by the toad
815 *Melanophryniscus rubriventris* in an unpredictable environment in Argentina 1871,
816 699–705. <https://doi.org/10.1139/Z06-038>

817 González-del-Pliego, P., Freckleton, R.P., Edwards, D.P., Koo, M.S., Scheffers, B.R.,
818 Pyron, R.A., Jetz, W., 2019. Phylogenetic and Trait-Based Prediction of Extinction
819 Risk for Data-Deficient Amphibians. *Curr. Biol.* 29, 1557-1563.e3.
820 <https://doi.org/10.1016/j.cub.2019.04.005>

821 Goodwin, T.W., 1984. The Biochemistry of the Carotenoids, 1st ed, The Biochemistry
822 of the Carotenoids - Volume II Animals. Springer Netherlands.
823 <https://doi.org/10.1007/978-94-009-5542-4>

824 Gosner, K.L., 1960. A simplified table for staging anuran embryos and larvae with
825 notes on identification. *Herpetologica* 16, 183–190.

826 Goutte, S., Dubois, A., Howard, S.D., Márquez, R., Rowley, J.J.L., Dehling, J.M.,
827 Grandcolas, P., Xiong, R.C., Legendre, F., 2018. How the environment shapes
828 animal signals: a test of the acoustic adaptation hypothesis in frogs. *J. Evol. Biol.*
829 31, 148–158. <https://doi.org/10.1111/jeb.13210>

830 Grant, T., Colombo, P., Verrastro, L., Saporito, R.A., 2012. The occurrence of
831 defensive alkaloids in non-integumentary tissues of the Brazilian red-belly toad
832 *Melanophryniscus simplex* (Bufonidae). *Chemoecology* 22, 169–178.
833 <https://doi.org/10.1007/s00049-012-0107-9>

834 Greenwood, P.J., 1980. Mating systems, philopatry and dispersal in birds and mammals.
835 *Anim. Behav.* 28, 1140–1162. [https://doi.org/10.1016/S0003-3472\(80\)80103-5](https://doi.org/10.1016/S0003-3472(80)80103-5)

836 Guimarães, T.C.S., De Figueiredo, G.B., Mesquita, D.O., Vasconcellos, M.M., 2011.
837 Ecology of *Hypsiboas albopunctatus* (Anura: Hylidae) in a Neotropical savanna. *J.*
838 *Herpetol.* 45, 244–250. <https://doi.org/10.1670/09-255.1>

839 Haad, B., Vera Candioti, F., Baldo, D., 2011. Shape variation in lentic and lotic tadpoles
840 of *Melanophryniscus* (Anura: Bufonidae). *Stud. Neotrop. Fauna Environ.* 46, 91–
841 99. <https://doi.org/10.1080/01650521.2011.593124>

842 Haddad, C. F., Prado, C. P., 2005. Reproductive modes in frogs and their unexpected
843 diversity in the Atlantic Forest of Brazil. *BioScience*, 55(3), 207-217.

844 Haddad, C.F.B., Machado, I.F., Giovanelli, J.G.R., Bataus, Y.S.L., Uhlig, V.M., Batista,
845 F.R.Q., Maciel, A.O.; Cruz, C.A.G. Loebmann, D., Silvano, D.L., Gonsales,
846 E.M.L., Nomura, F., Leite, F.S.F., Pinto, H.B.A., Amaral, I.B., Nascimento, L.B.,
847 Sturaro, M.J., Hoogmoed, M.S., Kienle, M.S. Souza, M.B., Maciel, N.M.,
848 Colombo, P., Feio, R.N., Lingnau, R., Bastos, R., 2018. *Melanophryniscus*
849 *admirabilis* Di-Bernardo, Grillo & Maneyro, 2006, in: Biodiversidade, I.C.M. de

850 C. da (Ed.), Livro Vermelho Da Fauna Brasileira Ameaçada de Extinção: Volume
851 V - Anfíbios. Brasília: ICMBio, pp. 30–32.

852 Halliday, T.R., Verrell, P.A., 1988. Body Size and Age in Amphibians and Reptiles. *J.*
853 *Herpetol.* 22(3), 253-265.

854 Hamilton, W.D., May, R.M., 1977. Dispersal in stable habitats. *Nature* 269, 578–581.
855 <https://doi.org/10.1038/269578a0>

856 Hantak, M.M., Grant, T., Reinsch, S., McGinnity, D., Loring, M., Toyooka, N.,
857 Saporito, R.A., 2013. Dietary Alkaloid Sequestration in a Poison Frog: An
858 Experimental Test of Alkaloid Uptake in *Melanophryniscus stelzneri* (Bufonidae).
859 *J. Chem. Ecol.* 39, 1400–1406. <https://doi.org/10.1007/s10886-013-0361-5>

860 Hayes, T., Chan, R., Licht, P., 1993. Interactions of temperature and steroids on larval
861 growth, development, and metamorphosis in a toad (*Bufo boreas*). *J. Exp. Zool.*
862 266, 206–215. <https://doi.org/10.1002/jez.1402660306>

863 Hoffman, E. a, Blouin, M.S., 2000. A review of colour and pattern polymorphisms in
864 anurans. *Biol. J. Linn. Soc.* 70, 633–665. <https://doi.org/10.1006/bijl.1999.0421>

865 Hourdry, J., Beaumont, A., 1985. Les métamorphoses des amphibiens. Elsevier
866 Masson. Paris: Singer Polignac, Masson.

867 Howard, S.D., Bickford, D.P., 2014. Amphibians over the edge: Silent extinction risk of
868 Data Deficient species. *Divers. Distrib.* 20, 837–846.
869 <https://doi.org/10.1111/ddi.12218>

870 Instrução Normativa MMA nº 3, de 27 de maio de 2003, 2003. Lista das Espécies da
871 Fauna Brasileira Ameaçadas de Extinção. Ministério do Meio Ambiente, Brasil.

872 IUCN, 2021. The IUCN Red List of Threatened Species [WWW Document]. Version
873 2021-1. URL <https://www.iucnredlist.org>

874 IUCN SSC Amphibian Specialist Group, 2013. *Melanophryniscus admirabilis* [WWW
875 Document]. IUCN Red List Threat. Species 2013 e.T135993A44846478. URL
876 <https://dx.doi.org/10.2305/IUCN.UK.2013-2.RLTS.T135993A44846478.en>.
877 (accessed 3.25.21).

878 Jeckel, A.M., Caorsi, V.Z., Grant, T., Borges-Martins, M., 2019. The Nuptial Pads of
879 *Melanophryniscus* (Anura: Bufonidae), with the Unexpected Occurrence of
880 Nuptial-Pad-Like Structures in Females of Two Species. *J. Herpetol.* 53, 53.
881 <https://doi.org/10.1670/18-104>

882 Jeckel, A.M., Saporito, R. A., Grant, T., 2015. The relationship between poison frog
883 chemical defenses and age, body size, and sex. *Front. Zool.* 12, 27.
884 <https://doi.org/10.1186/s12983-015-0120-2>

885 Kwet, A., Maneyro, R., Zillikens, A., Mebs, D., 2005. Advertisement calls of
886 *Melanophryniscus dorsalis* (Mertens, 1933) and *M. montevidensis* (Philippi, 1902),
887 two parapatric species from southern Brazil and Uruguay, with comments on
888 morphological variation in the *Melanophryniscus stelzneri* group (Anura:
889 Bufonidae). *Salamandra* 41(1/2), 3–20.

890 Köhler, J., Jansen, M., Rodríguez, A., Kok, P.J.R., Toledo, L.F., Emmrich, M., Glaw,
891 F., Haddad, C.F.B., Rödel, M.O., Vences, M., 2017. The use of bioacoustics in
892 anuran taxonomy: Theory, terminology, methods and recommendations for best
893 practice. *Zootaxa*. <https://doi.org/10.11646/zootaxa.4251.1.1>

894 Kuramoto, M., 1978. Correlations of Quantitative Parameters of Fecundity in
895 Amphibians. *Evolution* (N. Y). 32, 287. <https://doi.org/10.2307/2407596>

896 La Marca, E., 1984. Longevity in the Venezuelan Yellow Frog *Atelopus oxyrhynchus*
897 *carbonerensis* (Anura: Bufonidae). *Trans. Kansas Acad. Sci.* 87, 66–67.
898 <https://doi.org/10.2307/3627767>

899 Lavilla, E.O., Scrocchi, G.J., 1986. Morfometría larval de los géneros de Telmatobiinae

900 (Anura: Leptodactylidae) de Argentina y Chile. *Physis*, 44, 39–43.
 901 Lavilla, E., Vaira, M., 1997. La larva de *Melanophryniscus rubriventris rubriventris*
 902 (Vellard, 1947) (Anura, Bufonidae). *Alytes* 15, 19–25.
 903 Lyapkov, S.M., Kidov, A.A., Stepankova, I. V., Afrin, K.A., Litvinchuk, S.N., 2020.
 904 Age structure and growth in the lataste's toad, *Bufotes latastii* (Anura: Bufonidae).
 905 *Russ. J. Herpetol.* 27, 165–171. <https://doi.org/10.30906/1026-2296-2020-27-3-165-171>
 906
 907 Maneyro, R., Carreira, S., 2012. Guía de Anfibios del Uruguay. Ediciones de la Fuga.
 908 Montevideo. 1 – 207 pp.
 909 Marques, A.A.B., Fontana, C.S., Vélez, E., Bencke, G.A., Schneider, M., Reis, R.E.,
 910 2002. Lista de Referência da Fauna Ameaçada de Extinção no Rio Grande do Sul.
 911 Decreto no 41.672, de 11 junho de 2002. Publicações Avulsas FZB, 11, Porto
 912 Alegre: FZB/MCT-PUCRS/PANGEA.
 913 Minella, J.P.G., Merten, G.H., Walling, D.E., Reichert, J.M., 2009. Changing sediment
 914 yield as an indicator of improved soil management practices in southern Brazil.
 915 *Catena* 79, 228–236. <https://doi.org/10.1016/j.catena.2009.02.020>
 916 Monnet, J.-M., Cherry, M.I., 2002. Sexual size dimorphism in anurans. *Proc. R. Soc.*
 917 *London. Ser. B Biol. Sci.* 269, 2301–2307. <https://doi.org/10.1098/rspb.2002.2170>
 918 Moreno, J.A., 1961. Clima do Rio Grande do Sul. Secretaria Da Agricultura Do Estado
 919 Do Rio Grande Do Sul. Diretoria de Terras e Colonização, Seção de Geografia,
 920 Porto Alegre. <https://revistas.dee.spgg.rs.gov.br/index.php/boletim-geografico-rs/article/view/3236/3310>
 921
 922 Myers, N., Mittermeyer, R.A., Mittermeyer, C.G., Da Fonseca, G.A.B., Kent, J., 2000.
 923 Biodiversity hotspots for conservation priorities. *Nature* 403, 853–858.
 924 <https://doi.org/10.1038/35002501>
 925 Nali, R.C., Zamudio, K.R., Haddad, C.F.B., Prado, C.P.A., 2014. Size-dependent
 926 selective mechanisms on males and females and the evolution of sexual size
 927 dimorphism in frogs. *Am. Nat.* 184, 727–740. <https://doi.org/10.1086/678455>
 928 Penna, M., 2004. Amplification and spectral shifts of vocalizations inside burrows of
 929 the frog *Eupsophus calcaratus* (Leptodactylidae). *J. Acoust. Soc. Am.* 116, 1254–
 930 1260. <https://doi.org/10.1121/1.1768257>
 931 Penna, M., Hamilton-West, C., 2007. Susceptibility of evoked vocal responses to noise
 932 exposure in a frog of the temperate austral forest. *Anim. Behav.* 74, 45–56.
 933 <https://doi.org/10.1016/j.anbehav.2006.11.010>
 934 Pereira, G., Maneyro, R., 2018. Reproductive biology of *Melanophryniscus*
 935 *montevidensis* (Anura: Bufonidae) from Uruguay: reproductive effort, fecundity,
 936 sex ratio and sexual size dimorphism. *Stud. Neotrop. Fauna Environ.* 53, 10–21.
 937 <https://doi.org/10.1080/01650521.2017.1364952>
 938 Pereira, G., Maneyro, R., 2016. Movement Patterns in a Uruguayan Population of
 939 *Melanophryniscus montevidensis* (Philippi, 1902) (Anura: Bufonidae) Using
 940 Photo-Identification for Individual Recognition. *South Am. J. Herpetol.* 11, 119–
 941 126. <https://doi.org/10.2994/SAJH-D-15-00020.1>
 942 Pereyra, L.C., Lescano, J.N., Leynaud, G.C., 2011. Breeding-site selection by red-belly
 943 toads, *Melanophryniscus stelzneri* (Anura: Bufonidae), in Sierras of Córdoba,
 944 Argentina. *Amphib. Reptil.* 32, 105–112.
 945 <https://doi.org/10.1163/017353710X543029>
 946 Perrin, N., Mazalov, V., 2000. Local competition, inbreeding, and the evolution of sex-
 947 biased dispersal. *Am. Nat.* 155, 116–127. <https://doi.org/10.1086/303296>
 948 Portaria MMA N° 444, de 17 de dezembro de 2014, 2014. Lista Nacional Oficial de
 949 Espécies da Fauna Ameaçadas de Extinção. Ministério do Meio Ambiente, Brasil.

- 950 Rio Grande do Sul, G. do, 2014. Espécies da Fauna Silvestre Ameaçadas de Extinção
951 no Estado do Rio Grande do Sul. Decreto 51.797, de 08 de setembro de 2014.
952 Estado do Rio Grande do Sul, Brazil.
- 953 Röhr, D.L., Paterno, G.B., Camurugi, F., Juncá, F.A., Garda, A.A., 2016. Background
954 noise as a selective pressure: stream-breeding anurans call at higher frequencies.
955 Org. Divers. Evol. 16, 269–273. <https://doi.org/10.1007/s13127-015-0256-0>
- 956 Santos, R.R., Grant, T., 2010. Diel pattern of migration in a poisonous toad from Brazil
957 and the evolution of chemical defenses in diurnal amphibians. Evol. Ecol.
958 <https://doi.org/10.1007/s10682-010-9407-0>
- 959 Santos, R.R., Leonardi, S.B., Caorsi, V.Z., Grant, T., 2010. Directional orientation of
960 migration in an aseasonal explosive-breeding toad from Brazil. J. Trop. Ecol. 26,
961 415–421. <https://doi.org/10.1017/S0266467410000180>
- 962 Saporito, R.A., Donnelly, M.A., Spande, T.F., Garraffo, H.M., 2012. A review of
963 chemical ecology in poison frogs. Chemoecology. [https://doi.org/10.1007/s00049-](https://doi.org/10.1007/s00049-011-0088-0)
964 011-0088-0
- 965 Saporito, R.A., Spande, T.F., Garraffo, H.M., Donnelly, M.A., 2009. Arthropod
966 alkaloids in poison frogs: A review of the “dietary hypothesis.” Heterocycles 79,
967 277–297. [https://doi.org/10.3987/REV-08-SR\(D\)11](https://doi.org/10.3987/REV-08-SR(D)11)
- 968 Scott Jr., N.J., Woodward, B.D., 1994. Standard techniques for inventory and
969 monitoring: surveys at breeding sites., in: Heyer, W.R., Donnelly, M.A.,
970 McDiarmid, R.W., Hayek, L.C., Foster, M.S. (Eds.), Measuring and Monitoring
971 Biological Diversity, Standard Methods for Amphibians. Smithsonian Institution
972 Press, Washington DC, pp. 92–96.
- 973 Shine, R., 1979. Sexual Selection and Sexual Dimorphism in the Amphibia. Copeia
974 1979, 297. <https://doi.org/10.2307/1443418>
- 975 Silva, M.A., Magalhães, S., Prieto, R., Santos, R.S., Hammond, P.S., 2009. Estimating
976 survival and abundance in a bottlenose dolphin population taking into account
977 transience and temporary emigration. Mar. Ecol. Prog. Ser. 392, 263–276.
978 <https://doi.org/10.3354/meps08233>
- 979 Smith-Gill, S.J., Berven, K.A., 1979. Predicting amphibian metamorphosis. Am. Nat.
980 113, 563–585. <https://doi.org/10.1086/283413>
- 981 Spieler, M., Linsenmair, K.E., 1997. Choice of optimal oviposition sites by
982 *Hoplobatrachus occipitalis* (Anura: Ranidae) in an unpredictable and patchy
983 environment. Oecologia 109, 184–199. <https://doi.org/10.1007/s004420050073>
- 984 Stark, G., Meiri, S., 2018. Cold and dark captivity: Drivers of amphibian longevity.
985 Glob. Ecol. Biogeogr. 27, 1384–1397. <https://doi.org/10.1111/geb.12804>
- 986 Stephen Dobson, F., 1982. Competition for mates and predominant juvenile male
987 dispersal in mammals. Anim. Behav. 30, 1183–1192.
988 [https://doi.org/10.1016/S0003-3472\(82\)80209-1](https://doi.org/10.1016/S0003-3472(82)80209-1)
- 989 Umbers, K.D.L., Silla, A.J., Bailey, J.A., Shaw, A.K., Byrne, P.G., 2016. Dietary
990 carotenoids change the colour of Southern corroboree frogs. Biol. J. Linn. Soc.
991 119, 436–444. <https://doi.org/10.1111/bij.12818>
- 992 Vaira, M., 2005. Annual variation of breeding patterns of the toad, *Melanophryniscus*
993 *rubriventris* (Vellard, 1947). Amphibia-Reptilia 26, 193–199.
994 <https://doi.org/10.1163/1568538054253519>
- 995 Van Sluys, M., Guido-Castro, P., 2011. Influence of Temperature and Photoperiod on
996 the Activity of *Melanophryniscus moreirae* (Miranda-Ribeiro 1920) (Anura:
997 Bufonidae) on the Itatiaia Plateau, Southeastern Brazil. South Am. J. Herpetol. 6,
998 43–48. <https://doi.org/10.2994/057.006.0106>
- 999 Vargas-Salinas, F., Amézquita, A., 2014. Abiotic noise, call frequency and stream-

1000 breeding anuran assemblages. *Evol. Ecol.* 28, 341–359.
1001 <https://doi.org/10.1007/s10682-013-9675-6>
1002 Wells, K.D., 2010. *The Ecology and Behavior of Amphibians*. University of Chicago
1003 Press, Chicago.
1004 Wells, K.D., 1977. The social behaviour of anuran amphibians. *Anim. Behav.* 25, 666–
1005 693.
1006 Zank, C., Becker, F.G., Abadie, M., Baldo, D., Maneyro, R., Borges-Martins, M., 2014.
1007 Climate change and the distribution of neotropical red-bellied toads
1008 (*Melanophryniscus*, Anura, Amphibia): How to prioritize species and populations?
1009 *PLoS One* 9, 1–11. <https://doi.org/10.1371/journal.pone.0094625>
1010 Zepner, L., Karrasch, P., Wiemann, F., Bernard, L., 2020. ClimateCharts.net—an
1011 interactive climate analysis web platform. *Int. J. Digit. Earth.*
1012 <https://doi.org/10.1080/17538947.2020.1829112>
1013

1014 **Appendix A - Supplementary Information**

1015

1016 **Morphology, reproduction, longevity, and bioacoustics of the microendemic and**

1017

threatened Admirable Redbelly Toad

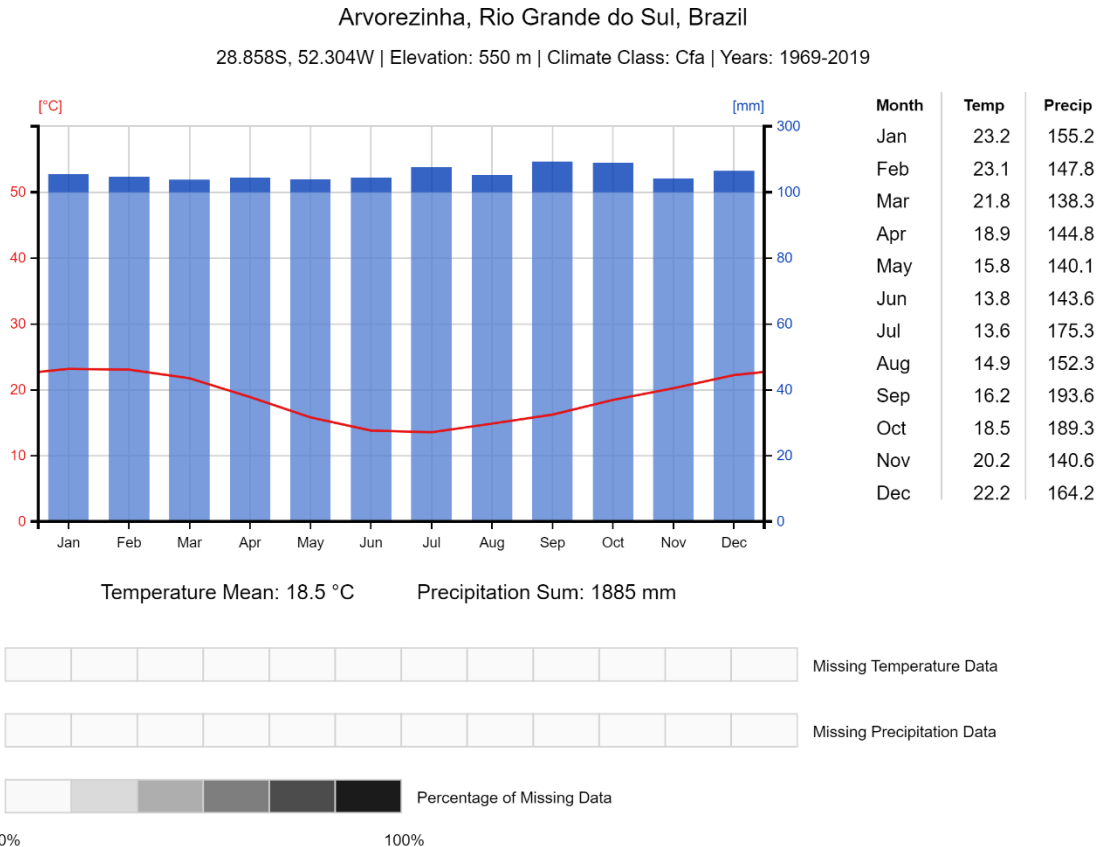
1018 *Michelle Abadie, Luis Fernando Marin da Fonte, Débora Bordignon, Thayná Mendes,*

1019 *Caroline Zank, Valentina Zaffaroni, Andreas Kindel, Raúl Maneyro, Márcio Borges-*

1020 *Martins.*

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1024 **Supplementary Figure S1.** Subtropical Humid Climate, variety *Cfa* (Koeppen’s climate

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classification) in the region of Perau de Janeiro, Arvorezinha, Rio Grande do Sul state,

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Brazil. The type *Cfa* climate is characterised by precipitation evenly distributed

1027

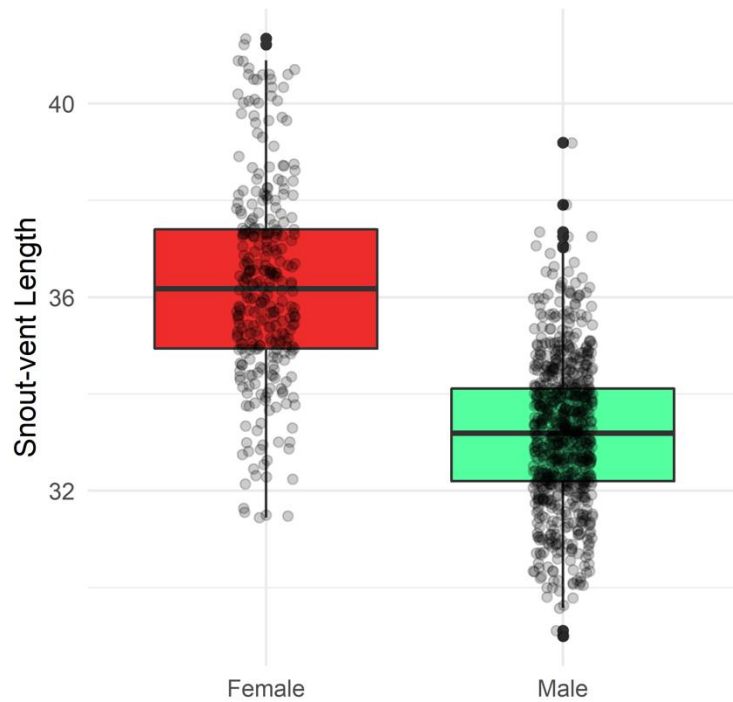
throughout the year, with hot summers (average temperatures above 22 °C for the hottest

1028

month) and cold winters (average temperatures varying between -3 and 18 °C for the

1029

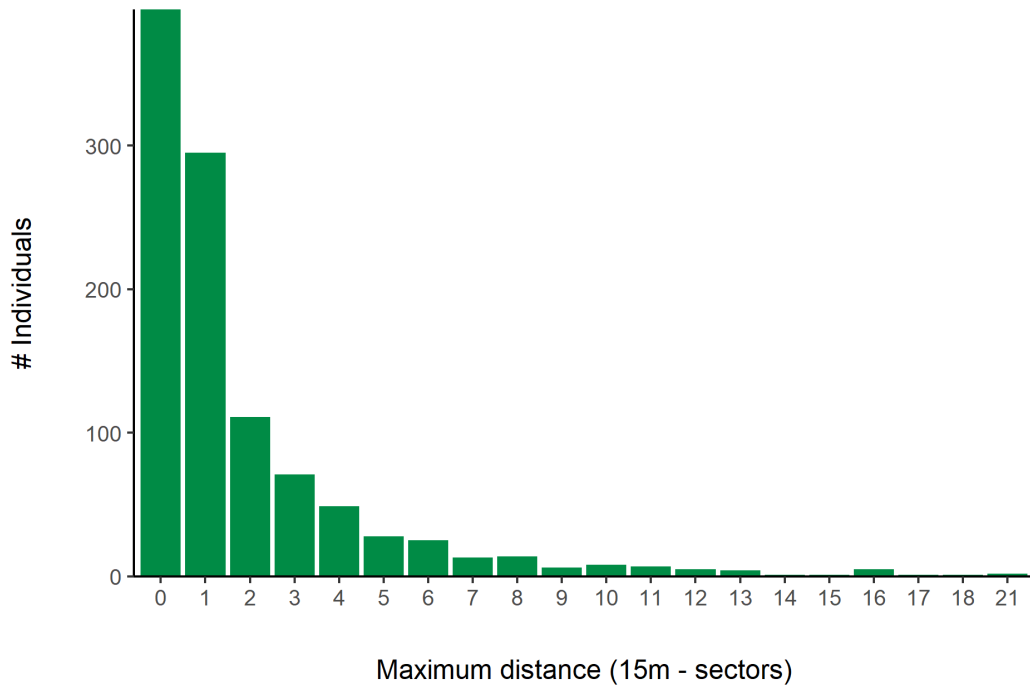
coldest month). Diagram generated in <https://climatecharts.net/>.



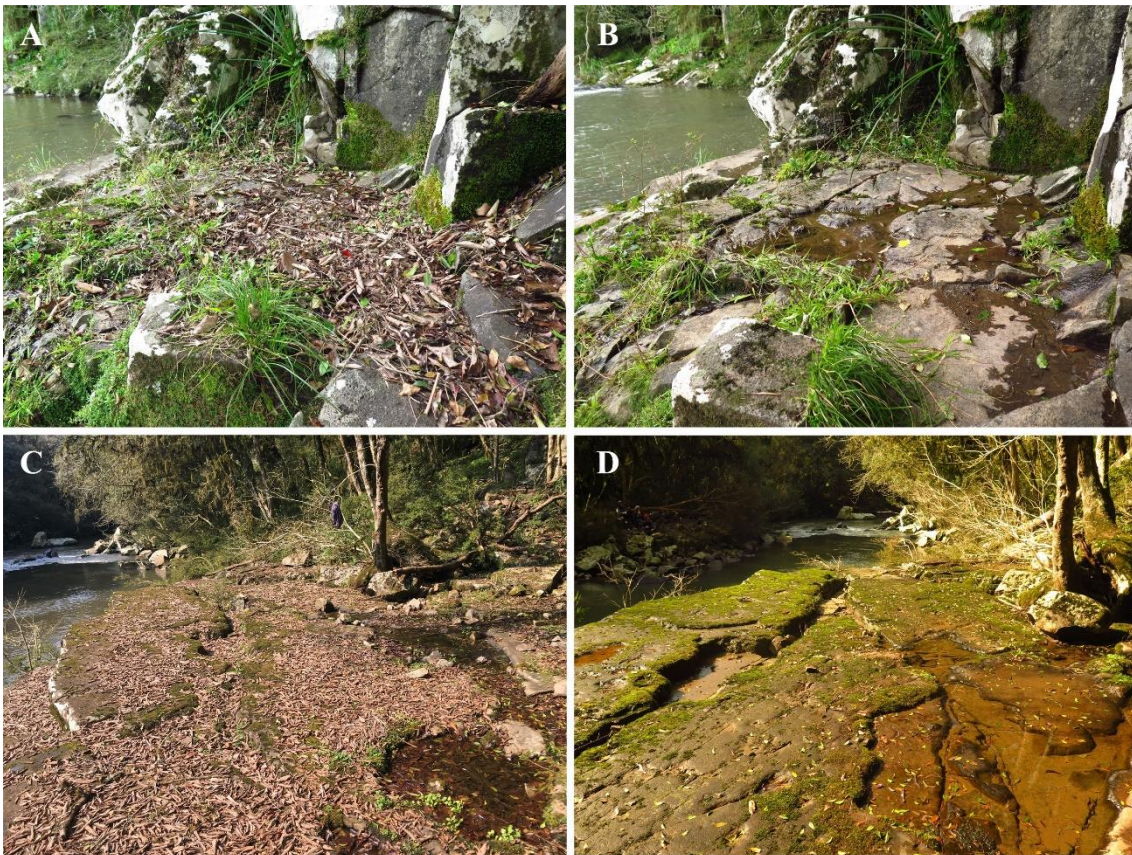
1030
1031 **Supplementary Figure S2.** Size sexual dimorphism on *Melanophryniscus admirabilis*.
1032 Boxplots showing the distribution of the snout-vent-length (mm) of 302 females and 760
1033 males (point cloud). The rectangular boxes contain the central 50% of the values, the
1034 heavy line in the centre of each box is the median, the whiskers extend to 1.5 times the
1035 interquartile range above and below the box and outliers are indicated by black filled
1036 points.
1037



1038
 1039 **Supplementary Figure S3.** When *Melanophryniscus admirabilis* is not breeding. (A-B)
 1040 two adult individuals in the adjacent forested steep slopes to the rocky river bank outcrop;
 1041 (C-D) some individuals hiding in spaces under rocks; (E-F) some individuals hiding
 1042 under vegetation and in a rocky slit, respectively, in moments when they are not breeding
 1043 but they are at the breeding site. Photos: (F) by Simone Leonardi.



1044
 1045 **Supplementary Figure S4.** Frequency of individuals for maximum distance between
 1046 sectors registered. Each sector has 15 metres of length.
 1047



1048
 1049 **Supplementary Figure S5.** Availability of water in the small pools at the breeding site
 1050 of *Melanophryniscus admirabilis*, in the type locality. (A and C) Pools filled by dry leaves

1051 at the end of austral winter; (B and D) the same places after raining and “washing” the
 1052 rocky river bank outcrop. Picture (B) was taken ten days after the picture (A).

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 1054

1055 **Supplementary Video S6.** Spawning behaviour of *Melanophryniscus admirabilis*. Video
 1056 available [clicking here](#).

1057
 1058

1059 **Supplementary Table S7.** Advertisement call characteristics of *Melanophryniscus*
 1060 *admirabilis*, based on nine calls from four individuals in the type locality Perau de
 1061 Janeiro, Arvorezinha, RS, Brasil. Call recordings were carried out under air temperatures
 1062 ranging between 17.5 and 23.9 °C.

#	Acoustic parameter	Average ± SD ^a (range)
1	Call duration (s)	21.00 ± 1.50 (2.90 – 68.80)
2	Call part A duration (s)	15.40 ± 19.10 (0.32 – 64.30)
3	Number of notes Call part A	8.20 ± 4.90 (2.00 – 19.00)
4	Duration of notes Call part A (s)	0.10 ± 0.04 (0.01 – 0.17)
5	Interval among notes Call part A (s)	2.00 ± 2.40 (0.01 – 18.80)
6	Dominant frequency Call part A (Hz)	1939 ± 383 (1500 – 2800)
7	Interval between call parts A and B (s)	1.50 ± 4.10 (0.06 – 12.60)
8	Call part B duration (s)	4.10 ± 1.70 (1.00 – 6.26)
9	Number of pulses Call part B	206.30 ± 88.60 (53 – 324)
10	Pulses rate Call part B	50.80 ± 2.00 (49.00 – 54.30)
11	Duration of pulses Call part B (s)	0.006 ± 0.002 (0.002 – 0.020)
12	Interval among pulses Call part B (s)	0.014 ± 0.002 (0.002 – 0.030)
13	Dominant frequency Call part B (Hz)	1749 ± 219.50 (1500 – 1968)

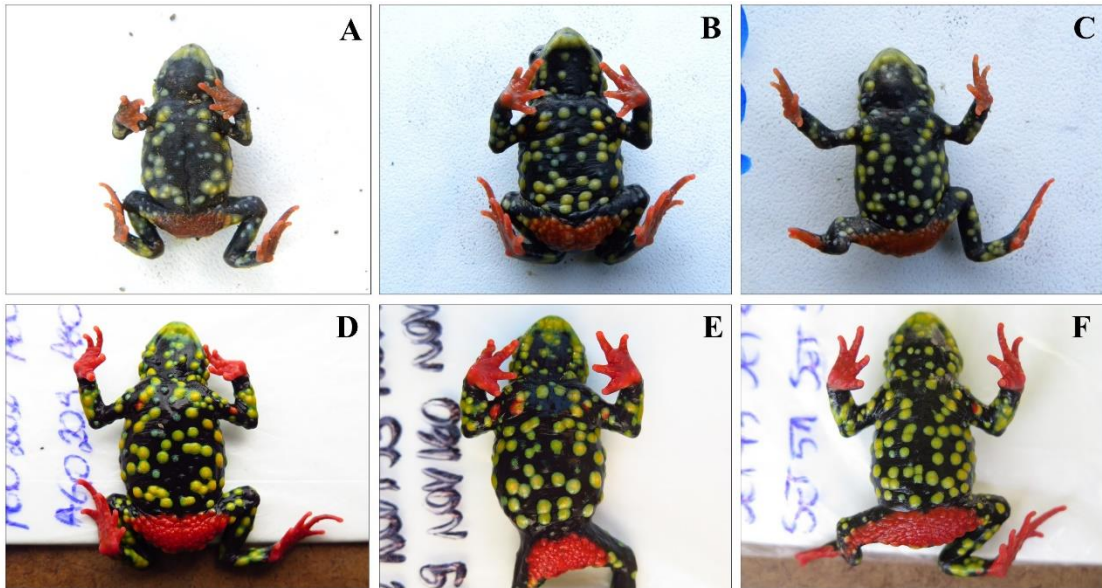
^aSD = standard deviation

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1067 **Supplementary Table S8.** Body and oral disc measurements were taken from tadpoles
 1068 of *M. admirabilis* (n=20) by development stage (*sensu* Gosner, 1960). Mean, standard
 1069 deviation and range values are shown in millimetres.

Measurement	32 (n=5)	34 (n=2)	35 (n=2)	36 (n=5)	37 (n=6)
TL	14.68 ± 0.55 (13.9 – 15.3)	14.85 ± 0.92 (14.2 – 15.5)	15.55 ± 1.06 (14.8 – 16.3)	16.52 ± 0.94 (15.4 – 17.6)	16.22 ± 0.76 (15.2 – 17.1)
BL	5.82 ± 0.31 (5.3 – 6.1)	6.10 ± 0.57 (5.7 – 6.5)	6.35 ± 0.35 (6.1 – 6.6)	6.62 ± 0.41 (6.1 – 7.2)	6.57 ± 0.29 (6.1 – 6.9)
TAL	9.08 ± 0.51 (8.5 – 9.7)	8.65 ± 0.35 (8.4 – 8.9)	9.2 ± 0.71 (8.7 – 9.7)	9.9 ± 0.60 (9.3 – 10.5)	9.63 ± 0.49 (9.0 – 10.2)
BMW	4.26 ± 0.11 (4.1 – 4.4)	4.50 ± 0.42 (4.2 – 4.8)	4.70 ± 0.00 (4.7)	4.66 ± 0.36 (4.1 – 5.1)	4.70 ± 0.25 (4.3 – 4.9)
BWN	2.80 ± 0.16 (2.6 – 3.0)	2.95 ± 0.07 (2.9 – 3.0)	2.90 ± 0.14 (2.8 – 3.0)	3.04 ± 0.15 (2.8 – 3.2)	3.02 ± 0.16 (2.8 – 3.3)
BWE	3.70 ± 0.12 (3.5 – 3.8)	3.90 ± 0.28 (3.7 – 4.1)	3.90 ± 0.14 (3.8 – 4.0)	4.04 ± 0.18 (3.8 – 4.2)	4.03 ± 0.12 (3.9 – 4.2)
BMH	3.48 ± 0.16 (3.3 – 3.7)	4.20 ± 0.42 (3.9 – 4.5)	4.00 ± 0.00 (4.0)	3.90 ± 0.16 (3.7 – 4.1)	4.00 ± 0.18 (3.8 – 4.3)
MTH	2.98 ± 0.19 (2.7 – 3.2)	2.90 ± 0.14 (2.8 – 3.0)	3.15 ± 0.21 (3.0 – 3.3)	3.38 ± 0.22 (3.2 – 3.7)	3.27 ± 0.26 (2.8 – 3.5)
TMH	1.14 ± 0.05 (1.1 – 1.2)	1.15 ± 0.07 (1.1 – 1.2)	1.20 ± 0.00 (1.2)	1.30 ± 0.10 (1.2 – 1.4)	1.32 ± 0.04 (1.3 – 1.4)
TMW	1.04 ± 0.11 (0.9 – 1.2)	1.15 ± 0.07 (1.1 – 1.2)	1.20 ± 0.00 (1.2)	1.28 ± 0.08 (1.2 – 1.4)	1.28 ± 0.13 (1.1 – 1.4)
RSD	3.72 ± 0.22 (3.4 – 4.0)	3.80 ± 0.0 (3.8)	4.10 ± 0.28 (3.9 – 4.3)	4.12 ± 0.23 (3.9 – 4.4)	4.05 ± 0.16 (3.8 – 4.3)
FN	0.76 ± 0.09 (0.7 – 0.9)	0.85 ± 0.07 (0.8 – 0.9)	0.90 ± 0.14 (0.8 – 1.0)	0.94 ± 0.13 (0.8 – 1.1)	0.83 ± 0.08 (0.7 – 0.9)
IND	0.95 ± 0.05 (0.9 – 1.0)	1.00 ± 0.0 (1.0)	1.05 ± 0.07 (1.0 – 1.1)	1.04 ± 0.05 (1.0 – 1.1)	1.05 ± 0.05 (1.0 – 1.1)
E	0.66 ± 0.07 (0.6 – 0.7)	0.67 ± 0.04 (0.7 – 0.7)	0.75 ± 0.07 (0.7 – 0.8)	0.80 ± 0.07 (0.7 – 0.9)	0.83 ± 0.10 (0.7 – 1.0)
IOD	1.75 ± 0.05 (1.7 – 1.8)	1.85 ± 0.07 (1.8 – 1.9)	1.90 ± 0.00 (1.9)	2.00 ± 0.10 (1.9 – 2.1)	2.00 ± 0.13 (1.8 – 2.1)
OD	2.02 ± 0.04 (2.0 – 2.1)	2.20 ± 0.28 (2.0 – 2.4)	2.35 ± 0.07 (2.3 – 2.4)	2.28 ± 0.28 (2.0 – 2.7)	2.27 ± 0.14 (2.1 – 2.4)
DG	1.30 ± 0.11 (2.3 – 2.8)	1.38 ± 0.04 (1.4 – 1.4)	1.60 ± 0.14 (1.5 – 1.7)	1.52 ± 0.15 (1.3 – 1.7)	1.53 ± 0.10 (1.4 – 1.7)
VG	0.98 ± 0.09 (1.7 – 2.2)	1.00 ± 0.21 (0.9 – 1.15)	1.05 ± 0.07 (1.0 – 1.1)	1.21 ± 0.19 (1.1 – 1.5)	1.14 ± 0.09 (1.0 – 1.3)

1070 TL= total length; BL= body length; TAL= tail length; BMW= body maximum width; BWN= body width
 1071 at nostrils; BWE= body width at eyes; BMH= body maximum height; MTH= maximum tail height;
 1072 TMH= tail musculature height; TMW= tail musculature width; RSD= rostrum-spiracular distance; FN=
 1073 fronto-nasal distance; IND= internarial distance; E= eye diameter; IOD= interorbital distance.
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1076
 1077 **Supplementary Figure S9.** Ontogenetic variation of the ventral colour pattern of three
 1078 individuals of *M. admirabilis*. Figures A-C are juveniles and D-F are their respective
 1079 adults. (A) Female juvenile (SVL= 15.52 mm) captured on October 2013 at sector #30
 1080 and (D) recaptured as adult (SVL = 35.04 mm) on August 2017 at sector #28. (B) Male
 1081 juvenile (SVL= 21.82 mm) captured on October 2013 at sector #22 and (E) recaptured as
 1082 adult (SVL = 32.86 mm) on November 2017 at sector #15. (C) Male juvenile (SVL=
 1083 15.88 mm) captured on October 2013 at sector #22 and (F) recaptured as adult (SVL =
 1084 31.23 mm) on September 2016 at sector #18.

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Capítulo 2

The abundance of the microendemic Admirable Redbelly Toad: a comparison of population size estimates.

Este manuscrito segue o formato da revista *Journal of Animal Ecology*. A fim de facilitar a leitura, as figuras e tabelas foram inseridas no corpo do texto.

The abundance of the microendemic Admirable Redbelly Toad: a comparison of population size estimates.

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1 Abstract

1. Limited range and habitat specificity make microendemic species particularly vulnerable to environmental change. Their persistence may be a result of random luck or of demographic processes that confer resistance to change. Population size and variability are key elements for understanding the demography of small populations as well as their chances of persistence. Among Neotropical anurans, the genus *Melanophryniscus* stands out as having a large number of species with small, microendemic populations that may be threatened.
2. The focus of this study is the Admirable Redbelly Toad (*Melanophryniscus admirabilis*), a Critically Endangered and microendemic anuran. We aim to estimate the size of the species population, as it varies through time in an eight-year window of observation.
3. We employ capture-recapture models to estimate population size, using individual recognition by photo-identification in a data set with more than seventeen hundred capture histories. We used three analytical approaches, the Closed population

16 (Closed) and the Jolly-Seber POPAN approaches which model data one year at a time
17 and Pollock's robust design (PRD), which simultaneously models data from the whole
18 observation period and provides annual estimates of abundance and apparent survival
19 probability.

20 4. Our estimates of population size varied more between methods in the same year
21 than between years, ranging from 561 ± 39 (estimate \pm standard error) to $1,734 \pm 361$
22 individuals over the whole study period. There is a substantial decrease in the estimated
23 number of individuals in the middle of the study period. POPAN and PRD abundance
24 estimates were closer to each other than either was to the Closed model estimates.

25 5. We conclude that the only known population has fewer than 2000 reproductive
26 individuals. Movements, within or between seasons, are likely the most important factor
27 behind temporal variation in the number of reproductive Admirable Toads. When multi-
28 year analyses are not possible, our results favor analyses that account for movement in
29 and out of the study area throughout one breeding season. Otherwise, the PRD offers
30 the most efficient approach for extracting information from the multi-year capture
31 histories.

32

33 **Key-words:** Capture-mark-recapture, Closed Population model, Jolly-Seber POPAN,
34 Pollock's Robust Design, *Melanophryniscus admirabilis*, microendemic, survival
35 estimates, threatened species.

36

37 **Introduction**

38 If extinction cannot happen without population decline, every population must
39 be rare at least once throughout its existence. Whether rarity is demographically
40 informative, however, is mostly a matter of time. Some populations may be rare for

41 only a moment before a cataclysmic event brings them to an end, while others may
42 remain rare—and observable—for extended periods. Rarity is a slippery concept with
43 multiple definitions (Rabinowitz 1981), but it always involves a small (or relatively
44 small) population size. The number of individuals in the population and the variability
45 of this number through time are key for interpreting the demography of rare populations
46 as well as their chances of persistence (Inchausti & Halley, 2003; Pertoldi et al., 2008).

47 Among rare species, persistent microendemics are the most intriguing, because
48 their limited range and often high habitat specificity make them vulnerable to
49 environmental change. Yet, their observed persistence may be a result of random luck
50 or of demographic processes that make them particularly resistant to change.

51 Neotropical anurans have particularly high endemism rates (Pimm et al., 2014;
52 Villalobos et al., 2013), and among them *Melanophryniscus* stands out as one of the
53 genera with more endemic species of strong conservation concern (Zank et al., 2014).
54 The focus of this study is the Admirable Redbelly Toad (*Melanophryniscus admirabilis*,
55 Anura: Bufonidae; Figure S1), a Critically Endangered and microendemic anuran
56 (IUCN SSC Amphibian Specialist Group, 2013) known from only one population,
57 which occurs over a stretch of a few hundred meters along the margins of one stream in
58 the southern end of the Brazilian Atlantic Forest (Di-Bernardo et al., 2006; Fonte et al.,
59 2014). Admirable Toads are small (about 35 mm in length) and occur in a very specific
60 environment consisting of forested river margins with flattened rocky outcrops along
61 the water's edge. They are difficult to find when they are not breeding, but occasionally
62 become conspicuous when hundreds of individuals aggregate for a few days at a time
63 on rocky outcrop pools where adults mate, clutches are laid, and eggs develop. Such
64 explosive breeding events (*sensu* Wells, 1977) take place during the second half of the
65 year (July – December: breeding season), coinciding with late winter, spring, and early

66 summer in the southern hemisphere (Abadie et al., 2021 – Chapter 1). This paper aims
67 to estimate the abundance of Admirable Redbelly Toad, a species whose individuals are
68 rarely and irregularly available for observation.

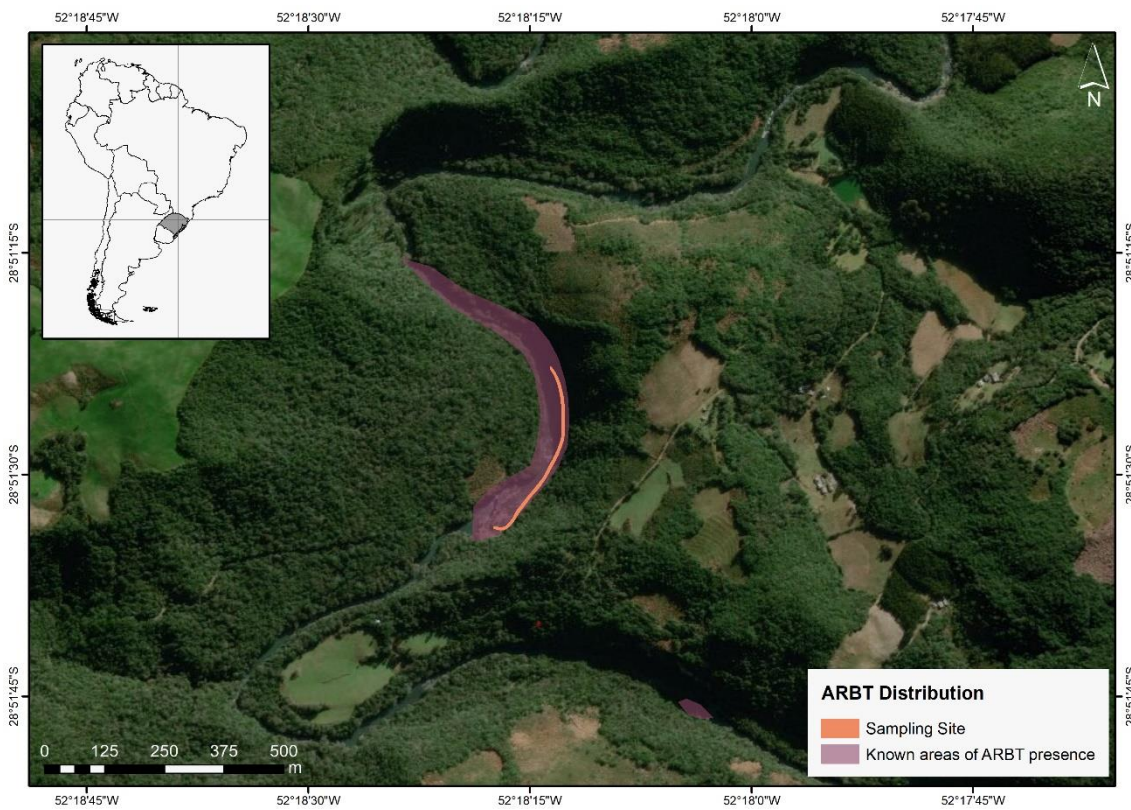
69 As we accumulated observations of individually recognizable toads over eight
70 years of study, we built more than seventeen hundred capture histories that we analyze
71 with a combination of three different statistical approaches that make different
72 assumptions about the data and Admirable Toad biology. We aim to inform
73 herpetologists about the advantages and disadvantages of each analytical approach and,
74 above all, to offer an assessment of *M. admirabilis* abundance, its variation through
75 time, and the demographic processes operating behind such variation.

76 **Materials and Methods**

77 *Study area*

78 We conducted our study at Perau de Janeiro, the Admirable Redbelly Toad type
79 locality, at the extreme south of the Atlantic Forest biome (Figure 1). Located in the
80 Brazilian state of Rio Grande do Sul, Arvorezinha municipality, Perau de Janeiro is part
81 of an approximately 5 km² patch of forest, which is one of the largest and few forest
82 fragments of the region. Regional forest cover is under pressure from the expansion of
83 eucalyptus, tobacco, soybean and livestock farming. Our study area extends ca. 400 m
84 along the margins of Forqueta River, which cuts through the forest patch at an elevation
85 of approximately 550 m above sea level and is part of the Taquari-Antas River basin.
86 The local humid subtropical climate has hot muggy summers and mild to cold winters,
87 with precipitation evenly distributed throughout the year (Alvares et al., 2013; Zepner et
88 al., 2020). The site is more humid than the surrounding area due to its forest cover and
89 steep slopes on both sides of the river (Figure S2). Our sampling area encompassed a

90 flattened rocky outcrop along the left bank of the river (Figure 1; Figure S3), where
91 most of the known Admirable Redbelly Toad population concentrates when breeding.
92 Admirable Toads find shelter in the forest and often spawn in hundreds of small
93 ephemeral pools on the rocky margins (Figure S3). Successful reproduction potentially
94 depends on the duration of the pools, which can be shortened by river flooding or by
95 high temperatures that lead to evaporation (Abadie et al., 2021 – Chapter 1).



96 **Figure1.** Known geographic distribution of Admirable Redbelly Toad at Perau de
97 Janeiro, municipality of Arvorezinha, Rio Grande do Sul state, Brazil. The sampling site
98 for this study is at the main breeding site of Admirable Redbelly Toad (ARBT), on the
99 flattened rocky outcrop along the left bank of the Forqueta River.
100
101

102 *Data collection*

103 We divided the sampling area in 27, 15-m-long sectors, and captured toads in
104 fifteen sectors per day of fieldwork. Nine of these fifteen sectors were fixed, because

105 they consistently held more animals than the others, while the remaining six were
 106 randomly selected each day. We visited the sectors while moving in opposite directions
 107 along the margin on consecutive days, to avoid always going through the same sector at
 108 the same time of the day. That is, if we visited fifteen sectors moving downstream in
 109 one day, we would visit the next set of fifteen sectors while moving upstream on the
 110 subsequent day. Surveys took place from 2010 to 2017, with gaps in 2012 and 2015 due
 111 to a lack of resources (Table 1). Up to and including 2014, we visited the site every 30
 112 to 60 days, regardless of species activity, and stayed there from one to eight days,
 113 depending on breeding activity. From 2016 onward, we hired a local resident to visit the
 114 site once a week and send us information about Admirable Toad activity. Intending to
 115 increase capture success, we only visited the site if there was evidence of a breeding
 116 event. On each visit of this second stage, we stayed at the site for three days or until
 117 reproductive activity ceased, whichever came first. We limited visit duration to three
 118 days and the frequency of visits to once per month to minimize disturbance to the
 119 population.

120 **Table 1.** Surveys dates, number of individuals, and the number of captures of the
 121 Admirable Redbelly Toad throughout the study period (2010 – 2017). Detected
 122 breeding events are identified by their abbreviated month and followed by the
 123 corresponding number of trapping days in brackets. ‘Individuals’ refers to the number
 124 of individuals captured at least once in each year. ‘Captures (days)’ and ‘Captures
 125 (events)’ are, respectively, the number of captures summed over all fieldwork days and
 126 the number of captures summed over all events. The latter number excludes recaptures
 127 from the same event. Letters ‘F’ and ‘M’ stand for ‘female’ and ‘male’, respectively.

Year	Reproductive events	Individuals		Captures (days)		Captures (events)	
		F	M	F	M	F	M
		2010	Oct(1); Nov(1)	24	106	24	110

2011	Jul(2); Aug(2); Sep(2); Oct(1)	111	394	123	496	116	482
2013	Aug(2); Oct(6); Dec(1)	126	363	145	457	130	399
2014	Jul(1); Sep(5)	90	359	98	469	92	371
2016	Sep(3); Oct(2); Nov(2); Dec(1)	177	477	207	767	190	687
2017	Aug(3); Sep(2); Oct(2); Nov(2)	202	513	264	1085	240	931
Total	19 events (41 survey days)	556	1211	861	3384	792	2980

128

129 In the field, we visually scanned and hand-captured toads throughout the day
130 beginning at 9 am and stopping, sometimes at night, when new individuals were no
131 longer detected in the selected sectors. All captured toads were sexed and photographed.
132 Sex was determined based on a brownish nuptial pad at the base of the male thumb, the
133 calling behavior of males, or the position of pairs in amplexus. To minimize disturbance
134 and handling time, we did not clip toes or use any form of physical marking. Instead, we
135 photographed the ventral patch of black pigmentation on each animal and used its
136 unique outline for individual recognition (Figure S4). An alphanumeric code was
137 assigned to each capture, and a photographic catalogue was assembled and updated after
138 each sampling visit. We released all photographed toads at the same site of capture.

139 The individual identification of ventral patch photographs followed a semi-
140 automatic procedure. The best photograph from each individual's capture was cropped
141 to the area of interest, between the throat and the cloaca, and presented to the software
142 Wild-ID (Bolger et al., 2012), which returned the 20 closest matches in our photo
143 catalogue. This list was subsequently manually examined by a researcher who attributed
144 a new or existing individual identity to the captured toad. This approach was
145 successfully tested and applied to other species of South American Redbelly toads by
146 Caorsi et al. (2012) and by Bardier et al. (2019). The mapping of individual identities
147 over fieldwork days produced a database of capture histories spanning the entire

148 duration of our sampling period and included all captured toads. Field procedures
149 followed recommendations by the Chico Mendes Institute for Biodiversity
150 Conservation (ICMBio) under licenses number 40004-5 and 10341-1 issued by the
151 Information and Authorization System in Biodiversity (SISBIO) to MBM.

152 *Data analysis overview*

153 We organized captures through time in a hierarchical data structure where the
154 entire study period is divided into years, which are further divided into breeding events.
155 Each breeding event can also be divided into capture days, with the possibility for some
156 individuals to be recaptured on different days of the same event. For simplicity,
157 however, our analyses used the breeding event as the shortest time unit, treating
158 multiple captures of the same individual within one event as a single capture. Some
159 capture days, before 2016, did not coincide with a breeding event and were excluded
160 from our analysis based on criteria specified in the results section. In our data set, a
161 sample capture history could be given by:

162
$$X = 00\ 0110\ 011\ 00\ 1100\ 1011,$$

163 where year i represents the survey years in the study period ($i= 1 - 6$), and the breeding
164 event j varies from 1 to l_i , the maximum number of breeding events in a year i . Table 1
165 shows that l_i takes the values $\{2, 4, 3, 2, 4, 4\}$ as i varies from 1 to 6. A capture history
166 element X_{ij} equals 1 when the individual is captured at least once during the breeding
167 event j of the year i and equals 0 otherwise.

168 Our statistical analyses aimed to estimate the population size in each year and
169 understand the variation between years. To achieve this goal, we modeled the set of all
170 capture histories under three different approaches. Each approach has its own model

171 structure, but they all share the recognition that individuals may be alive and present in
172 the study site while not captured. First, we model capture histories within each year
173 using a closed-population capture-recapture model (henceforth called 'Closed'; Otis et
174 al., 1978). As a second approach, which considers the population open but still models
175 each year separately from the others, we employed the POPAN formulation of the Jolly-
176 Seber model (POPAN; Schwarz & Arnason, 2019; Schwarz & Arnason, 1996). Our
177 third approach, Pollock's Robust Design (PRD; Kendall, 2019; Kendall et al., 1997;
178 Pollock, 1982), considers the population closed within each year but open between
179 years. The PRD model combines information from all years in the same analysis. All
180 three approaches estimate yearly numbers of reproductively active males and females
181 present at the sampling site during the reproductive season. All models were
182 implemented in the RMark package (Laake & Rexstad, 2014), an R interface to build,
183 run and rank capture-recapture models in a likelihood framework using Program
184 MARK (White & Burnham, 1999). We ranked models within each approach using the
185 Akaike Information Criterion (AIC) adjusted for small sample sizes (AICc; Burnham &
186 Anderson, 2002) and drew inference about abundance from model-averaged estimates.
187 The following sections detail assumptions and model structure for each approach.

188 *Closed Population Capture-recapture*

189 The Closed model uses marked toads in successive breeding events of the same
190 year to estimate capture (p) and recapture (c) probabilities and the number of
191 individuals not captured (f_0) to obtain the abundance estimate. The approach assumes
192 geographic and demographic closure, i.e. no migration, recruitment, or mortality within
193 each breeding season. This model also assumes that marks are permanent. Our use of
194 natural marks for individual identification supports this assumption since the Admirable
195 Toad ventral patches do not change with age to the point of precluding individual

196 identification (Caorsi et al., *in prep*; Elgue et al., 2014). At the same time, the
197 noninvasiveness of our identification technique is unlikely to cause a trap response.
198 Finally, the Closed model assumes that all animals are available to be detected during
199 the annual survey period. We have relatively less evidence to support this assumption
200 and address the possibility of its failure with the POPAN and PRD models.

201 We implemented our Closed analysis using the ‘Full likelihood p and c ’ data
202 type (Lukacs, 2016), which directly estimates p (the probability of the first capture), c
203 (the probability of recapture) and f_0 (the number, or *frequency*, of individuals that were
204 observed 0 times, i.e., the estimated number of individuals in the population that were
205 never captured). Estimated population size (\hat{N}) under the Closed model is a derived
206 parameter, obtained with the equation:

$$207 \quad \hat{N} = \hat{f}_0 + M_{t+1},$$

208 where M_{t+1} is the total number of marked animals. Thus, since M_{t+1} is a constant
209 number, uncertainty about \hat{N} equals uncertainty about f_0 . We estimate f_0 separately for
210 males and females and use the logit link to model p and c as functions of covariates. We
211 let p vary with sex, amount of rain accumulated during the seven days before the onset
212 of surveys (rain), and with the median temperature of the first survey day in the event
213 (temperature). The parameter c is modeled as a function of sex and time, taking one
214 value for each event. We note that, even in the absence of trap aversion, it is reasonable
215 to consider the possibility of $p \neq c$, as well as their variation with sex and environmental
216 factors, because males and females may remain at the breeding site with different
217 probabilities, which in turn will be affected by the availability of pools. We built twelve
218 models that represent twelve hypothetical combinations of covariate effects and fit them
219 separately to data from each year with more than two reproductive events (Table S1).
220 The model set includes the possibility of constant p and c , represented by model ‘ $p(.)$

221 $c(\cdot)$ ' in Table S1. The year 2013 has only seven models because, in this year, we could
222 not obtain convergence for models with an effect of temperature on p .

223 *Jolly-Seber POPAN*

224 As in the Closed analysis, our implementation of POPAN models use each
225 year's data independently of the others. In contrast with Closed, however, POPAN
226 accounts for the possibility of entry and departure of individuals between breeding
227 events of the same year (Schwarz & Arnason, 2019). Specifically, the model includes
228 probabilities of entry (b) and survival (φ) between breeding events within a year. From
229 these two parameters, POPAN derives an estimate of the superpopulation size (N),
230 which is the total number of individuals entering the breeding site in one year. The
231 parameter b is the probability of each individual of the superpopulation entering the
232 study site before a given breeding event. Using indices to designate event-specific
233 parameter values, the expected number of individuals present during each breeding
234 event is given by:

$$235 \quad E[N_1] = Nb_0$$

$$236 \quad E[N_{t+1}] = E[N_t]\varphi_t + Nb_t,$$

237 where b_0 is the probability of each individual of the superpopulation (N) to be available
238 in the first breeding event. To account for imperfect detection, POPAN also estimates a
239 probability of detection per capture occasion (p). Parameters p , b , and φ can be held
240 constant or modeled as functions of covariates.

241 To keep the number of models down to an interpretable level, we followed a
242 sequential procedure of first fitting a set of capture probability functions while holding b
243 and φ at the most parameterized structure, which was φ as a function of sex and b as an

244 additive function of sex and breeding event. Our set of seven capture probability
245 functions consisted of all the additive combinations of the effects of sex, rain
246 (accumulated in the seven days before the sampling event), and temperature, including a
247 $p(\cdot)$ option of constant capture probability (Table S2). Combining the presence or
248 absence of an effect of sex on φ with the four possible additive combinations of the
249 effects of sex and time ('event') on b , we obtain eight possible joint models of φ and b .
250 In the second stage of our sequential procedure, we fit combinations of the eight joint
251 models of φ and b with all the models of p from the first stage that have $w > 0.1$ in each
252 year (Tables S3, S4, S5, and S6). The POPAN superpopulation estimate (N) for each
253 year is given for both males and females.

254 *Pollock's Robust Design*

255 The PRD approach combines the advantages of both open and closed population
256 models, providing estimates of abundance, apparent survival, and temporary emigration
257 (Kendall, 1999). This is the only one out of our three approaches that combines
258 information from all years in one single analysis. The PRD data structure aggregates
259 data on two temporal scales: the primary periods and, within each of these, multiple
260 secondary occasions. There should be sufficient time between consecutive primary
261 periods for the population to change via recruitment, mortality, and/or migration.
262 Secondary occasions, on the other hand, should be close enough in time to allow for
263 closure within each primary period. In our analysis, each year's reproductive season is a
264 primary period, subdivided in breeding events, which constitute secondary occasions.
265 According to the PRD model, each individual that is alive on primary period t survives
266 or does not permanently emigrate to primary period $t+1$ with probability φ , termed
267 apparent survival. Those individuals that survive or do not permanently emigrate may
268 be unavailable for detection during primary period $t+1$ with probability γ , termed

269 temporary emigration. Those that are available, with probability $(1 - \gamma)$, will be captured
270 according to probabilities of first capture (p) and recapture (c), as in the Closed model.
271 Also as in the Closed model, the PRD model derives yearly estimates of abundance
272 from the sum of the number of individuals captured in that year (or primary period) with
273 the estimated number of individuals that were alive and available at the site in that year
274 but were never captured (f_0). Thus, our estimates of abundance from this analysis in
275 year t refer to that part of the superpopulation, which was available to be captured at the
276 site (i.e. breeding site) in year t . In our application of the PRD, temporary emigration
277 may be random or Markovian (Kendall & Nichols, 1995; Kendall et al., 1997).
278 Markovian emigration draws a distinction between the probability γ'' that an individual
279 is unavailable in year $t+1$ given that it was available in year t , and the probability γ' that
280 an individual was unavailable in year $t+1$, given that it was unavailable in t . Under the
281 random emigration model, individuals become unavailable in year $t+1$ independently of
282 their state in year t .

283 As with the POPAN analysis, we use a sequential procedure to fit the sampling
284 and biological components of the PRD model. The most parameterized form of the
285 biological component represents ϕ , γ'' , and γ' as different additive functions of sex and
286 year. While holding the biological component of the model in its most parameterized
287 form, we fit 25 candidate models where c is either constant or a function of sex, and p
288 appears as various functions of effort, rain, temperature (linear and quadratic effects),
289 and sex (Table S7). The covariate effort equals the number of sampling days in a
290 breeding event (Table 1). The covariate rain is the amount of rainfall accumulated
291 during the seven days before the onset of sampling. After selecting the lowest AIC
292 structure for p and c , we subsequently fit and ranked 24 models of apparent survival and
293 temporary emigration (Table S8). These consist of Markovian and random temporary

294 emigration versions of twelve models of φ and γ . Apparent survival (φ) appears in four
295 structures: constant, as a function of sex, as a function of year, and as an additive
296 function of sex and year. Temporary emigration appears in the same structures except as
297 a constant parameter, because we didn't find that option biologically informative. We
298 considered the irregular time intervals between the primary periods to obtain annual
299 apparent survival and temporary emigration.

300 We compared estimates of abundance under the three analytical approaches;
301 nonetheless, we know that PRD extracts information from our multi-year dataset.
302 Therefore, we derive sex ratio and life expectancy statistics from the PRD results alone.
303 We obtain variances of derived statistics based on the Delta method (Powell, 2007)
304 implemented in the R package 'emdbook' (Bolker, 2019). The sex ratio values result
305 from the division of the estimated number of males by the estimated number of females.
306 We used the average annual apparent survival probability ($\bar{\varphi}$) to derive life expectancy
307 for the age of reproductive maturity e_m using Seber's (1973): $e_m = 0.5 + 1 / (1 - \bar{\varphi})$,
308 assuming that all adult individuals found in the breeding site are mature adults

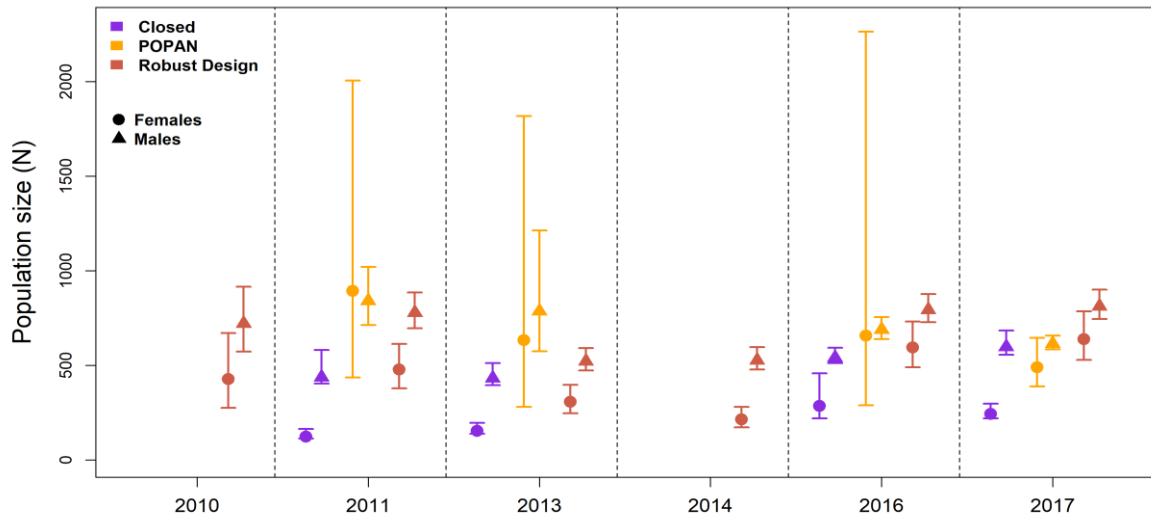
309 **Results**

310 *Overview of data and abundance estimates*

311 We obtained 4,620 captures of 1,862 adult Admirable Toad individuals during
312 88 days of fieldwork between October 2010 and November 2017. Capture effort and
313 capture success were unevenly distributed between the periods before and after 2016.
314 There was less effort in the latter period (18 days in 8 events), but an almost fourfold
315 increase in the number of captures per unit effort, from an average of 33 to 130 captures
316 per day. To focus on breeding events, we only analyze days with more than 30 captures,
317 thus working with a dataset of 41 capture days, all with evidence of reproductive

318 activity (*e.g.* recent clutches, pairs in amplexus, calling males). These contain 4,245
319 captures of 1,767 adults (Table 1), which is more than 90% of both captures and
320 individuals in the complete dataset. The number of breeding events detected per year
321 varied between two and four (Table 1). We did not fit POPAN and Closed Population
322 models to data from 2010 and 2014 because these years had only two breeding events
323 each. Fifty-one percent of all the individuals captured were recaptured at least once in a
324 different breeding event, while 16% were recaptured at least three times. The number of
325 males captured per breeding event was always between two and five times larger than
326 the number of females.

327 Estimates of the number of reproductive adults in the population varied more
328 between methods in the same year than between years. Abundance estimates (\pm
329 standard error) ranged from 561 ± 39 individuals (95% C.I. 484–638), for the Closed
330 Population model in 2011, to $1,734 \pm 361$ individuals (95% C.I. 1,026–2,442) for the
331 POPAN model, also in 2011 (Figure 2). POPAN results showed the least evidence of
332 temporal variability, while, at the same time, producing the least precise estimates.
333 Abundances estimated by the Closed Population model were consistently lower and
334 more precise than those obtained with POPAN or PRD. POPAN and PRD estimates
335 were closer to each other than either was to the estimates of the Closed Population
336 model. In all but one instance (POPAN 2011), the estimated number of males was
337 higher than that of females (Figure 2). Capture probability was also consistently higher
338 for males than for females, except for the closed population model results from 2011
339 (Table S9). Likewise, recapture probabilities estimated under the Closed and PRD
340 Models were higher for males than for females (Table S10). The apparent survival
341 estimated under POPAN is particularly low for females in 2016 (Table S11).

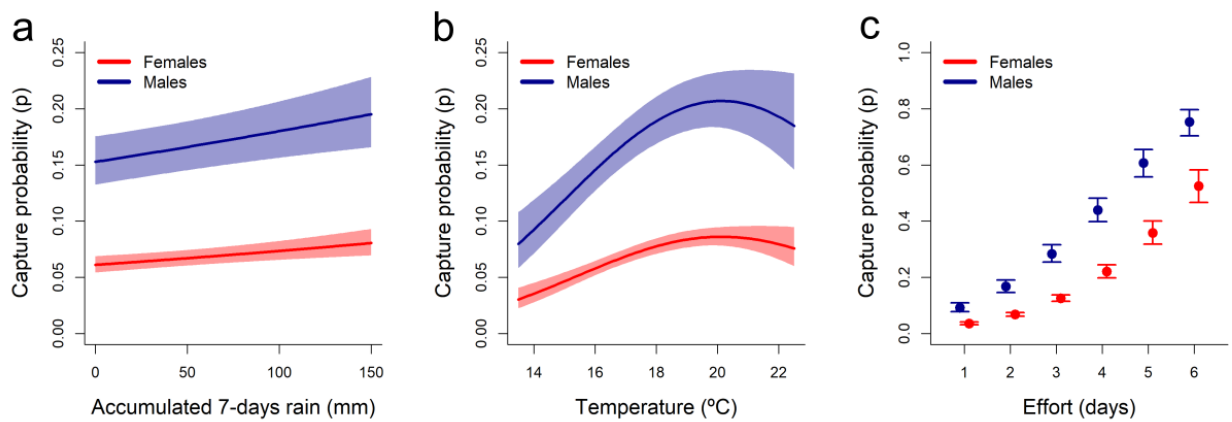


342 **Figure 2.** Estimates of adult female and male Admirable Redbelly Toad abundances per
 343 year, following the three approaches explored in this study: Closed Population, POPAN
 344 Jolly-Seber, and Pollock's Robust Design.

345
 346 *Robust Design Results*

347 The best-ranking model in our comparison of 25 PRD detection models with the same
 348 biological component has an AIC weight of 99% (Table S7). We thus model recapture
 349 probability (c) as a function of sex and capture probability (p) as a function of sampling
 350 effort, cumulative rainfall, a quadratic effect of temperature, and sex. The big difference
 351 in AIC between models #12 and #13 in Table S7 reveals the strong impact of sampling
 352 effort on p . Models with an effect of rainfall on p consistently rank better than similar
 353 models without rainfall. The same applies to models with a quadratic effect of
 354 temperature, which consistently rank better than similar models with only a linear effect
 355 of temperature. Sex explains a large part of the variation in detection probabilities (p
 356 and c), as expressed, for p , by the consistently higher ranking of models with an effect
 357 of sex, and for c by the more than 200 unit difference in AIC between models #23 and
 358 #25. Males presented higher capture and recapture probabilities than females (Table 2).
 359 Rainfall had a positive effect on p (Figure 3A), as expressed by a $\Delta AIC > 8$ for model

360 #2 in Table S7. The relationship between temperature and p shows a maximum capture
 361 probability near 20°C (Figure 3B). The quadratic nature of such relationship is
 362 supported by the considerable difference in AIC between models #3 and #1 (>10), as
 363 well as between models #10 and #8 (>5) in Table S7. The effort covariate explains most
 364 of the variation in p , as shown in Fig. 3C and in the more than 450 unit increase in AIC
 365 between model #12, the lowest-ranking model with effort, and model #13, the highest-
 366 ranking model without effort (Table S7).



367
 368 **Figure 3.** Relationship of capture probability (p) of adult female and male Admirable
 369 Redbelly Toad with accumulated rainfall (A), temperature (B), and sampling effort (C),
 370 under the top-ranking Pollock Robust Design model. Each plot's prediction holds the
 371 other two covariates at their average value. The structure of the top-ranking model is
 372 given by: $\phi(\text{year}) \gamma(\text{sex} + \text{year}) p(\text{sex} + \text{rain} + \text{temp}2 + \text{effort}) c(\text{sex}) f_{\theta}(\text{sex} * \text{year})$.

373

374 Model-averaged PRD parameter estimates show a decrease in abundance for the
 375 years 2013 and 2014 (Figure 2, Table 2). Such decrease coincides with relatively high
 376 temporary emigration in 2011 and 2013 when compared with 2010; also in agreement,
 377 survival probability for 2011 and 2012, before the period of decreased abundance, is
 378 estimated at its lowest value throughout the study period (Table 2). The lowest
 379 abundance estimate, for the year 2014, coincides with the most biased sex ratio, of
 380 nearly two males per female in the population. When abundance was highest, in 2017,

381 the sex ratio was statistically indistinguishable from 1. There is no evidence of
 382 differential survival between males and females throughout the period, but females do
 383 show consistently higher values of temporary emigration than males. In agreement with
 384 the model selection results of the previous paragraph, we see no evidence of difference
 385 between model-averaged γ'' and γ' . The model-averaged point estimate of life
 386 expectancy for females (8.46 ± 1.65 years) is slightly higher than that of males
 387 (7.50 ± 0.77 years) but the uncertainty about those estimates is too large to establish a
 388 statistical difference between the sexes.

389 **Table 2.** Yearly abundance, sex ratio, annual apparent survival, and annual temporary
 390 emigration of adult female and male Admirable Redbelly Toad obtained under the top-
 391 ranking Pollock Robust Design model. Sex ratios are expressed as the estimated number
 392 of males (M) divided by females (F). Temporary Emigration'' is the probability that a
 393 toad present in the current year leaves the site, whereas Temporary Emigration' is the
 394 probability that a toad absent in the current year remains out of the breeding site. Values
 395 of temporary emigration and survival are not estimable for the last year of the study,
 396 and temporary emigration', likewise, cannot be estimated for the first year. Asterisks
 397 indicate uninterpretable estimates, with confidence intervals between zero and one.
 398 Survival and temporary emigration estimates for the years 2011 and 2014 are derived
 399 under the simplifying assumption of constant survival and temporary emigration during
 400 the two-year intervals 2011-2012 and 2014-2015, respectively.

Year	Abundance	Sex Ratio	Survival		Temporary Emigration''		Temporary Emigration'	
			F	M	F	M	F	M
2010	1148±141	1.68±0.14	0.99±0.01*	0.99±0.01*	0.20±0.20	0.09±0.09	-	-
2011	1256±87	1.63±0.14	0.80±0.04	0.78±0.01	0.60±0.08	0.32±0.06	0.57±0.16	0.30±0.09
2013	828±59	1.70±0.11	0.93±0.04	0.92±0.02	0.65±0.07	0.37±0.05	0.66±0.08	0.36±0.06
2014	747±49	2.45±0.31	0.82±0.03	0.80±0.04	0.001±0.012*	0.000±0.004*	0.000±0.015*	0.000±0.012*
2016	1388±82	1.33±0.13	0.94±0.05	0.93±0.05	0.002±0.052*	0.001±0.017*	0.001±0.033*	0.001±0.033*
2017	1451±87	1.27±0.22	-	-	-	-	-	-

401

402 **Discussion**

403 Knowledge about population size is key for effective wildlife management and
404 for the assessment of species' conservation status (IUCN, 2012). If we may consider the
405 Pollock Robust Design as the most adequate model for this biological system, our
406 results reveal a breeding population of Admirable Toad varying between 650 and 1,650
407 individuals (PRD model, lower confidence limit – higher confidence limit) over eight
408 years, while spending more than half of the time above 1,000 individuals. Accounting
409 for temporary emigration, in those years for which we could estimate it, the total
410 population of breeding and non-breeding toads would fall between 1,500 and 2,500. Is
411 this the global Admirable Redbelly Toad population size? Since 2010, we have searched
412 for other populations near our study site, along the Forqueta River, and throughout the
413 Taquari-Antas basin. Despite our search over more than seven kilometers of forested
414 river margins (Fonte et al., 2014), all we found was a group of fewer than 20
415 individuals, in 2017, breeding on a small rocky outcrop 1,500 m downstream from our
416 site (Abadie et al., 2021 – Chapter 3). Across the Forqueta River from our sampling
417 area, there is another small (~200 m²) rocky outcrop with confirmed *M. admirabilis*
418 breeding activity. We cannot confirm that breeding on the right bank of the river occurs
419 as frequently as on the left. Nonetheless, even if it does, and if it includes approximately
420 15% of the individuals estimated for the left bank, the total breeding population size
421 will be lower than 2000 individuals.

422 Our methodological choice of estimating abundance with three different
423 approaches aims at comparing results under a variety of assumptions. The variation
424 between approaches spans hundreds of individuals, an arguably small difference but
425 still revealing informative patterns. The Closed population model consistently produced
426 the lowest abundance estimates, especially for females, in all years, regardless of effort

427 or number of captures. The Closed model assumes closure throughout the sampling
428 period and applies to data from only one year at a time. Therefore, any individual that is
429 not available for capture because it did not breed in our sampling area in one year will
430 be treated as non-existent. Understandably, such a model should produce relatively low
431 estimates, even if it is one of the most reasonable analytical options with a single year of
432 data. One should look for alternative approaches when data from different years reveal
433 information about temporary unavailability. Also, when there are intervals of several
434 weeks between breeding events—as in our case—one should be cautious about
435 assuming closure within a whole year, or breeding season.

436 Pellet et al. (2011) suggested that POPAN is a useful approach for amphibians
437 with reproductive strategies that are less amenable to closure assumptions, as in the case
438 of prolonged (i.e. not explosive) reproduction. Using a simulation study, they estimated
439 abundances under scenarios of high and low survival and capture probabilities and
440 found that lower values of these probabilities led to higher inaccuracy in abundance
441 estimates. In our study, POPAN results are the least precise among the three
442 approaches, particularly for female estimates, likely due to low capture probabilities
443 (Table S9). The POPAN estimates of apparent survival refer to intervals between
444 breeding events and are thus partly interpretable as emigration from the breeding site.
445 This estimated apparent survival is particularly low for females in 2016, in agreement
446 with the low number of 2016 female recaptures, evident in Table 1. When working with
447 data from only one year, one may face the choice between Closed and POPAN
448 approaches. We believe that this choice should not be guided by the low precision of the
449 POPAN estimates, as it is better to be uncertain about a reasonable result than to be
450 certain about a biased estimate. POPAN is an interesting way of accounting for
451 inevitable changes in the availability of individuals between breeding events. Although

452 we agree with Pellet et al.'s (2011) suggestion that POPAN is appropriate for prolonged
453 breeders, since it models the superpopulation of toads using the breeding site
454 throughout one reproductive season, we believe that it applies equally well to seasonal
455 explosive breeders. The choice between POPAN and Closed, however, may also be
456 informed by comparison with results from a PRD analysis, which simultaneously
457 models data from the whole study period.

458 The PRD analysis assumes closure within each year, just like the Closed model
459 does; however, individuals that are unavailable in any given year are treated by the PRD
460 model as temporarily emigrated. A probability of temporary emigration is estimated,
461 and both survival and capture parameter estimates will reflect the inference on
462 emigration. One key benefit of PRD models is this ability to quantify temporary
463 emigration from the information of individuals that are not captured in one year but
464 reappear in future years, being unequivocally part of the superpopulation (Kendall,
465 1999). The difference between Closed and PRD abundance estimates, however, goes
466 beyond the estimation of temporary emigration. Our PRD estimates of capture
467 probability (p) were always lower than those obtained under the Closed analysis, for
468 both males and females (Table S9). This consistent difference in p explains why the
469 PRD abundances tend to be higher than their Closed counterparts, even after accounting
470 for temporary emigration. In the end, despite the lack of precision in the POPAN
471 estimates for 2011, 2013, and 2016, we would recommend the use of POPAN to anyone
472 who had only one year of data obtained under circumstances similar to those of our
473 study. Figure 2 and the inevitable lack of within-year closure lead us to believe that
474 Closed models are underestimating abundance, and especially so for females.

475 The analysis of capture histories that span multiple breeding seasons, afforded
476 by the PRD, also improves our ability to examine temporal changes in population size.

477 The estimated number of breeding individuals is lowest in 2013 and 2014 (Table 2).
478 Such a decrease coincides with the highest estimated temporary emigration
479 probabilities, for both sexes, which occurred in 2013 (Table 2). It also follows the
480 lowest estimated apparent survival for males and females, obtained in 2011. Since
481 temporary emigration varies more than survival, we are inclined to attribute most of the
482 variation in abundance to temporary emigration, but some of the decrease in abundance
483 could also result from reduced recruitment in 2012. The (austral) summer of 2012 saw
484 the most severe drought of the previous 15 years in Southern Brazil (Zepner et al., 2020;
485 Estado do Rio Grande do Sul, 2012). If Admirable Toads take one or two years to reach
486 maturity, which appears likely in the similar-sized *Melanophryniscus moreirae* (Jeckel,
487 Saporito, & Grant, 2015), unfavorable conditions to reproduce (and consequently fewer
488 couples breeding), clutch desiccation and/or high juvenile mortality in 2012 could
489 impact breeding population size in 2013 and 2014.

490 Admirable Redbelly Toad breeding events often concentrate several individuals
491 in one pond with multiple males trying to mate with one female at the same time.
492 Understandably, females may find such episodes stressful and possibly evolve
493 mechanisms for successful breeding while minimizing exposure to large numbers of
494 sexually active males. Besides anecdotal evidence of behavioral avoidance, there at
495 least two lines of population-level evidence that are compatible with exposure
496 minimization: first, females have higher temporary emigration (Table 2); and second,
497 among individuals that are available for capture, females have lower capture (Table S9)
498 and recapture (Table S10) probabilities than males. After accounting for differences in
499 capture probability between sexes, we estimate sex ratios between 1.27 and 2.45 males
500 per female. These estimates are lower than the average 3.3 ratio of captured males to
501 females in Table 1, but they are statistically different from 1 in all but the last year.

502 Male-biased sex ratios have been reported for other bufonid species based on
503 capture counts (Lampo et al., 2012), and for other species of *Melanophryniscus* based
504 on capture counts (Cairo et al., 2013; Pereira & Maneyro, 2018; Vaira, 2005) and
505 abundance estimates (Bardier et al., 2019). According to Wells (1977), anuran males
506 usually outnumber females at explosive mating aggregations as well as throughout the
507 breeding season, a pattern that should result in competition for mates (Lodé et al.,
508 2005). Zug & Zug (1979) compiled evidence of intraspecific variation in the sex ratio of
509 anuran populations and suggested that extensive variation among *Rhinella marina*
510 populations was more due to the local environment than to phylogenetic history. Some
511 anuran females become reproductively active at an older age than males. This seems to
512 apply to *Melanophryniscus moreirae* as suggested by Jeckel et al (2015) and is common
513 in sexually dimorphic species where females are larger than males (Monnet & Cherry,
514 2002). Considering we found evidence of similar survival for both sexes (or slightly
515 higher survival for females), and assuming balanced sex ratios at birth, we could expect
516 to find more males than females in the breeding population as a possible role of age
517 maturity in the determination of sex ratios.

518 Demographic information about amphibians is scarce and patchy (Conde et al.,
519 2019), but we found our estimates of Admirable Toad annual survival probability are
520 generally higher than those published for another redbelly toad (*M. montevidensis*,
521 Bardier et al., 2019) or other bufonids (Lampo et al., 2012; Vasconcellos & Colli,
522 2009). If *M. admirabilis* survival is indeed high, we hypothesize that it may be due to
523 chemical defense against predators. Among Bufonidae anurans, *Melanophryniscus* is
524 the only genus that has lipophilic alkaloids for chemical defense (Daly et al., 1984;
525 Hantak et al., 2013). Species of the genus accumulate abundant and diverse alkaloids
526 throughout their lives, from the ingestion of arthropods rich in these toxic substances

527 (Jeckel et al., 2015). The older the toad, the greater the diversity of alkaloids found in
528 the skin and internal organs (Grant et al., 2012; Jeckel et al., 2015). Although not yet
529 clear, alkaloid sequestration may reduce the range of predators that can feed on
530 sequestering species (Savitzky et al., 2012; Stynoski et al., 2014; Toledo & Jared,
531 1995), as well as minimize the chance of parasite infection (Grant et al., 2012; Mina et
532 al., 2015). In our case, annual survival estimates resulted in a life expectancy of about
533 eight years for an adult toad. Such life expectancies are surprisingly high for animals as
534 small as Admirable Redbelly Toad, but they are well supported by our data, which
535 includes live recaptures seven years after the first capture for both sexes (see Abadie et
536 al, 2021 – Chapter 1).

537 Based on this study, we are confident that the only known population of *M.*
538 *admirabilis* has fewer than 2000 reproductive individuals. Our comparison of analytical
539 approaches highlights the usefulness of carrying out multi-year monitoring efforts that
540 account for population openness due to recruitment, mortality, and movement of live
541 individuals. When multi-year analyses are not possible, our results suggest it is best to
542 employ analysis that accounts for movement in and out of the study area throughout one
543 breeding season. Such movements, within or between seasons, may be the most
544 important factor behind the substantial temporal variation in the number of reproductive
545 Admirable Toads, even though we see reasons to suspect that breeding failure during an
546 extremely dry year may have contributed as well. Eight years of observation do not
547 inform a proper assessment of extinction risk (Fieberg & Ellner, 2000) but we find it
548 striking that such a small population could decrease to almost one half of its initial size
549 and subsequently rebound during such a short time. If high survival probability is a
550 characteristic trait of the Admirable Redbelly Toad, we wonder how much does it
551 contribute to the persistence of the species at a low population size.

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569 **References**

- 570 Alvares, C.A., Stape, J.L., Sentelhas, P.C., De Moraes Gonçalves, J.L. & Sparovek, G.
571 (2013). Köppen's climate classification map for Brazil. *Meteorologische*
572 *Zeitschrift*, 22(6), 711–728. <https://doi.org/10.1127/0941-2948/2013/0507>
- 573 Bardier, C., Martínez-Latorraca, N., Porley, J. L., Bortolini, S. V., Cabrera Alonzo, N.,
574 Maneyro, R., & Toledo, L. F. (2019). Seasonal demography of the threatened
575 Montevideo Redbelly Toad (*Melanophryniscus montevidensis*) in a protected area
576 of Uruguay. *Canadian Journal of Zoology*, 97(2), 131–141. doi: 10.1139/cjz-2017-
577 0362
- 578 Bolger, D. T., Morrison, T. A., Vance, B., Lee, D., & Farid, H. (2012). A computer-
579 assisted system for photographic mark-recapture analysis. *Methods in Ecology and*

580 *Evolution*, 3(5), 813–822. doi: 10.1111/j.2041-210X.2012.00212.x

581 Bolker, B. (2019). *Package ‘emdbook.’*

582 Burnham, K. P., & Anderson, D. R. (2002). *Model selection and multimodel inference:*
583 *a practical information-theoretic approach.* Springer-Verlag.

584 Cairo, S. L., Zalba, S. M., & Úbeda, C. A. (2013). Reproductive pattern in the
585 southernmost populations of South American redbelly toads. *Journal of Natural*
586 *History*, 47(31–32), 2125–2134. doi: 10.1080/00222933.2013.769644

587 Caorsi, V. Z., Santos, R. R., & Grant, T. (2012). Clip or Snap? An Evaluation of Toe-
588 Clipping and Photo-Identification Methods for Identifying Individual Southern
589 Red-Bellied Toads, *Melanophryniscus cambaraensis*. *South American Journal of*
590 *Herpetology*, 7(2), 79–84. doi: 10.2994/057.007.0210

591 Caorsi, V., Abadie, M., Santos, R.F., Colombo, P. & Borges-Martins, M. (*in prep*).
592 Long-term colour pattern study in South American Red-Bellied Toads (Genus
593 *Melanophryniscus*).

594 Conde, D. A., Staerk, J., Colchero, F., da Silva, R., Schöley, J., Baden, H. M., ...
595 Vaupel, J. W. (2019). Data gaps and opportunities for comparative and
596 conservation biology. *Proceedings of the National Academy of Sciences*, 116(19),
597 9658–9664. doi: 10.1073/pnas.1816367116

598 Daly, J. W., Highet, R. J., & Myers, C. W. (1984). Occurrence of skin alkaloids in non-
599 dendrobatid frogs from Brazil (Bufonidae), Australia (Myobatrachidae) and
600 Madagascar (Mantellinae). *Toxicon*, 22(6), 905–919. doi: 10.1016/0041-
601 0101(84)90182-X

602 Di-Bernardo, M., Maneyro, R., & Grillo, H. (2006). New Species of *Melanophryniscus*
603 (Anura : Bufonidae) from Rio Grande do Sul, Southern Brazil. *Journal of*
604 *Herpetology*, 40(2), 261–266.

605 Elgue, E., Pereira, G., Achaval-Coppes, F., & Maneyro, R. (2014). Validity of photo-
606 identification technique to analyze natural markings in *Melanophryniscus*
607 *montevideensis* (Anura: Bufonidae). *Phyllomedusa*, 13(1), 59–66. doi:
608 10.11606/issn.2316-9079.v13i1p59-66

609 Estado do Rio Grande do Sul. (2012). Conselho Permanente de Agrometeorologia
610 Aplicada do Estado do Rio Grande do Sul: Boletim de informações nº 31, 19 de
611 janeiro de 2012.
612 http://www.fepagro.rs.gov.br/upload/20120120125230boletim_copaaergs_extraord
613 [inariano_jan2012.pdf](http://www.fepagro.rs.gov.br/upload/20120120125230boletim_copaaergs_extraord). Last accessed on 15 November 2019.

614 Fieberg, J., & Ellner, S. P. (2000). When is it meaningful to estimate an extinction
615 probability? *Ecology*, 81(7), 2040–2047. doi: 10.1890/0012-
616 9658(2000)081[2040:WIIMTE]2.0.CO;2

617 Fonte, L. F. M. da, Abadie, M., Mendes, T., Zank, C., & Borges-Martins, M. (2014).
618 The Times they are a-Changing: How a MultiInstitutional Effort Stopped the
619 Construction of a Hydroelectric Power Plant that Threatened a Critically
620 Endangered Red-Belly Toad in Southern Brazil. *FrogLog*, 22(4), 18–21.

621 Grant, T., Colombo, P., Verrastro, L., & Saporito, R. A. (2012). The occurrence of
622 defensive alkaloids in non-integumentary tissues of the Brazilian red-belly toad
623 *Melanophryniscus simplex* (Bufonidae). *Chemoecology*, 22(3), 169–178. doi:
624 10.1007/s00049-012-0107-9

625 Hantak, M. M., Grant, T., Reinsch, S., McGinnity, D., Loring, M., Toyooka, N., &
626 Saporito, R. A. (2013). Dietary Alkaloid Sequestration in a Poison Frog: An
627 Experimental Test of Alkaloid Uptake in *Melanophryniscus stelzneri* (Bufonidae).
628 *Journal of Chemical Ecology*, 39(11–12), 1400–1406. doi: 10.1007/s10886-013-
629 0361-5

- 630 Inchausti, P., & Halley, J. (2003). On the relation between temporal variability and
631 persistence time in animal populations. *Journal of Animal Ecology*, 72(6), 899–
632 908. doi: 10.1046/j.1365-2656.2003.00767.x
- 633 IUCN. (2012). *Guidelines for application of IUCN red list criteria at regional and*
634 *national levels: version 4.0*. Retrieved from [www.iucnredlist.org/technical-](http://www.iucnredlist.org/technical-documents/categories-and-criteria)
635 [documents/categories-and-criteria](http://www.iucnredlist.org/technical-documents/categories-and-criteria)
- 636 IUCN SSC Amphibian Specialist Group. (2013). *Melanophryniscus admirabilis*. In *The*
637 *IUCN Red List of Threatened Species 2013*. Retrieved from
638 <http://dx.doi.org/10.2305/IUCN.UK.2013-2.RLTS.T135993A44846478.en>.
- 639 Jeckel, A. M., Saporito, R. a., & Grant, T. (2015). The relationship between poison frog
640 chemical defenses and age, body size, and sex. *Frontiers in Zoology*, 12(1), 27.
641 doi: 10.1186/s12983-015-0120-2
- 642 Kendall, W. (1999). Robustness of closed capture–recapture methods to violations of
643 the closure assumption. *Ecology*, 80(8), 2517–2525.
- 644 Kendall, W. (2019). The “robust design”... In E. G. Cooch & G. C. White (Eds.),
645 *Program MARK: A gentle introduction* (pp. 1–48). Retrieved from
646 <http://www.phidot.org/software/mark/docs/book/pdf/chap15.pdf>
- 647 Kendall, W., & Nichols, J. D. (1995). On the use of secondary capture-recapture
648 samples to estimate temporary emigration and breeding proportions. *Journal of*
649 *Applied Statistics*, 22(5–6), 751–762. doi: 10.1080/02664769524595
- 650 Kendall, W., Nichols, J., & Hines, J. E. (1997). Estimating temporary emigration using
651 capture-recapture data with Pollock’s Robust Design. *Ecology*, 78(2), 563–578.
- 652 Laake, J., & Rexstad, E. (2014). RMark - an alternative approach to building linear
653 models in MARK. *Mark*, 113pp.
- 654 Lampo, M., Celsa, S. J., Rodríguez-Contreras, A., Rojas-Runjaic, F., & García, C. Z.
655 (2012). High Turnover Rates in Remnant Populations of the Harlequin Frog
656 *Atelopus cruciger* (Bufonidae): Low Risk of Extinction? *Biotropica*, 44(3), 420–
657 426. doi: 10.1111/j.1744-7429.2011.00830.x
- 658 Lodé, T., Holveck, M. J., & Lesbarrères, D. (2005). Asynchronous arrival pattern,
659 operational sex ratio and occurrence of multiple paternities in a territorial breeding
660 anuran, *Rana dalmatina*. *Biological Journal of the Linnean Society*, 86(2), 191–
661 200. doi: 10.1111/j.1095-8312.2005.00521.x
- 662 Lukacs, P. (2016). Chapter 14 – Closed population abundance models. In *A gentle*
663 *introduction to MARK* (pp. 1–43).
- 664 Mina, A. E., Ponti, A. K., Woodcraft, N. L., Johnson, E. E., & Saporito, R. A. (2015).
665 Variation in alkaloid-based microbial defenses of the dendrobatid poison frog
666 *Oophaga pumilio*. *Chemoecology*, 25(4), 169–178. doi: 10.1007/s00049-015-0186-
667 5
- 668 Monnet, J.-M., & Cherry, M. I. (2002). Sexual size dimorphism in anurans.
669 *Proceedings. Biological Sciences / The Royal Society*, 269(1507), 2301–2307. doi:
670 10.1098/rspb.2002.2170
- 671 Otis, D. L., Burnham, K. P., White, G. C., & Anderson, D. R. (1978). Statistical
672 inference from capture data on closed animal populations. *Wildlife Monographs*,
673 (62), 3–135.
- 674 Pellet, J., Schmidt, B. R., Wagner, N., Lötters, S., & Schmitt, T. (2011). The
675 superpopulation approach for estimating the population size of “prolonged”
676 breeding amphibians: Examples from Europe. *Amphibia-Reptilia*, 32(3), 323–332.
- 677 Pereira, G., & Maneyro, R. (2018). Reproductive biology of *Melanophryniscus*
678 *montevideensis* (Anura: Bufonidae) from Uruguay: reproductive effort, fecundity,
679 sex ratio and sexual size dimorphism. *Studies on Neotropical Fauna and*

680 *Environment*, 53(1), 10–21. doi: 10.1080/01650521.2017.1364952

681 Pertoldi, C., Bach, L. A., & Loeschcke, V. (2008). On the brink between extinction and
682 persistence. *Biology Direct*, 3(1), 47. doi: 10.1186/1745-6150-3-47

683 Pimm, S. L., Jenkins, C. N., Abell, R., Brooks, T. M., Gittleman, J. L., Joppa, L. N., ...
684 Sexton, J. O. (2014). The biodiversity of species and their rates of extinction,
685 distribution, and protection. *Science*, 344(6187). doi: 10.1126/science.1246752

686 Pollock, K. H. (1982). A Capture-Recapture Design Robust to Unequal Probability of
687 Capture. *The Journal of Wildlife Management*, 46(3), 752. doi: 10.2307/3808568

688 Powell, L. A. (2007). Approximating Variance of Demographic Parameters Using the
689 Delta Method : A Reference for Avian Biologists. *The Condor*, 109(4), 949–954.

690 Rabinowitz, D. (1981). Seven forms of rarity. *The Biological Aspects of Rare Plant*
691 *Conservation*, 205–217. doi: 10.1177/004728702237415

692 Savitzky, A. H., Mori, A., Hutchinson, D. A., Saporito, R. A., Burghardt, G. M.,
693 Lillywhite, H. B., & Meinwald, J. (2012). Sequestered defensive toxins in tetrapod
694 vertebrates : principles , patterns , and prospects for future studies. *Chemoecology*,
695 141–158. doi: 10.1007/s00049-012-0112-z

696 Schwarz, C J, & Arnason, A. N. (2019). Jolly-Seber models in MARK. In E. G. Cooch
697 & G. C. White (Eds.), *Program MARK: A gentle introduction*, (pp. 1–51).

698 Schwarz, Carl James, & Arnason, A. N. (1996). A General Methodology for the
699 Analysis of Capture-Recapture Experiments in Open Populations. *Biometrics*,
700 52(3), 860. doi: 10.2307/2533048

701 Seber, G. A. F. (1973). *The Estimation of Animal Abundance and Related Parameters*.
702 New York: Hafner Press.

703 Stynoski, J. L., Torres-Mendoza, Y., Sasa-Marin, M., & Saporito, R. A. (2014).
704 Evidence of maternal provisioning of alkaloid-based chemical defenses in the
705 strawberry poison frog *Oophaga pumilio*. *Ecology*, 95(3), 587–593. doi:
706 10.1890/13-0927.1

707 Toledo, R. C., & Jared, C. (1995). Cutaneous granular glands and amphibian venoms.
708 *Comparative Biochemistry and Physiology*, Vol. 111, pp. 1–29. doi: 10.1016/0300-
709 9629(95)98515-1

710 Vaira, M. (2005). Annual variation of breeding patterns of the toad, *Melanophryniscus*
711 *rubriventris* (Vellard, 1947). *Amphibia-Reptilia*, 26(2), 193–199. doi:
712 10.1163/1568538054253519

713 Vasconcellos, M. M., & Colli, G. R. (2009). Factors Affecting the Population Dynamics
714 of Two Toads (Anura: Bufonidae) in a Seasonal Neotropical Savanna. *Copeia*,
715 2009(2), 266–276. doi: 10.1643/CE-07-099

716 Villalobos, F., Dobrovolski, R., Provete, D. B., & Gouveia, S. F. (2013). Is Rich and
717 Rare the Common Share? Describing Biodiversity Patterns to Inform Conservation
718 Practices for South American Anurans. *PLoS ONE*, 8(2), e56073. doi:
719 10.1371/journal.pone.0056073

720 Wells, K. D. (1977). The social behaviour of anuran amphibians. *Animal Behaviour*, 25,
721 666–693.

722 White, G. C., & Burnham, K. P. (1999). Program MARK: survival estimation from
723 populations of marked animals. *Bird Study*, 46(February), 120–139. doi:
724 10.1080/00063659909477239

725 Williams, B. K., Nichols, J. D., & Conroy, M. J. (2002). *Analysis and management of*
726 *animal populations*. Academic Press.

727 Zank, C., Becker, F. G., Abadie, M., Baldo, D., Maneyro, R., & Borges-Martins, M.
728 (2014). Climate change and the distribution of neotropical red-bellied toads
729 (*Melanophryniscus*, Anura, Amphibia): How to prioritize species and populations?

730 *PLoS ONE*, 9(4), 1–11. doi: 10.1371/journal.pone.0094625
731 Zepner, L., Karrasch, P., Wiemann, F. & Bernard, L. (2020). ClimateCharts.net – an
732 interactive climate analysis web platform. *International Journal of Digital Earth*,
733 14(3), 338-356. <https://doi.org/10.1080/17538947.2020.1829112>
734 Zug, G. R., & Zug, P. B. (1979). The marine toad, *Bufo marinus*: a natural history
735 resumé of native populations. *Smithsonian Contributions to Zoology*, (284), 1–58.
736 doi: 10.5479/si.00810282.284.
737

738 Online Supporting Information

739 **Abundance of the microendemic Admirable Redbelly Toad:**
740 **a comparison of population size estimates.**

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Michelle Abadie, Márcio Borges-Martins, Thayná Mendes, Murilo Guimarães.



744

745 **Figure S1.** An individual of Admirable Redbelly Toad, *Melanophryniscus admirabilis* (snout-vent length
746 about 3.5 cm). Photo: Documentation of Threatened Species Project (DoTS).



747

748 **Figure S2.** Forest cover and steep slopes on both sides of the Forqueta River at the Perau de Janeiro
749 region. Photo: Documentation of Threatened Species Project (DoTS).

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751

752 **Figure S3.** Breeding site of Admirable Redbelly Toad, at Perau de Janeiro, Arvorezinha/RS, Brazil. (a)
753 flattened rocky outcrop along the left bank of the Forqueta River; (b) a small ephemeral pool on the rocky
754 margins; (c) male individual of ARBT calling on a small pool at rocky outcrop; and (d) a pair in
755 amplexus surrounded by egg clutches.



756

757 **Figure S4.** Individual variation in ventral patch of black pigmentation used as marking for individual
758 recognition of the Admirable Redbelly Toad.

759

760 **Table S1.** Model-selection results for year-specific Closed population model analyses of adult female
761 and male Admirable Redbelly Toad. Letters in front of parentheses stand for first capture (p) and
762 recapture (c) probabilities. Words in parentheses indicate covariates for each parameter, where ‘sex’
763 denotes differences between males and females, ‘rain’ for rainfall accumulated in seven days before
764 sampling, ‘temp’ for the median temperature of the first sampling day, and ‘effort’ for the number of days
765 in each sampling event. For simplicity, we omit model structure specification for the number of
766 individuals that were never caught, which was fixed as $f0$ (sex).

Year	#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
2011	1	$p(\text{rain} + \text{temp}) c(\text{sex} + \text{event})$	-2878.4	0	0.73	9	54.3900
	2	$p(\text{sex} + \text{rain} + \text{temp}) c(\text{sex} + \text{event})$	-2876.4	2.00	0.27	10	54.3714
	3	$p(\text{temp}) c(\text{sex} + \text{event})$	-2866.0	12.39	0.00	8	68.8033
	4	$p(\text{sex} + \text{temp}) c(\text{sex} + \text{event})$	-2864.4	14.00	0.00	9	68.3904
	5	$p(\text{sex} + \text{rain} + \text{temp}) c(\text{sex})$	-2859.5	18.91	0.00	8	75.3155
	6	$p(\text{rain} + \text{temp}) c(\text{event})$	-2855.5	22.84	0.00	8	79.2484
	7	$p(\text{sex} + \text{temp}) c(\text{sex})$	-2847.5	30.91	0.00	7	89.3345
	8	$p(\text{rain}) c(\text{sex} + \text{event})$	-2828.9	49.53	0.00	8	105.9376
	9	$p(\text{sex} + \text{rain}) c(\text{sex} + \text{event})$	-2826.9	51.54	0.00	9	105.9285
	10	$p(\text{sex}) c(\text{sex} + \text{event})$	-2805.4	73.02	0.00	8	129.4241
	11	$p(\text{sex}) c(\text{sex})$	-2788.5	89.93	0.00	6	150.3682
	12	$p(.) c(.)$	-2770.6	107.79	0.00	4	172.2563
2013	1	$p(\text{rain}) c(\text{sex} + \text{event})$	-3708.0	0	0.70	7	14.8753
	2	$p(\text{sex} + \text{rain}) c(\text{sex} + \text{event})$	-3706.1	1.84	0.28	8	14.6955
	3	$p(\text{sex} + \text{rain}) c(\text{event})$	-3701.3	6.67	0.02	7	21.5471
	4	$p(\text{sex} + \text{rain}) c(\text{sex})$	-3648.1	59.90	0.00	7	74.7794
	5	$p(\text{sex}) c(\text{sex} + \text{event})$	-3220.7	487.31	0.00	7	502.1853
	6	$p(\text{sex}) c(\text{sex})$	-3162.6	545.37	0.00	6	562.2692
	7	$p(.) c(.)$	-3159.8	548.15	0.00	4	569.0736
2016	1	$p(\text{sex} + \text{rain} + \text{temp}) c(\text{sex} + \text{event})$	-3883.4	0	0.98	10	70.2127
	2	$p(\text{rain} + \text{temp}) c(\text{sex} + \text{event})$	-3875.0	8.41	0.01	9	80.6358
	3	$p(\text{sex} + \text{rain}) c(\text{sex} + \text{event})$	-3867.5	15.95	0.00	9	88.1785
	4	$p(\text{rain}) c(\text{sex} + \text{event})$	-3857.8	25.58	0.00	8	99.8269
	5	$p(\text{sex} + \text{temp}) c(\text{sex} + \text{event})$	-3818.6	64.85	0.00	9	137.0755
	6	$p(\text{rain} + \text{temp}) c(\text{event})$	-3811.0	72.45	0.00	8	146.6957
	7	$p(\text{temp}) c(\text{sex} + \text{event})$	-3809.8	73.60	0.00	8	147.8461
	8	$p(\text{sex}) c(\text{sex} + \text{event})$	-3778.2	105.19	0.00	8	179.4330
	9	$p(\text{sex} + \text{rain} + \text{temp}) c(\text{sex})$	-3761.7	121.77	0.00	8	196.0085
	10	$p(\text{sex} + \text{temp}) c(\text{sex})$	-3696.8	186.61	0.00	7	262.8713
	11	$p(\text{sex}) c(\text{sex})$	-3656.5	226.96	0.00	6	305.2288
	12	$p(.) c(.)$	-3585.1	298.29	0.00	4	380.5720

767 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes

768 ^b Δ AICc = difference in the AICc values between top model and the current model

769 ^c w = AIC weight

770 ^d k = number of estimated parameters

771 ^e Deviance = difference between the saturated model and the current model

772 **Table S1.** (Cont.)

Year	#	Model name	AICc ^a	Δ AICc ^b	w ^c	k ^d	Deviance ^e
2017	1	<i>p</i> (rain + temp) <i>c</i> (sex + event)	-3611.0	0	0.29	9	52.5343
	2	<i>p</i> (temp) <i>c</i> (sex + event)	-3610.3	0.75	0.20	8	55.2941
	3	<i>p</i> (rain) <i>c</i> (sex + event)	-3609.6	1.45	0.14	8	55.9965
	4	<i>p</i> (sex + rain + temp) <i>c</i> (sex + event)	-3609.5	1.47	0.14	10	51.9948
	5	<i>p</i> (sex + temp) <i>c</i> (sex + event)	-3608.7	2.30	0.09	9	54.8364
	6	<i>p</i> (sex + rain) <i>c</i> (sex + event)	-3608.0	3.01	0.06	9	55.5500
	7	<i>p</i> (sex) <i>c</i> (sex + event)	-3607.3	3.71	0.04	8	58.2585
	8	<i>p</i> (sex + rain + temp) <i>c</i> (sex)	-3605.8	5.25	0.02	8	59.7956
	9	<i>p</i> (sex + temp) <i>c</i> (sex)	-3604.9	6.08	0.01	7	62.6372
	10	<i>p</i> (sex) <i>c</i> (sex)	-3603.5	7.49	0.01	6	66.0593
	11	<i>p</i> (rain + temp) <i>c</i> (time)	-3484.0	126.98	0.00	8	181.5340
	12	<i>p</i> (.) <i>c</i> (.)	-3479.8	131.18	0.00	4	193.7670

773

774 ^a **AICc** = Akaike's Information Criteria adjusted for small sample sizes

775 ^b Δ **AICc** = difference in the AICc values between top model and the current model

776 ^c **w** = AIC weight

777 ^d **k** = number of estimated parameters

778 ^e **Deviance** = difference between the saturated model and the current model

779

780 **Table S2.** Year-specific model-selection results for capture probability structures using the POPAN
781 model of adult female and male Admirable Redbelly Toad. The letter *p*, in front of parentheses, stands for
782 detection probability, with words in parentheses indicating detection covariates. ‘sex’ denotes differences
783 between males and females, ‘rain’ for rainfall accumulated in seven days before sampling, and ‘temp’ for
784 the median temperature of the first sampling day. All models contain the structures $\phi(\text{sex}) b(\text{sex} + \text{event})$,
785 as explained in the methods section.

Year	#	Model name	AICc ^a	ΔAICc^b	w^c	k^d	Deviance ^e
2011	1	$p(\text{sex} + \text{temp})$	631.0	0	0.43	11	-1205.44
	2	$p(\text{sex} + \text{rain} + \text{temp})$	631.6	0.53	0.33	12	-1206.98
	3	$p(\text{sex} + \text{rain})$	632.2	1.17	0.24	11	-1204.26
	4	$p(\text{sex})$	639.2	8.20	0.00	10	-1195.16
	5	$p(\cdot)$	643.3	12.30	0.00	9	-1188.99
	6	$p(\text{rain} + \text{temp})$	644.2	13.14	0.00	11	-1192.30
	7	$p(\text{temp})$	645.1	14.09	0.00	10	-1189.31
	8	$p(\text{rain})$	645.3	14.24	0.00	10	-1189.12
2013	1	$p(\text{sex} + \text{temp})$	272.0	0	0.49	10	-541.75
	2	$p(\text{temp})$	273.7	1.64	0.21	9	-538.04
	3	$p(\text{sex} + \text{rain} + \text{temp})$	274.1	2.05	0.17	11	-541.79
	4	$p(\text{rain} + \text{temp})$	275.6	3.57	0.08	10	-538.18
	5	$p(\text{sex} + \text{rain})$	277.4	5.31	0.03	10	-536.44
	6	$p(\text{rain})$	280.1	8.08	0.00	9	-531.59
	7	$p(\cdot)$	585.2	313.15	0.00	8	-224.45
	8	$p(\text{sex})$	601.8	329.75	0.00	9	-209.93
2016	1	$p(\text{rain} + \text{temp})$	1102.5	0	0.72	11	-1363.43
	2	$p(\text{sex} + \text{rain} + \text{temp})$	1104.4	1.91	0.28	12	-1363.58
	3	$p(\text{sex} + \text{temp})$	1146.1	43.64	0.00	11	-1319.78
	4	$p(\text{temp})$	1153.5	51.01	0.00	10	-1310.36
	5	$p(\text{sex})$	1202.2	99.75	0.00	10	-1261.63
	6	$p(\text{rain})$	1229.6	127.14	0.00	10	-1234.23
	7	$p(\text{sex} + \text{rain})$	1230.7	128.20	0.00	11	-1235.23
	8	$p(\cdot)$	1236.6	134.13	0.00	9	-1225.19
2017	1	$p(\text{sex})$	1697.3	0	0.27	10	-1769.86
	2	$p(\text{sex} + \text{rain} + \text{temp})$	1697.5	0.19	0.24	12	-1773.74
	3	$p(\text{sex} + \text{temp})$	1697.7	0.43	0.22	11	-1771.46
	4	$p(\text{sex} + \text{rain})$	1697.9	0.55	0.20	11	-1771.34
	5	$p(\text{rain} + \text{temp})$	1701.5	4.15	0.03	11	-1767.74
	6	$p(\text{temp})$	1702.8	5.54	0.02	10	-1764.32
	7	$p(\text{rain})$	1704.4	7.05	0.00	10	-1762.80
	8	$p(\cdot)$	1705.9	8.58	0.00	9	-1759.24

786 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes

787 ^b ΔAICc = difference in the AICc values between top model and the current model

788 ^c w = AIC weight

789 ^d k = number of estimated parameters

790 ^e Deviance = difference between the saturated model and the current model

791 **Table S3.** Biological process model-selection results for the POPAN analysis 2011. Model name letters
792 in front of parentheses stand for apparent survival (ϕ), detection probability (p), and the probability of
793 entrance (b). Words in parentheses indicate covariates for each parameter: ‘sex’ denotes differences
794 between males and females, ‘rain’ for accumulated rainfall for the last seven days before the sampling
795 day, ‘temp’ for the median temperature of the first sampling day, and ‘event’ for temporal variation
796 among sampling events. We omit specification of the parameter N , derived from ϕ and b for males and
797 females.

#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
1	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{temp})$	631.0	0	0.30	11	-1205.4
2	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	631.6	0.53	0.23	12	-1207.0
3	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain})$	632.2	1.17	0.17	11	-1204.3
4	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	634.4	3.34	0.06	11	-1202.1
5	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain})$	634.6	3.58	0.05	10	-1199.8
6	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{temp})$	634.9	3.94	0.04	10	-1199.4
7	$\phi(.) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	635.0	3.99	0.04	10	-1199.4
8	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{rain})$	635.1	4.08	0.04	10	-1199.3
9	$\phi(.) b(\text{event}) p(\text{sex} + \text{rain})$	635.4	4.39	0.03	9	-1196.9
10	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	636.7	5.62	0.02	11	-1199.8
11	$\phi(.) b(\text{event}) p(\text{sex} + \text{temp})$	640.7	9.63	0.00	9	-1191.7
12	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{temp})$	642.7	11.62	0.00	10	-1191.7
13	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{rain} + \text{temp})$	644.2	13.14	0.00	11	-1192.3
14	$\phi(.) b(.) p(\text{sex} + \text{rain} + \text{temp})$	644.7	13.64	0.00	8	-1185.6
15	$\phi(.) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	646.0	14.97	0.00	9	-1186.3
16	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	649.2	18.20	0.00	10	-1185.2
17	$\phi(\text{sex}) b(.) p(\text{sex} + \text{rain} + \text{temp})$	652.4	21.36	0.00	9	-1179.9
18	$\phi(.) b(.) p(\text{sex} + \text{temp})$	652.7	21.71	0.00	7	-1175.5
19	$\phi(.) b(\text{sex}) p(\text{sex} + \text{temp})$	653.2	22.18	0.00	8	-1177.0
20	$\phi(\text{sex}) b(.) p(\text{sex} + \text{temp})$	653.6	22.56	0.00	8	-1176.7
21	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{temp})$	654.9	23.91	0.00	9	-1177.4
22	$\phi(.) b(.) p(\text{sex} + \text{rain})$	656.5	25.50	0.00	7	-1171.7
23	$\phi(\text{sex}) b(.) p(\text{sex} + \text{rain})$	656.8	25.76	0.00	8	-1173.5
24	$\phi(.) b(\text{sex}) p(\text{sex} + \text{rain})$	658.5	27.49	0.00	8	-1171.7

798 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes
799 ^b Δ AICc = difference in the AICc values between top model and the current model
800 ^c w = AIC weight
801 ^d k = number of estimated parameters
802 ^e Deviance = difference between the saturated model and the current model
803

804 **Table S4.** Biological process model-selection results for the POPAN analysis 2013. Model name letters
805 in front of parentheses stand for apparent survival (ϕ), detection probability (p), and the probability of
806 entrance (b). Words in parentheses indicate covariates for each parameter: ‘sex’ denotes differences
807 between males and females, ‘rain’ for accumulated rainfall for the last seven days before the sampling
808 day, ‘temp’ for the median temperature of the first sampling day, and ‘event’ for temporal variation
809 among sampling events. We omit the specification of the parameter N , which is derived from ϕ and b .

#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
1	$\phi(.) b(\text{event}) p(\text{sex} + \text{temp})$	268.4	0.00	0.18	8	-541.2
2	$\phi(.) b(.) p(\text{sex} + \text{rain} + \text{temp})$	268.9	0.50	0.14	8	-540.7
3	$\phi(.) b(.) p(\text{sex} + \text{temp})$	270.2	1.74	0.08	7	-537.4
4	$\phi(.) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	270.3	1.84	0.07	9	-541.5
5	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{temp})$	270.3	1.87	0.07	9	-541.4
6	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{temp})$	270.5	2.07	0.06	9	-541.2
7	$\phi(\text{sex}) b(.) p(\text{sex} + \text{rain} + \text{temp})$	270.7	2.30	0.06	9	-541.0
8	$\phi(.) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	270.7	2.33	0.06	9	-541.0
9	$\phi(.) b(\text{sex}) p(\text{sex} + \text{temp})$	271.4	3.02	0.04	8	-538.2
10	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	272.0	3.62	0.03	10	-541.8
11	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{temp})$	272.1	3.64	0.03	10	-541.8
12	$\phi(\text{sex}) b(.) p(\text{sex} + \text{temp})$	272.1	3.72	0.03	8	-537.5
13	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	272.3	3.88	0.03	10	-541.5
14	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	272.8	4.34	0.02	10	-541.1
15	$\phi(\text{sex}) b(\text{event}) p(\text{temp})$	273.0	4.56	0.02	8	-536.7
16	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{temp})$	273.1	4.73	0.02	9	-538.6
17	$\phi(.) b(\text{event}) p(\text{temp})$	273.2	4.81	0.02	7	-534.4
18	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{temp})$	273.7	5.28	0.01	9	-538.0
19	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	274.1	5.68	0.01	11	-541.8
20	$\phi(\text{sex}) b(.) p(\text{temp})$	274.4	5.94	0.01	7	-533.2
21	$\phi(.) b(.) p(\text{temp})$	274.9	6.53	0.01	6	-530.6
22	$\phi(\text{sex}) b(\text{sex}) p(\text{temp})$	275.6	7.16	0.01	8	-534.1

810 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes
811 ^b Δ AICc = difference in the AICc values between top model and the current model
812 ^c w = AIC weight
813 ^d k = number of estimated parameters
814 ^e Deviance = difference between the saturated model and the current model
815

816 **Table S5.** Biological process model-selection results for the POPAN analysis 2016. Model name letters
817 in front of parentheses stand for apparent survival (ϕ), detection probability (p), and the probability of
818 entrance (b). Words in parentheses indicate covariates for each parameter: ‘sex’ denotes differences
819 between males and females, ‘rain’ for accumulated rainfall for the last seven days before the sampling
820 day, ‘temp’ for the median temperature of the first sampling day, and ‘event’ for temporal variation
821 among sampling events. We omit the specification of the parameter N , which is derived from ϕ and b .

#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
1	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{rain} + \text{temp})$	1102.5	0.00	0.60	11	-1363.4
2	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1104.4	1.91	0.23	12	-1363.6
3	$\phi(\text{sex}) b(\text{sex}) p(\text{rain} + \text{temp})$	1105.6	3.17	0.12	9	-1356.2
4	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	1107.6	5.17	0.05	10	-1356.2
5	$\phi(.) b(.) p(\text{sex} + \text{rain} + \text{temp})$	1117.6	15.12	0.00	8	-1342.2
6	$\phi(.) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	1118.5	16.04	0.00	9	-1343.3
7	$\phi(\text{sex}) b(.) p(\text{sex} + \text{rain} + \text{temp})$	1119.6	17.13	0.00	9	-1342.2
8	$\phi(.) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1120.6	18.16	0.00	10	-1343.2
9	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1121.3	18.86	0.00	11	-1344.6
10	$\phi(\text{sex}) b(\text{event}) p(\text{rain} + \text{temp})$	1147.3	44.84	0.00	10	-1316.5
11	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1148.5	46.03	0.00	11	-1317.4
12	$\phi(\text{sex}) b(.) p(\text{rain} + \text{temp})$	1153.9	51.39	0.00	8	-1305.9
13	$\phi(.) b(\text{sex} + \text{event}) p(\text{rain} + \text{temp})$	1166.2	63.72	0.00	10	-1297.7
14	$\phi(.) b(\text{sex}) p(\text{rain} + \text{temp})$	1170.7	68.24	0.00	8	-1289.1
15	$\phi(.) b(\text{event}) p(\text{rain} + \text{temp})$	1184.4	81.93	0.00	9	-1277.4
16	$\phi(.) b(.) p(\text{rain} + \text{temp})$	1186.9	84.47	0.00	7	-1270.8

822 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes
823 ^b Δ AICc = difference in the AICc values between top model and the current model
824 ^c w = AIC weight
825 ^d k = number of estimated parameters
826 ^e Deviance = difference between the saturated model and the current model
827

828 **Table S6.** Biological process model-selection results for the POPAN analysis 2017. Model name letters
829 in front of parentheses stand for apparent survival (ϕ), detection probability (p), and the probability of
830 entrance (b). Words in parentheses indicate covariates for each parameter: ‘sex’ denotes differences
831 between males and females, ‘rain’ for accumulated rainfall for the last seven days before the sampling
832 day, ‘temp’ for the median temperature of the first sampling day, and ‘event’ for temporal variation
833 among sampling events. We omit the specification of the derived parameter N , which is a function of sex.

#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
1	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	1696.8	0.00	0.09	10	-1770.4
2	$\phi(\text{sex}) b(\text{event}) p(\text{sex})$	1697.0	0.28	0.08	9	-1768.1
3	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex})$	1697.3	0.55	0.07	10	-1769.9
4	$\phi(\text{sex}) b(.) p(\text{sex} + \text{rain} + \text{temp})$	1697.4	0.61	0.07	9	-1767.8
5	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{temp})$	1697.4	0.61	0.07	10	-1769.8
6	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain})$	1697.4	0.67	0.07	10	-1769.7
7	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1697.5	0.74	0.06	12	-1773.7
8	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1697.6	0.81	0.06	11	-1771.6
9	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex})$	1697.7	0.89	0.06	9	-1767.5
10	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{temp})$	1697.7	0.98	0.06	11	-1771.5
11	$\phi(\text{sex}) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain})$	1697.9	1.10	0.05	11	-1771.3
12	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{temp})$	1698.0	1.19	0.05	10	-1769.2
13	$\phi(.) b(\text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1698.0	1.23	0.05	10	-1769.2
14	$\phi(\text{sex}) b(\text{event}) p(\text{sex} + \text{rain})$	1698.3	1.49	0.04	10	-1768.9
15	$\phi(.) b(\text{sex} + \text{event}) p(\text{sex} + \text{rain} + \text{temp})$	1699.4	2.65	0.03	11	-1769.8
16	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{temp})$	1699.5	2.72	0.02	9	-1765.6
17	$\phi(\text{sex}) b(.) p(\text{sex} + \text{temp})$	1699.7	2.90	0.02	8	-1763.4
18	$\phi(\text{sex}) b(.) p(\text{sex} + \text{rain})$	1701.0	4.25	0.01	8	-1762.1
19	$\phi(\text{sex}) b(\text{sex}) p(\text{sex} + \text{rain})$	1701.1	4.38	0.01	9	-1764.0
20	$\phi(\text{sex}) b(.) p(\text{sex})$	1703.4	6.64	0.00	7	-1757.7
21	$\phi(\text{sex}) b(\text{sex}) p(\text{sex})$	1704.6	7.83	0.00	8	-1758.5
22	$\phi(.) b(\text{event}) p(\text{sex} + \text{temp})$	1706.8	10.04	0.00	9	-1758.3
23	$\phi(.) b(\text{event}) p(\text{sex} + \text{rain})$	1709.5	12.74	0.00	9	-1755.6
24	$\phi(.) b(\text{sex}) p(\text{sex} + \text{temp})$	1710.9	14.12	0.00	8	-1752.2
25	$\phi(.) b(\text{sex}) p(\text{sex})$	1711.2	14.47	0.00	7	-1749.8
26	$\phi(.) b(\text{sex}) p(\text{sex} + \text{rain})$	1711.4	14.59	0.00	8	-1751.7
27	$\phi(.) b(\text{sex}) p(\text{sex} + \text{rain} + \text{temp})$	1711.6	14.84	0.00	9	-1753.5
28	$\phi(.) b(\text{event}) p(\text{sex})$	1712.6	15.85	0.00	8	-1750.5
29	$\phi(.) b(.) p(\text{sex} + \text{temp})$	1718.6	21.81	0.00	7	-1742.5
30	$\phi(.) b(.) p(\text{sex} + \text{rain})$	1719.1	22.30	0.00	7	-1742.0
31	$\phi(.) b(.) p(\text{sex})$	1719.2	22.43	0.00	6	-1739.9
32	$\phi(.) b(.) p(\text{sex} + \text{rain} + \text{temp})$	1719.4	22.63	0.00	8	-1743.7

834 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes

835 ^b Δ AICc = difference in the AICc values between top model and the current model

836 ^c w = AIC weight

837 ^d k = number of estimated parameters

838 ^e Deviance = difference between the saturated model and the current model

839

840 **Table S7.** Sampling process model-selection results for the Pollock Robust Design analysis. Model name
841 letters in front of parentheses stand for first capture (p) and recapture probability (c). Words in
842 parentheses indicate covariates for each parameter, with ‘sex’ standing for the sex and ‘rain’ for rainfall
843 accumulated in seven days before sampling. The words ‘temp’ and ‘temp2’ stand, respectively, for linear
844 and quadratic effects of the median temperature of the first sampling day. The word ‘effort’ is the number
845 of trapping days in the sampling event. For simplicity, we omit specification of the biological component
846 of the model, which was fixed as $\phi(\text{sex} + \text{year}) \gamma''(\text{sex} + \text{year}) \gamma'(\text{sex} + \text{year})$. We also omit the model
847 structure for parameter f_0 , fixed as $f_0(\text{sex} * \text{year})$, as explained in the methods section.

#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
1	$p(\text{effort} + \text{rain} + \text{temp2} + \text{sex}) c(\text{sex})$	-13427.4	0	0.99	37	-2398.27
2	$p(\text{effort} + \text{temp2} + \text{sex}) c(\text{sex})$	-13418.4	8.98	0.01	36	-2387.24
3	$p(\text{effort} + \text{rain} + \text{temp} + \text{sex}) c(\text{sex})$	-13415.3	12.15	0.00	36	-2384.08
4	$p(\text{effort} + \text{rain} + \text{temp2}) c(\text{sex})$	-13414.4	13.07	0.00	36	-2383.16
5	$p(\text{effort} + \text{rain} * \text{temp} + \text{sex}) c(\text{sex})$	-13413.3	14.14	0.00	37	-2384.12
6	$p(\text{effort} + \text{temp} + \text{sex}) c(\text{sex})$	-13.411.1	16.36	0.00	35	-2377.83
7	$p(\text{effort} + \text{rain} + \text{temp}) c(\text{sex})$	-13405.2	22.21	0.00	35	-2371.98
8	$p(\text{effort} + \text{temp2}) c(\text{sex})$	-13403.8	23.65	0.00	35	-2370.54
9	$p(\text{effort} + \text{rain} * \text{temp}) c(\text{sex})$	-13403.3	24.11	0.00	36	-2372.12
10	$p(\text{effort} + \text{temp}) c(\text{sex})$	-13399.3	28.11	0.00	34	-2364.04
11	$p(\text{effort} + \text{rain} + \text{sex}) c(\text{sex})$	-13362.4	65.08	0.00	35	-2329.11
12	$p(\text{effort} + \text{rain}) c(\text{sex})$	-13340.0	87.45	0.00	34	-2304.70
13	$p(\text{rain} + \text{temp2}) c(\text{sex})$	-12850.0	577.41	0.00	35	-1816.78
14	$p(\text{rain} + \text{temp2} + \text{sex}) c(\text{sex})$	-12849.1	578.37	0.00	36	-1817.86
15	$p(\text{rain} * \text{temp} + \text{sex}) c(\text{sex})$	-12803.2	624.27	0.00	36	-1771.95
16	$p(\text{rain} * \text{temp}) c(\text{sex})$	-12802.8	624.60	0.00	35	-1769.59
17	$p(\text{rain} + \text{temp}) c(\text{sex})$	-12782.4	645.08	0.00	34	-1747.07
18	$p(\text{rain} + \text{temp} + \text{sex}) c(\text{sex})$	-12781.8	645.65	0.00	35	-1748.54
19	$p(\text{rain} + \text{sex}) c(\text{sex})$	-12776.7	650.76	0.00	34	-1741.39
20	$p(\text{temp2} + \text{sex}) c(\text{sex})$	-12758.6	668.81	0.00	35	-1725.38
21	$p(\text{temp} + \text{sex}) c(\text{sex})$	-12452.6	974.82	0.00	34	-1417.33
22	$p(\text{sex}) c(\text{sex})$	-12409.9	1017.5	0.00	33	-1372.57
23	$p(.) c(\text{sex})$	-12409.4	1018.0	0.00	32	-1370.00
24	$p(\text{sex}) c(.)$	-12207.3	1220.1	0.00	32	-1167.93
25	$p(.) c(.)$	-12206.8	1220.7	0.00	31	-1165.37

848 ^a AICc = Akaike’s Information Criteria adjusted for small sample sizes
849 ^b Δ AICc = difference in the AICc values between top model and the current model
850 ^c w = AIC weight
851 ^d k = number of estimated parameters
852 ^e Deviance = difference between the saturated model and the current model
853

854 **Table S8.** Biological process model-selection results for the Pollock Robust Design analysis. Model
855 name letters in front of parentheses stand for survival probability (ϕ) and the probability of temporary
856 emigration (γ). Models where γ has no prime(s) treat temporary emigration as a random process
857 independent from the migration state of individuals in the previous year. The remaining models portray
858 temporary emigration as a Markov process where the state of an individual at time t depends on its state
859 at time $t-1$, as given by the probability γ' that an individual stays out of the study area in primary
860 occasion t , given that it was already out in $t-1$; and the probability γ'' that an individual leaves the study
861 area in primary occasion t , given that it was present there in $t-1$. Words in parentheses indicate covariates
862 for each parameter: 'sex' stands for the sex, and 'year' for temporal variation among years. For
863 simplicity, we omit specification of the detection component of the model, which was fixed as $p(\text{sex} +$
864 $\text{rain} + \text{temp2} + \text{effort}) c(\text{sex}) f0(\text{sex} * \text{year})$, as explained in the methods section.

#	Model name	AICc ^a	Δ AICc ^b	w^c	k^d	Deviance ^e
1	$\phi(\text{year}) \gamma(\text{sex} + \text{year})$	-13435.7	0.00	0.66	31	-2394.34
2	$\phi(\text{sex} + \text{year}) \gamma(\text{sex} + \text{year})$	-13434.3	1.43	0.32	32	-2394.95
3	$\phi(\text{year}) \gamma''(\text{sex} + \text{year}) \gamma'(\text{sex} + \text{year})$	-13428.7	6.99	0.02	36	-2397.54
4	$\phi(\text{sex} + \text{year}) \gamma''(\text{sex} + \text{year}) \gamma'(\text{sex} + \text{year})$	-13427.4	8.30	0.01	37	-2398.27
5	$\phi(\text{year}) \gamma(\text{year})$	-13424.4	11.34	0.00	30	-2380.97
6	$\phi(.) \gamma(\text{sex} + \text{year})$	-13423.6	12.11	0.00	27	-2374.11
7	$\phi(\text{sex} + \text{year}) \gamma(\text{year})$	-13423.5	12.19	0.00	31	-2382.16
8	$\phi(\text{sex}) \gamma(\text{sex} + \text{year})$	-13421.9	13.82	0.00	28	-2374.43
9	$\phi(.) \gamma''(\text{sex} + \text{year}) \gamma'(\text{sex} + \text{year})$	-13420.4	15.29	0.00	32	-2381.09
10	$\phi(\text{sex}) \gamma''(\text{sex} + \text{year}) \gamma'(\text{sex} + \text{year})$	13418.8	16.89	0.00	33	-2381.52
11	$\phi(\text{year}) \gamma''(\text{year}) \gamma'(\text{year})$	-13417.3	18.38	0.00	34	-2382.07
12	$\phi(\text{sex} + \text{year}) \gamma''(\text{year}) \gamma'(\text{year})$	-13416.7	19.08	0.00	35	-2383.41
13	$\phi(.) \gamma(\text{year})$	-13411.4	24.37	0.00	26	-2359.82
14	$\phi(\text{sex}) \gamma(\text{year})$	-13410.3	25.42	0.00	27	-2360.80
15	$\phi(.) \gamma''(\text{year}) \gamma'(\text{year})$	-13406.7	29.07	0.00	30	-2363.24
16	$\phi(\text{sex}) \gamma''(\text{year}) \gamma'(\text{year})$	-13405.4	30.29	0.00	31	-2364.05
17	$\phi(\text{year}) \gamma(\text{sex})$	-13392.7	43.03	0.00	27	-2343.19
18	$\phi(\text{sex} + \text{year}) \gamma(\text{sex})$	-13392.1	43.65	0.00	28	-2344.60
19	$\phi(\text{year}) \gamma''(\text{sex}) \gamma'(\text{sex})$	-13391.3	44.45	0.00	29	-2345.83
20	$\phi(\text{sex} + \text{year}) \gamma''(\text{sex}) \gamma'(\text{sex})$	-13389.5	46.28	0.00	30	-2346.04
21	$\phi(\text{sex}) \gamma(\text{sex})$	-13349.2	86.57	0.00	24	-2293.57
22	$\phi(.) \gamma(\text{sex})$	-13348.2	87.55	0.00	23	-2290.56
23	$\phi(.) \gamma''(\text{sex}) \gamma'(\text{sex})$	-13346.3	89.45	0.00	25	-2292.72
24	$\phi(\text{sex}) \gamma''(\text{sex}) \gamma'(\text{sex})$	-13345.5	90.25	0.00	26	-2293.94

865 ^a AICc = Akaike's Information Criteria adjusted for small sample sizes
866 ^b Δ AICc = difference in the AICc values between top model and the current model
867 ^c w = AIC weight
868 ^d k = number of estimated parameters
869 ^e Deviance = difference between the saturated model and the current model

870

871 **Table S9.** Model-averaged estimates \pm SE of capture probability for Closed, POPAN, and PRD models.
872 Each row corresponds to one breeding event identified by month and year. The letter 'F' indicates
873 females and 'M' males.
874

Month/ Year	Closed		POPAN		Robust Design	
	F	M	F	M	F	M
Oct/2010	-	-	-	-	0.02 \pm 0.003	0.05 \pm 0.007
Nov/2010	-	-	-	-	0.04 \pm 0.004	0.10 \pm 0.007
Jul/2011	0.20 \pm 0.024	0.20 \pm 0.020	0.08 \pm 0.080	0.37 \pm 0.164	0.08 \pm 0.008	0.19 \pm 0.016
Aug/2011	0.50 \pm 0.046	0.50 \pm 0.041	0.06 \pm 0.028	0.36 \pm 0.061	0.09 \pm 0.009	0.22 \pm 0.016
Sep/2011	0.46 \pm 0.078	0.46 \pm 0.076	0.03 \pm 0.018	0.20 \pm 0.042	0.05 \pm 0.005	0.14 \pm 0.009
Oct/2011	0.58 \pm 0.081	0.53 \pm 0.079	0.03 \pm 0.018	0.21 \pm 0.056	0.03 \pm 0.003	0.09 \pm 0.006
Aug/2013	0.13 \pm 0.026	0.14 \pm 0.020	0.12 \pm 0.222	0.24 \pm 0.219	0.06 \pm 0.005	0.15 \pm 0.009
Oct/2013	0.77 \pm 0.067	0.78 \pm 0.058	0.22 \pm 0.136	0.48 \pm 0.135	0.35 \pm 0.039	0.60 \pm 0.038
Dec/2013	0.13 \pm 0.026	0.14 \pm 0.020	0.01 \pm 0.008	0.04 \pm 0.012	0.04 \pm 0.006	0.10 \pm 0.013
Jul/2014	-	-	-	-	0.02 \pm 0.003	0.07 \pm 0.006
Sep/2014	-	-	-	-	0.40 \pm 0.040	0.66 \pm 0.034
Sep/2016	0.18 \pm 0.051	0.35 \pm 0.029	0.25 \pm 0.097	0.27 \pm 0.018	0.14 \pm 0.013	0.32 \pm 0.017
Oct/2016	0.37 \pm 0.087	0.60 \pm 0.040	0.44 \pm 0.141	0.47 \pm 0.029	0.08 \pm 0.007	0.20 \pm 0.011
Nov/2016	0.21 \pm 0.057	0.40 \pm 0.030	0.30 \pm 0.110	0.32 \pm 0.020	0.08 \pm 0.007	0.19 \pm 0.011
Dec/2016	0.09 \pm 0.040	0.20 \pm 0.050	0.07 \pm 0.033	0.08 \pm 0.014	0.03 \pm 0.003	0.09 \pm 0.007
Aug/2017	0.32 \pm 0.032	0.33 \pm 0.023	0.22 \pm 0.090	0.51 \pm 0.131	0.10 \pm 0.009	0.24 \pm 0.014
Sep/2017	0.38 \pm 0.055	0.39 \pm 0.049	0.21 \pm 0.073	0.50 \pm 0.037	0.09 \pm 0.008	0.22 \pm 0.013
Oct/2017	0.39 \pm 0.054	0.40 \pm 0.047	0.20 \pm 0.083	0.49 \pm 0.032	0.09 \pm 0.011	0.22 \pm 0.021
Nov/2017	0.39 \pm 0.075	0.40 \pm 0.070	0.21 \pm 0.081	0.50 \pm 0.041	0.08 \pm 0.007	0.20 \pm 0.012

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Table S10. Model-averaged estimates \pm SE of recapture probabilities for Closed and PRD models. Each row corresponds to one breeding event identified by month and year. The letter 'F' indicates females and 'M' males. Estimates of recapture probability under our PRD model were considered as constant.

Month/Year	Closed		Robust Design	
	F	M	F	M
Nov/2010	-	-		
Aug/2011	0.05 \pm 0.025	0.27 \pm 0.047		
Sep/2011	0.02 \pm 0.012	0.14 \pm 0.021		
Oct/2011	0.01 \pm 0.007	0.08 \pm 0.014		
Oct/2013	0.15 \pm 0.072	0.38 \pm 0.061		
Dec/2013	0.01 \pm 0.006	0.04 \pm 0.010		
Sep/2014	-	-	0.06 \pm 0.007	0.24 \pm 0.007
Oct/2016	0.08 \pm 0.023	0.38 \pm 0.035		
Nov/2016	0.05 \pm 0.014	0.27 \pm 0.022		
Dec/2016	0.01 \pm 0.003	0.06 \pm 0.011		
Sep/2017	0.14 \pm 0.024	0.48 \pm 0.036		
Oct/2017	0.11 \pm 0.018	0.42 \pm 0.024		
Nov/2017	0.10 \pm 0.016	0.37 \pm 0.023		

880

881 **Table S11.** Model-averaged estimates \pm SE of apparent survival and entrance probabilities for POPAN
 882 models. Each row corresponds to one interval of the breeding event identified by month and year. The
 883 letter 'F' indicates females and 'M' males. Estimates of apparent survival probability do not vary with
 884 time. Asterisks indicate uninterpretable estimates, with confidence intervals between zero and one.

Year	Apparent Survival		Probability of Entrance		
	F	M	F	M	
					886
					887
					888
Jul/2011			0.16 \pm 0.224	0.41 \pm 0.118	889
Aug/2011	0.89 \pm 0.143	0.63 \pm 0.105	0.07 \pm 0.128	0.26 \pm 0.093	
Sep/2011			*	*	890
Aug/2013			0.054 \pm 0.243	0.55 \pm 0.233	891
Oct/2013	0.90 \pm 0.147	0.92 \pm 0.114	0.09 \pm 0.183	0.10 \pm 0.190	
Sep/2016			0.19 \pm 0.068	*	892
Oct/2016	0.16 \pm 0.107	0.83 \pm 0.041	0.13 \pm 0.045	*	893
Nov/2016			0.34 \pm 0.107	*	
Aug/2017			0.15 \pm 0.161	0.024 \pm 0.138	894
Sep/2017	0.71 \pm 0.151	0.89 \pm 0.032	0.06 \pm 0.056	0.09 \pm 0.045	
Oct/2017			0.04 \pm 0.044	0.04 \pm 0.035	895

896

Capítulo 3

Assessment of threats, extinction risk and conservation priority actions for the microendemic Admirable Redbelly Toad

Este manuscrito segue o formato da revista *Perspectives in Ecology and Conservation*. A fim de facilitar a leitura, as figuras e tabelas foram inseridas no corpo do texto.

**Assessment of threats, extinction risk and conservation priority actions for the
microendemic Admirable Redbelly Toad**

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1 **Abstract**

2 Red Lists are essential conservation tools, which influence conservation outputs, such as
3 scientific knowledge gain and public awareness. Carefully identifying all known and
4 potential threats to a species is important not only to assess the extinction risk of a
5 species but also to plan priority actions for its conservation. In this study, we used
6 geographical distribution metrics and global population size, combined with an
7 objective approach to rank the threats, to reassess the conservation status of the
8 microendemic *Melanophryniscus admirabilis* (Anura: Bufonidae), the Admirable
9 Redbelly Toad, under three IUCN criteria. We found a known distribution of 0.049 km²
10 and an area of occupancy (AOO) of 4 km² as a maximum distribution for *M.*
11 *admirabilis*. The most relevant threats to the species are Bd and agrochemicals from
12 tobacco and soy plantations, and the most concerning one in terms of intensity and
13 extent is the hydropower plant construction, even though it is under control. Finally, we
14 discussed priority actions for the conservation of the Admirable toad in order to reduce
15 the species' extinction risk. Although a great effort is needed to protect the toad, the
16 distribution of *M. admirabilis* is so small that only finding a new population could
17 remove the species from threatened categories.

18 **Keywords:** *Melanophryniscus admirabilis*, Critically Endangered, area of occupancy,
19 extent of occurrence, IUCN Red List criteria, ranking of threats

20 **1. Introduction**

21 A quantitative framework widely used to assess the conservation status of a
22 taxon is the IUCN Red List Categories and Criteria, which highlights those species that
23 are most likely to become extinct in a not-so-distant future (IUCN Standards and
24 Petitions Committee, 2019). This standardized approach has been periodically updated

25 and improved and is now widely applied to regional, national, and local species
26 assessments worldwide, even though it was designed for global assessments (Mace et
27 al, 2008). Red Lists are essential conservation tools, influencing conservation outputs
28 such as scientific knowledge gain and public awareness, although measuring their
29 impact on positive changes for species persistence is still a challenge (Betts et al.,
30 2020).

31 The IUCN Red List approach is based on five criteria, which refer to currently
32 available knowledge about population size, geographic distribution, trends in population
33 and distribution, and recent, current, or projected threats acting on the species (IUCN
34 Standards and Petitions Committee, 2019). Data for assessing the complete set of
35 criteria are not available for the majority of species, but the approach is flexible enough
36 to be applicable to most of the known taxa. For example, in the absence of information
37 on abundance and population trends, about 85% of threatened amphibians have their
38 assessments based exclusively on the criteria of geographic distribution (criteria B and
39 D2; IUCN, 2021). For assessing a species' extinction risk adequate data are required for
40 at least one criterion; otherwise the species is assessed as Data Deficient (DD).

41 Carefully identifying threats is important not only to assess the extinction risk of
42 a species but also to plan priority actions for its conservation. Sound planning on how to
43 mitigate threats includes monitoring and reviewing the plan, to subsequently improve its
44 implementation (CMP, 2020). Conservation Action Plans are usually focused on
45 promoting threat reduction to conservation targets (TNC, 2007). At the global level, in
46 response to the global amphibian crisis (Young et al., 2001; Stuart et al., 2004), the
47 Amphibian Conservation Action Plan - ACAP (Gascon et al., 2007; Wren et al., 2015)
48 addressed the most important threats and the priority action steps for amphibian
49 conservation (Bishop et al, 2012). At the national level in Brazil, there are tools

50 designed for prioritizing conservation actions through threat suppression or mitigation,
51 such as the National Action Plans for Conservation of Species Threatened with
52 Extinction (PAN; Instrução Normativa MMA nº 21, 2018; e.g. Baptista et al., 2019).

53 *Melanophryniscus admirabilis* (Admirable Redbelly Toad) is an example of a
54 priority species for conservation (Portaria MMA Nº 350, 2019). It is a microendemic
55 and Critically Endangered anuran that faced a high risk of extinction due to the
56 imminent construction of a hydroelectric power plant just 500 meters upstream of the
57 species' breeding site (Fonte et al., 2014). At that time, the species was not legally
58 protected by any environmental law in Brazil, and it had been categorized as Near
59 Threatened (NT) in The IUCN Red List of Threatened Species (Maneyro, 2008). This
60 assessment was questioned by local researchers, and its conservation status was
61 reassessed with the scarce information that was produced in a year (Fonte et al., 2014).
62 It was reassessed as Critically Endangered based on criteria B1ab(iii,v)+2ab(iii,v)
63 (IUCN SSC Amphibian Specialist Group, 2013). Simultaneously, a team of researchers
64 and conservationists began a long-term project to produce as much information as
65 possible about its population, habitat and threats.

66 The main objective of this study is to present all the acquired information of the
67 Admirabilis Project in the past ten years and offer a reassessment of the conservation
68 status of the Admirable Redbelly Toad. Specifically, we addressed: (1) distribution of
69 the species in terms of area of occupancy (AOO) and extent of occurrence (EOO); (2)
70 population size of the species; (3) identification, description and ranking of threats; and
71 (4) conservation status. Finally, we discuss some possible strategies to protect the
72 population of the Admirable Redbelly Toad.

73 **2. Study species and area**

74 *Melanophryniscus admirabilis* is a colorful (bright green back, black and bright
75 red belly), small (< 42 mm), poisonous (lipophilic alkaloids from diet) and explosive
76 breeding bufonid. The species occurs in a very specific environment, breeding on a
77 flattened river bank outcrop, which is surrounded by forested steep slopes in a river
78 valley (Figure 1A). Small and shallow temporary pools on the rocky outcrop are used
79 for calling, spawning, and development of tadpoles (Figure 1B-D). The species can live
80 at least 9 years and seems to have high site fidelity, particularly among adults (Abadie
81 et al, 2021 – Chapter 1).



82
83 **Figure 1.** Admirable Redbelly Toad (*Melanophryniscus admirabilis*) and its only known locality.
84 (A) Forested steep slopes in a river valley, in Southernmost Atlantic Forest; (B) colorful
85 Admirable toads breeding in a small pool; (C) a calling male; and (D) amplexus pair spawning in
86 a small pool. Photos: Pedro Peloso - DoTS Project, Michelle Abadie, Valentina Caorsi, and
87 Simone Leonardi, respectively.

88

89 The Admirable Redbelly Toad is known from a single locality on the margins of
90 the Forqueta River, in Perau de Janeiro, on the border between the municipalities of
91 Arvorezinha and Soledade, Rio Grande do Sul, Brazil (28° 51' 25.3" S; 52° 18' 12.3"

92 W). This area is located in the Southernmost Atlantic Forest biome, about 550 m asl, on
93 the southern border slopes of the Brazilian Southern Plateau. The climate of the region
94 is classified as Subtropical Humid, without a dry season and with hot summers (*Cfa*,
95 Koeppen's climate classification; Alvares et al, 2013). The species' type locality is part
96 of the largest remaining forest fragments in the region (about 500 ha), which is under
97 pressure mainly due to the expansion of soy, tobacco and eucalyptus monocultures and
98 livestock production. The only known population of the species is found in an
99 unprotected and easily accessible area. A nearby tourist facility and upstream pesticide
100 usage expose the site to direct and indirect human threats.

101 **3. Distribution**

102 The original description of *Melanophryniscus admirabilis* informed its
103 occurrence on 200 m along the Forqueta River margins (Di-Bernardo et al., 2006). To
104 improve our understanding of the distribution of *M. admirabilis* and to verify whether
105 the species represented a true microendemism or whether its range was underestimated
106 due to lack of sampling, we conducted three different approaches: active search by the
107 project team, review of environmental impact assessment studies of hydropower
108 projects in the region, and a BioBlitz.

109 *3.1 Project Team Search*

110 We searched for new occurrences of the species along the Forqueta River and
111 other rivers in the same basin, and areas where the local people reported that the species
112 could occur (Figure 2B-C). We adjusted our methodology and sampling effort, both
113 spatially and temporally, as we improved our knowledge about the Admirable Toad
114 behavior and reproduction (see Abadie et al., 2021– Chapter 1).

115 Since the region is characterized mostly by steep and inaccessible slopes, from
116 October 2010 to March 2011, we started by searching accessible areas of the river
117 margins. Before searches, we always checked if there was reproductive activity at the
118 known breeding site. In 2017, when we knew more about the species behavior, we used
119 satellite images to select and prioritize potential searching sites as riverbanks associated
120 with steep slopes of forest (slope > 45°) and rocky outcrops and concentrated surveys
121 between August and December. In the remaining years (2013 to 2016), we worked to
122 refine knowledge about the local distribution at Perau de Janeiro, the regular breeding
123 site of the species.

124 From 2010 to 2017, we conducted visual and audio surveys (Visual Encounter
125 Survey, Crump and Scott Jr., 1994; Surveys at Breeding Sites, Scott Jr. and Woodward,
126 1994) looking for adult individuals or any other evidence of the species presence, as
127 clutches of eggs or tadpoles. We mainly focused on environmental conditions similar to
128 the known reproductive site of the species (rocky river bank outcrops with nearby forest
129 remnants), as a clue for potential new areas of occurrence. We visited at least twice all
130 potential rocky river bank outcrops to reduce false-negative observations from
131 unsuccessful survey events.

132 *3.2 Review of environmental impact assessment studies of hydropower plant*
133 *projects*

134 Simultaneously to the beginning of fieldwork, we analyzed databases from pre-
135 permitting biodiversity inventories and follow-up studies conducted between 2005 and
136 2010 in other rivers in the Taquari-Antas river basin (Figure 2A; Biolaw Consultancy
137 Company, *unpublished data*). This information and 2010-2011 project team surveys
138 were used to assess the global conservation status of the species in 2013 (IUCN SSC

139 Amphibian Specialist Group, 2013) and national and regional status in 2014 (Portaria
140 MMA N° 444, 2014; Rio Grande do Sul, 2014).

141 *3.3 BioBlitz*

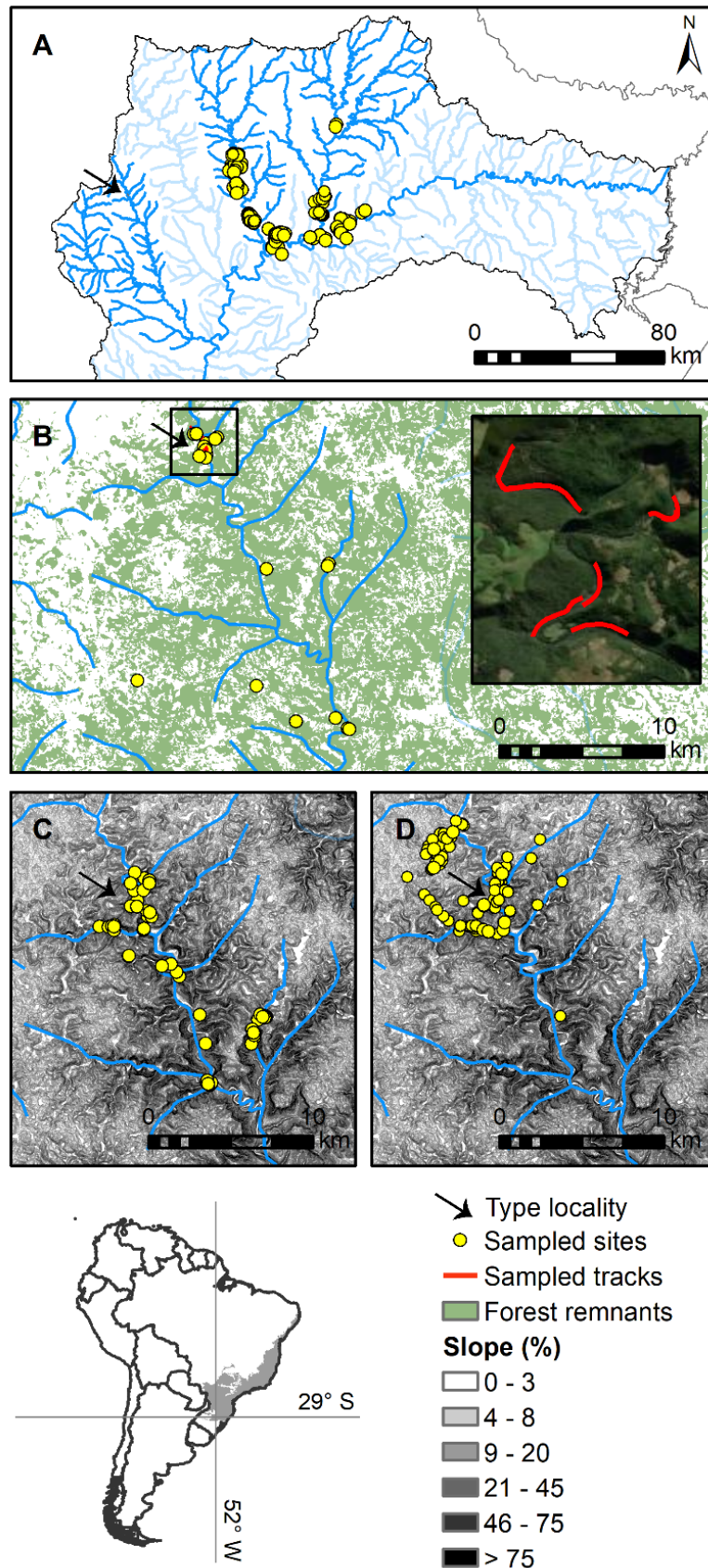
142 In 2017, we also conducted a five-day modified BioBlitz approach with the
143 involvement of taxonomic experts, a socio-environmental NGO and undergraduate
144 students. The goals of this BioBlitz were to record as many vertebrates as possible in
145 the Perau de Janeiro region and to generate data to design and propose a protected area
146 for the Admirable Redbelly Toad (Instituto Curicaca, *unpublished data*). Since the
147 Admirable toad was the target species, we also searched for adult individuals, tadpoles
148 or eggs through transects in riparian forests and along the riverbanks in the surroundings
149 of the breeding site (Figure 2D).

150 *3.4 EOO, AOO and range description and calculation*

151 We used the coordinates of the searched sites and transects to produce a
152 sampling effort map. Only confirmed records were used to calculate the extent of
153 occurrence (EOO) and the area of occupancy (AOO). The EOO was measured by a
154 minimum convex polygon (MCP), and the AOO by counting the occupied cells in a 2 x
155 2 km cell grid (IUCN Standards and Petitions Committee 2019). We also delimited the
156 range of the species, defined by “current known limits of distribution, accounting for all
157 known, inferred or projected sites of occurrence”, that is, the occupied sites and the
158 surrounding river margin forests (~50 m to each margin). Besides AOO and EOO being
159 essential to assess the conservation status under criterion B of IUCN standards (IUCN
160 Standards and Petitions Committee. 2019), distribution metrics, including the species
161 range presented here, are also useful for monitoring habitat loss, mapping threats and
162 planning and locating priority conservation actions.

163 *3.5 Admirable Toad range*

164 We searched for the species over 40 sampling days and covered 65 km along the
165 riverbanks of the Forqueta River sub-basins (Figure 2). The areas sampled by
166 herpetologist consultants (for environmental impact assessment studies) covered 16 km
167 in areas along the sub-basins of Carreiro and Turvo Rivers. We found three potential
168 new population sites, which seemed appropriate for the Admirable toad's reproduction,
169 as temporary pools in rocky outcrops. Although we have returned several times to these
170 sites, we have never found any evidence of the species. These sites are located in areas
171 lacking either adjacent forests or steep slopes. Probably as a consequence, they also lack
172 herbaceous vegetation, are exposed to the sunlight and are drier than the sites where the
173 species is usually found. This lower humidity possibly hinders the persistence of the
174 pools and reduces the minimum necessary period for tadpole development.



175

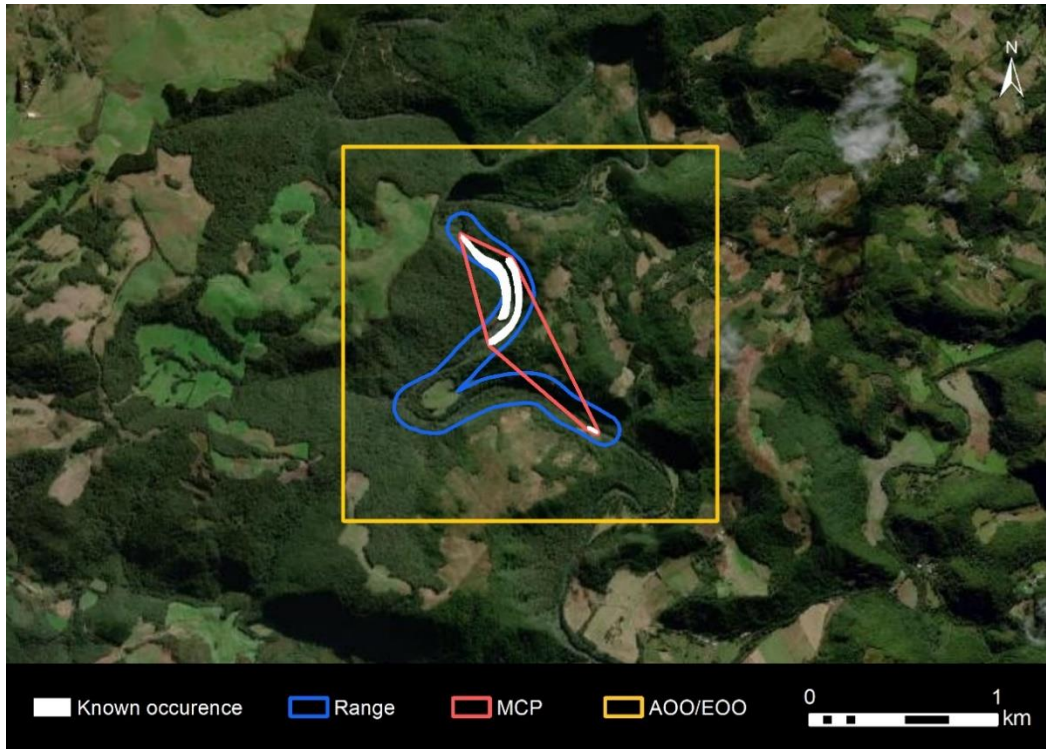
176

177

Figure 2. Areas searched for *Melanophryniscus admirabilis*. (A) sites from the review of environmental impact assessment studies of hydropower projects in Taquari-Antas Basin; (B)

178 sites from active searches by the project team in 2010-2011 and (C) 2017 ; and (D) sites from the
179 Bioblitz.

180 We found *M. admirabilis* in a unique new site, expanding its range by only 1.43
181 km downstream and 492 m in Euclidean distance from the previously known site.
182 Species presence was detected after heavy rains in September of 2017 based on auditory
183 encounters (male calling) and confirmed by the visual encounter and capture of some
184 individuals. The new site is a rocky river bank outcrop with less than 100 m², few pools,
185 and surrounding steep slope forests. This new record increases the known EOO
186 (measured by a minimum convex polygon – MCP) from 0.035 to 0.253 km². The new
187 AOO is a 4 km² square encompassing the area where the toad lives, following the novel
188 approaches applied by IUCN (Figure 3). As AOO should not exceed EOO (IUCN
189 Standards and Petitions Committee, 2019), we adjusted EOO to 4 km². The range was
190 estimated to be 0.418 km² (Figure 3). We estimated the known species occurrence to be
191 0.049 km² along the river, including both river margins and 50 m of adjacent forests
192 (see Abadie et al, 2021 – Chapter 1).



193

194 **Figure 3.** Distribution of *Melanophryniscus admirabilis* under different metrics. Range is the
 195 current known limits of distribution; MCP is the minimum convex polygon; AOO is the area of
 196 occupancy and EOO is the adjusted extent of occurrence.

197 The new record considerably expands the species distribution (almost 10 x the
 198 previous EOO; IUCN SSC Amphibian Specialist Group, 2013); however, the range is
 199 still very restricted, and we kept considering it as a single population and locality.

200 **4. Population Size**

201 For assessment purposes, we followed the IUCN Red List Categories and
 202 Criteria v. 3.1 (2012) nomenclature which defines ‘population’ as the total number of
 203 individuals of the taxon, and ‘population size’ as the number of mature individuals of
 204 the population. Since most of the mature individuals of *M. admirabilis* reproduce at the
 205 same moment in explosive breeding events, we considered the breeding adults (i.e.,
 206 participating in the reproductive events at the breeding site) as ‘mature individuals’.

207 Abadie et al. (2021 – Chapter 2) estimated the abundance of Admirable Redbelly
 208 Toad breeding adults over an eight-year window (2010-2017), using individual photo-
 209 identification and capture-recapture models (Pollock Robust Design; Pollock, 1982).
 210 The abundance estimate was obtained from the site where most of the population
 211 concentrates for breeding. This breeding site is about 400 m long and one to 14 m wide
 212 (Abadie et al., 2021 – Chapter 1). The authors provided annual estimates of abundance
 213 for males and females, which varied over time from 742 (CI= 645-838) individuals in
 214 2014 to 1451 (CI= 1280-1622) individuals in 2017 (Table 1). They also presented a
 215 male-biased sex ratio for the species, even after accounting for imperfect detection
 216 (from 1.27:1, in 2017, to 2.45:1, in 2014).

217 **Table 1.** Estimated annual population size for males and females of *Melanophryniscus*
 218 *admirabilis* and respective sex ratio, from 2010 to 2017 using capture-recapture data. Adapted
 219 from Abadie et al., 2021 – Chapter 2.

Year	Population Size		Sex Ratio
	(-95% CI – +95% CI)		
	F	M	
2010	428 (276-670)	721 (573-916)	1.68±0.14
2011	477 (378-612)	779 (695-885)	1.63±0.14
2013	307 (247-396)	521 (473-591)	1.70±0.11
2014	215 (172-280)	527 (478-596)	2.45±0.31
2016	594 (491-732)	794 (728-876)	1.33±0.13
2017	638 (528-785)	813 (745-900)	1.27±0.22

229 According to IUCN Standards and Petitions Committee (2019), when the
 230 population size fluctuates or has a biased sex ratio, we should use the lowest estimate.
 231 Although *M. admirabilis* population has a male-biased sex ratio, females attend to more

232 than one breeding event per year (Abadie et al, 2021 – Chapter 1). Thus, to assess the
233 species conservation status we used the estimate from 2014 (females and males = 742
234 individuals, CI= 645-838; Abadie et al., 2021 – Chapter 2).

235 In addition to the main breeding site where the capture-recapture study was
236 conducted, there are three other small breeding sites, which represent less than 10% of
237 the surface of the main one. Even if we extrapolate a density estimate to these sites, the
238 total population size would remain below 1,000 individuals.

239 **5. Threats**

240 The list of threats presented here is the result of 10 years of the Admirabilis
241 Project and was elaborated based on regular visits to the locality and regular
242 communication and knowledge shared with local people. The final selection is a
243 consensus list that was gathered through a series of specific meetings with the project
244 team to enumerate, describe and rank all current and potential direct threats.

245 First, we sorted all direct threats that we considered relevant for *M. admirabilis*
246 from the CMP Direct Threats Classification v 2.0 (Salafsky et al., 2008; CMP, 2016).
247 We then ranked all threats using three criteria based on the Threat Reduction
248 Assessment (TRA) approach (Salafsky and Margoluis, 1999): 1) Extent: portion of the
249 area of occupancy (AOO) that is already or will be affected by the threat; 2) Intensity:
250 degree of damage or disturbance severity caused by the threat (i.e. will the threat change
251 the overall supporting system, such as environmental conditions, potentially affecting
252 the entire population or will it affect some individuals' survival?); 3) Urgency: the
253 immediacy of the threat (i.e. is the threat occurring now or may the impact arise only in
254 future years?). We adapted the TRA criteria as a tool for ordering and prioritizing the
255 species threats and do not conduct a threat reduction assessment. The number of ranking

256 levels for the three criteria varied from one to the total number of threats, and the same
257 number could not be repeated within the same criterion. Since the species is
258 microendemic, the differences between threats of the same criterion are very small in
259 absolute terms, making it sometimes difficult to assign rankings. Thus, it was essential
260 to weigh the differences and discuss the ordering until a consensus was reached. We
261 then added up the rankings for all three criteria for each threat to obtain the total
262 ranking. Finally, we calculated the Threat Relevance Index (TRI), which is the
263 proportion of a given threat to the total sum of the three criteria:

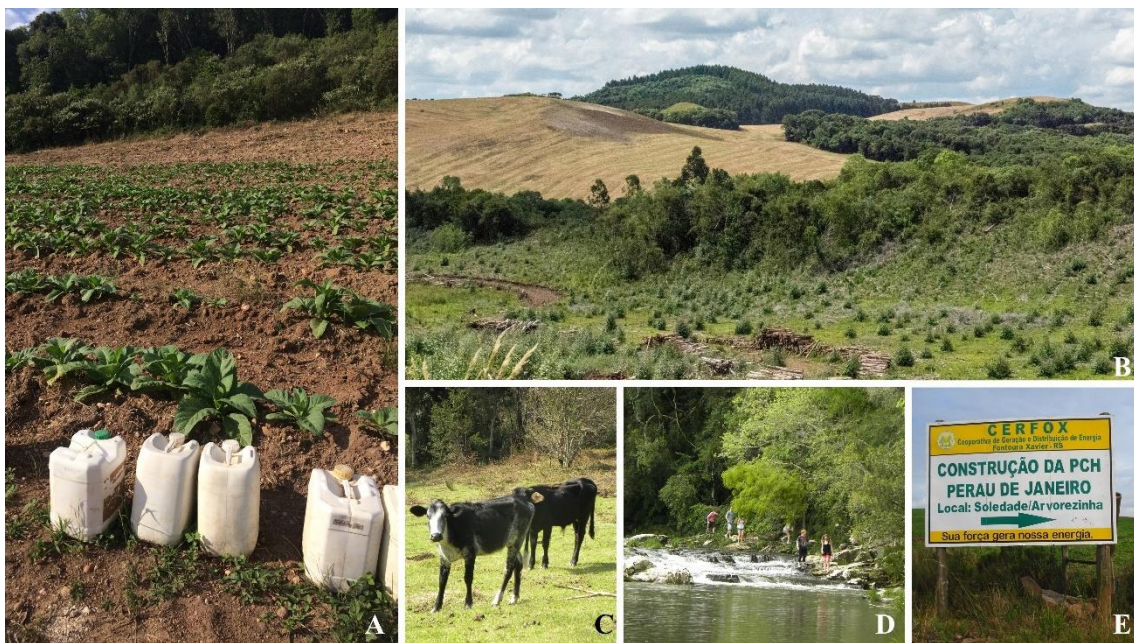
$$264 \quad TRI_i = (E_i + I_i + U_i) / (\sum_{i=1}^n E_i + \sum_{i=1}^n I_i + \sum_{i=1}^n U_i),$$

265 where TRI is the Threat Relevance Index for each threat $i \{1, \dots, n = 10\}$, E is extent, I
266 is intensity, and U is urgency.

267 We kept the threats related to Climate Change, such as large temperature
268 variations or changes in the precipitation regime, out of our classification and ranking
269 not because these are not potential threats to the species, but because we have no direct
270 influence on them.

271 We identified a total of ten direct threats to the Admirable Redbelly Toad and its
272 habitat (Table 2; Figure 4). Amphibian fungal disease (Chytridiomycosis) followed by
273 agricultural pesticides were classified as the greatest general concern. Both livestock
274 and American bullfrog (*Lithobates catesbeianus*) invasion were equally ranked as of
275 least general concern. Although cattle-raising is an expected threat to Atlantic Forest
276 (Fundación Vida Silvestre Argentina and WWF, 2017), particularly by virtue of use of
277 forests as shelter by cattle, with risk of trampling and changes in litter humidity, the
278 slope in the adjacent areas and the site itself do not favor this activity. Thus, cattle

279 presence is sporadic. The risk of a hydropower plant construction has the highest rank
280 on extension and intensity criteria. But it was considered as the lowest urgency because
281 this threat is currently controlled due to the prohibition for new HPP projects on the
282 Upper Forqueta River Basin (Becker et al., 2017). Agricultural pesticide was the most
283 urgent threat because soy production is quickly expanding and dominating the
284 landscape matrix. Although there is a general demand for poison and colorful frogs for
285 the pet trade (CITES, 2020), we do not have any record of *M. admirabilis* individuals
286 found in trade.



287
288 **Figure 4.** Identified direct threats to *Melanophryniscus admirabilis*. (A) Pesticides used in
289 tobacco plantations; (B) soy and eucalyptus plantations; (C) cattle breeding; (D) tourists at the
290 breeding site of the species; and (E) sign indicating the construction of a hydropower plant.
291 Photos: Talita M. Ribeiro (A), Pedro Peloso - DoTS Project (B), Michelle Abadie (C-E).

292 **Table 2.** *Melanophryniscus admirabilis* threats ranking according to CMP Direct Threats Classification v 2.0 levels. The ranking is in descending order within each
 293 criterion (E, I, U), where “10” is the highest level for the threats. E: Extent - portion of the area of occupancy (AOO) that is already or will be affected by the threat;
 294 I: Intensity – degree of damage or disturbance severity caused by the threat; U: Urgency - the immediacy of the threat; TRI: Threat Relevance Index of each threat
 295 (not in order).

CMP Direct Threats Classification v 2.0 levels

General type	Definition	Example for <i>M. admirabilis</i>	Definition/Justification	E	I	U	TR	TRI (%)
2. Agriculture & Aquaculture								
2.1 Annual & Perennial Non-Timber Crops	Crops planted for food, fodder, fiber, fuel, or other uses	Tobacco and soy plantations	Native vegetation areas have been converted into tobacco and soy plantations. It threatens the population of <i>M. admirabilis</i> due to the use of pesticides in the plantations (see item 9.3) and the loss of local humidity from the native forest. Together these mechanisms lead to the loss of local habitat quality	6	4	6	16	9.70
2.2 Wood & Pulp Plantations	Stands of trees planted for timber or fiber outside of natural forests, often with non-native species	Eucalyptus plantations	Native forest areas have been replaced by eucalyptus plantations (slower than conversion into tobacco and soy plantations). It threatens the population of <i>M. admirabilis</i> due to the use of pesticides in the plantations (see item 9.3) and the loss of local humidity from the native forest. Together both mechanisms lead to the loss of local habitat quality	5	3	2	10	6.06
2.3 Livestock Farming & Ranching	Domestic terrestrial animals raised in one location on farmed or non-local resources (farming); also domestic or semi-domesticated animals allowed to roam in the wild and supported by natural habitats (ranching)	Cattle breeding	Small-scale cattle breeding. It affects the population of <i>M. admirabilis</i> due to trampling on seedlings of riparian native flora, hindering the regeneration of the local vegetation and resulting in forest floor drying	3	1	5	9	5.45
5. Biological Resource Use								
5.1 Hunting & Collecting Terrestrial Animals	Killing or trapping terrestrial wild animals or animal products for commercial, recreation, subsistence, research or cultural purposes, or for control/persecution reasons; includes accidental mortality/bycatch	Collection of specimens for the pet trade	Sporadic capture of specimens from the wild. Potential use of the species in the pet trade due to its colorful pattern and easy access to the locality. It affects the population of <i>M. admirabilis</i> due to the removal of mature individuals from the population	1	8	4	13	7.88

General type	Definition	Example for <i>M. admirabilis</i>	Definition/Justification	A	I	U	TR	TRI (%)
5.3 Logging & Wood Harvesting	Harvesting trees and other woody vegetation for timber, fiber, or fuel, including site preparation and other forestry management practices	Harvesting trees	Tree harvesting for firewood, which is used in the tobacco drying process. It threatens the population of <i>M. admirabilis</i> due to the loss of local humidity from the native forest, leading to the loss of local habitat quality	7	6	8	21	12.73
6. Human Intrusions & Disturbance								
6.1 Recreational Activities	People spending time in nature or traveling in vehicles outside of established transport corridors, usually for recreational reasons	Tourism	Local people and tourists walking across the reproductive site of the species. It affects the population of <i>M. admirabilis</i> due to the trampling of individuals and pools used for reproduction	2	7	7	16	9.70
7. Natural System Modifications								
7.2 Dams & Water Management / Use	Changing water flow patterns from their natural range of variation either deliberately or as a result of other activities	Risk of dam construction	Possible construction of hydropower plants for energy production. Dam construction would affect the <i>M. admirabilis</i> population due to changes in the hydrologic regime and quality	10	10	1	21	12.73
8. Invasive & Problematic Species, Pathogens & Genes								
8.1 Invasive Non-Native / Alien Plants & Animals	Harmful plants and animals not originally found within the ecosystem(s) in question and directly or indirectly introduced and spread into it by human activities	American Bullfrog (<i>Lithobates catesbeianus</i>)	Presence of the American Bullfrog (<i>Lithobates catesbeianus</i>). It can affect the <i>M. admirabilis</i> population due to its potential as a vector of diseases	4	2	3	9	5.45
8.4 Pathogens & Microbes	Harmful native and non-native agents that cause disease or illness to a host, including bacteria, viruses, prions, fungi, and other microorganisms	Chytridiomycosis	Presence of the fungus <i>Batrachochytrium dendrobatidis</i> (Bd) in specimens and/or in the environment. It can affect the population of <i>M. admirabilis</i> due to the development of the chytridiomycosis disease	8	9	9	26	15.76
9. Pollution								
9.3 Agricultural & Forestry Effluents	Water-borne pollutants from agricultural, silvicultural and aquaculture systems that include nutrients, toxic chemicals and/or sediments including the effects of these pollutants on the site where they are applied	Agricultural pesticides	Use of pesticides in tobacco, soy and eucalyptus plantations on the upper slope of the Forqueta River and upstream. It affects the population of <i>M. admirabilis</i> due to the flow of residues from the plantations to the reproductive site of the species, which can cause malformations or even the direct death of individuals	9	5	10	24	14.55
TOTAL				55	55	55	165	100

299 **6. Conservation Status Reassessment**

300 According to the Guidelines for Using the IUCN Red List Categories and
301 Criteria (2019), it is important to use the best available data to assess taxa status against
302 all five criteria (IUCN Standards and Petitions Committee, 2019). Based on the
303 information gathered in this study, we could assess the extinction risk of *M. admirabilis*
304 under criterion B (geographic range metrics, severe fragmentation, number of locations,
305 decline or fluctuations), criterion C (small and declining population size and
306 fragmentation, fluctuations, or few subpopulations), and criterion D (very small
307 population or very restricted distribution). We were not able to assess under criteria A
308 (population size reduction) and E (quantitative analysis of extinction risk) because we
309 do not have ten years (or three generations) of population monitoring, and there is not
310 enough available information for a population viability analysis.

311 *6.1. Criterion B:* (1) The species has a measured EOO of 0.253 km², adjusted
312 to 4 km², which is within the threshold for Critically Endangered (100 km²) under
313 criterion B1; (2) The species has an estimated AOO of 4 km², which is within the
314 threshold for Critically Endangered (10 km²) under criterion B2; (3) With the observed
315 presence of the fungus *Batrachochytrium dendrobatidis* (Bd) in individuals of *M.*
316 *admirabilis* (M.R. Pontes, *personal communication*), if a Bd outbreak occurs, it could
317 cause a fast decline in the entire population (number of locations=1; subcriterion a); (4)
318 There is an observed ongoing decline of the species' terrestrial and aquatic habitat
319 quality, as a result of heavy and continued usage of agrochemicals and loss of forest to
320 agriculture and harvesting of trees for fuel (subcriterion b, item iii); (5) the locality is a
321 target area for hydroelectric power plants. Under this criterion, *M. admirabilis* would
322 remain categorized as Critically Endangered, based on criteria B1ab(iii) + 2ab(iii).

323 6.2. *Criterion C*: (1) The species has less than 2,500 mature individuals,
324 which is within the threshold for Endangered; (2) There is no observed, estimated,
325 projected, or inferred continued decline in the number of mature individuals. Therefore,
326 under this criterion, *M. admirabilis* could not be classified in any of the threat
327 categories.

328 6.3. *Criterion D*: (1) The species population size was estimated at less than
329 one thousand individuals, which is within the threshold for Vulnerable (< 1,000 mature
330 individuals) under criterion D1; (2) The species occurs in a very restricted area of
331 occupancy (AOO < 20 km²) and has less than five locations, which is within the
332 threshold for Vulnerable under criterion D2. Also, the species may be prone to “future
333 threats that could drive the taxon to CR or EX in a very short time”, such as the HPP
334 construction, Bd infection or forest loss. Under this criterion, *M. admirabilis* must be
335 categorized as Vulnerable, based on criteria D1 + 2.

336 Given the precautionary principle adopted by IUCN guidelines, which
337 recommend that the taxon should be listed under the most threatened category, we
338 propose *M. admirabilis* to remain categorized as Critically Endangered (criterion B).

339 **7. Conclusions and Future Directions**

340 For Red Lists to be useful for conservation planning and effectiveness
341 monitoring, the regular reassessment of the species conservation status is fundamental.
342 New threats can emerge and affect the priority of conservation actions, particularly for
343 species with restricted distributions. In this study, we conducted a detailed conservation
344 status reassessment of *M. admirabilis*, a microendemic species from the Southern
345 Atlantic Forest, addressing three out of five criteria from IUCN Red List Categories and
346 Criteria. We compiled information about the species distribution and population size

347 and adapted a methodology used to monitor threat reduction to rank threats by extent,
348 intensity and urgency.

349 The most concerning threats, Bd infection and agrochemicals, are of complex
350 control because they have a diffuse origin and chronic action, that is, even if these
351 threats are introduced in some local quite far upstream catchment, they could affect the
352 species and/or the quality of the species' habitat in a difficult way to control. There is
353 evidence that *M. admirabilis* has a potential xenobiotic degradation pathway in the oral
354 microbiota (Mann et al., 2021), which may be an indicator of contact with
355 contaminants, although the tadpole is quite resistant to pesticides (da Silva et al, 2021).
356 Synergistic effects among pesticides, skin microbiome and chytridiomycosis are still
357 poorly understood for amphibians (Jani and Briggs, 2018), but environmental
358 contaminants could play an important role in altering the amphibian skin microbiome
359 and therefore disease susceptibility (McCoy and Peralta, 2018). The quantification and
360 monitoring of both agrochemical concentration and Bd incidence in *M. admirabilis*
361 range are imperative to anticipate crashes in population size. Promoting best practices
362 and/or alternatives to intensive agrochemical usage among landowners in the entire
363 river basin is also urgent.

364 An important conservation action would be the creation of a protected area for
365 the Admirable Redbelly Toad, and this was formally recognized in the National Action
366 Plan for the Conservation of Amphibians and Reptiles in Southern Brazil (Portaria
367 MMA N° 350, 2019). Here, it is important to note that the area is already considered a
368 global priority for conservation, being recognized as a Key Biodiversity Area and an
369 Alliance for Zero Extinction site (Key Biodiversity Areas Partnership, 2020; AZE,
370 2020). This strategic action could protect the population from most of the prioritized
371 threats but is scale-dependent. Tree harvesting, livestock disturbance, tobacco,

372 eucalyptus and soy plantations, tourist activities and illegal collection of specimens at or
373 near the main reproductive site could be eradicated by a small-scale protected area.
374 However, agrochemical contamination effects could only be reduced with broader-scale
375 economic incentives. Tobacco plantations are monocultures dependent on neurotoxic
376 pesticides (Krawczyk et al., 2014), which besides harming flora, fauna and funga,
377 directly affect farmers (Arcury and Quandt, 2006). Also, soy plantations and their inputs
378 are expanding within the species' range. Another conservation action would be to
379 subsidize the substitution of tobacco and soy monocultures for yerba mate (cultural
380 South American tea) in agroforestry and organic systems, availing on the region's
381 vocation known as the "land of yerba mate". Linking yerba mate culture to consumption
382 of organic produce and the conservation of an endemic species in the region might be a
383 solution that benefits the entire production chain involved. Moreover, promoting
384 mitigation strategies to reduce pesticide inputs in the entire basin should be a priority
385 for government programs, since the deliberate abuse of pesticides (often used above
386 legal levels) directly impacts water quality and produces different health impacts for
387 humans and wildlife (Pimentel et al., 1992). Although difficult to put in place, the
388 replacement of monocultures by yerba mate agroforestry and the creation of a protected
389 area have the potential to reduce the threats listed above. These priority actions could
390 genuinely change the extinction risk degree of *M. admirabilis* from Critically
391 Endangered B1ab (iii) + 2ab (iii) to Vulnerable D1 + 2.

392 We considered the hydropower plant (HPP) construction as the most extensive
393 and intensive (but least urgent) threat to *M. admirabilis* (see Fonte et al., 2014).
394 Currently, the species is protected by a legal instrument that prohibits the HPP
395 construction in its habitat (Resolução Consema, N° 388, 2018). If for any reason the
396 validity of this regulation ceases, the species may be susceptible to loss of habitat

397 quality and a potential reduction in the number of mature individuals. A recent study
398 found that this kind of regulation is the main reason for HPP cancellations (Macedo,
399 2021). Therefore, the permanent avoidance of this potential threat would only be
400 ensured by either the maintenance of the referred legal instrument throughout the next
401 governments or the creation of a protected area encompassing all the potential HPP sites
402 upstream. With this perspective and if the number of mature individuals increases above
403 the one thousand individuals threshold, the species could even be categorized as Near
404 Threatened (NT). Nonetheless, this would only happen if other populations were found
405 in other areas, expanding the AOO, and if the likelihood of a plausible future threat that
406 could quickly drive the species to Critically Endangered or Extinct is removed. With the
407 current distribution, even if those conditions were achieved, the species would remain
408 categorized as Vulnerable because of criterion D2 ($AOO < 20 \text{ km}^2$).

409 Some research actions also deserve further and continuous efforts due to their
410 repercussion for conservation planning and species persistence. First, it is necessary to
411 keep monitoring the population size and habitat quality to track for any positive or
412 negative trends. Second, the search for new populations should be reinforced since their
413 finding and protection would increase the likelihood of the species' persistence. Since
414 *M. admirabilis* is a quite elusive explosive breeder, indirect detection techniques, such
415 as environmental DNA and passive acoustic recorders, could improve the chance to
416 detect the species in new areas (e.g. Lopes et al., 2020).

417 Microendemic species like the Admirable toad are a great challenge for
418 conservation. There are many examples of fast decline and disappearance in the
419 amphibian literature (see Bishop et al., 2012). The restricted range alone is an
420 underlying condition for increased extinction risk. However, it also gives us a great
421 opportunity to control most of the main mechanisms or factors that could affect the

422 species persistence since the landscape to be targeted for action is relatively small. Time
423 will be the limiting dimension for planning and implementing conservation actions
424 since any significant shift in the extent or intensity of the main threats could drive
425 important population losses. This requires that proper indicators of threats and the
426 population need to be regularly and continuously monitored, with obvious pressures on
427 sustainable project fundraising. This challenge could only be overcome with
428 increasingly close cooperation between researchers, locals, NGOs, environmental
429 agencies and other government structures and funders. In the recent history of the
430 Admirable toad conservation program, this kind of coordinated partnership was
431 fundamental for the toad's conservation (Fonte et al., 2014; and the present study).

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447 **References**

- 448 Alvares, C.A., Stape, J.L., Sentelhas, P.C., De Moraes Gonçalves, J.L., Sparovek, G.,
 449 2013. Köppen's climate classification map for Brazil. *Meteorol. Zeitschrift* 22,
 450 711–728. <https://doi.org/10.1127/0941-2948/2013/0507>
- 451 Arcury, T.A., Quandt, S.A., 2006. Health and social impacts of Tobacco production. *J.*
 452 *Agromedicine* 11, 71–81. https://doi.org/10.1300/J096v11n03_08
- 453 AZE (Alliance for Zero Extinction sites), 2020. Areas of Biodiversity Importance:
 454 summaries of globally relevant systems to identify areas of importance for
 455 biodiversity. [https://www.biodiversitya-z.org/content/alliance-for-zero-extinction-](https://www.biodiversitya-z.org/content/alliance-for-zero-extinction-sites-aze)
 456 [sites-aze](https://www.biodiversitya-z.org/content/alliance-for-zero-extinction-sites-aze)
- 457 Baptista, J.R., Giné, G.A.F., Schiavetti, A., 2019. Performance of Single-versus Multi-
 458 Species Recovery Plans in Brazil. *Environ. Conserv.* 46, 211–218.
 459 <https://doi.org/10.1017/S0376892919000134>
- 460 Becker, F.G., Pineda, M.D., Perelló, L.F.C., Pagel, S.M., Ribeiro, G.V.B., Leite, E.H.,
 461 Audibert, E.A., Guadagnin, D.I., 2017. Síntese da Avaliação Ambiental Regional
 462 na Bacia Hidrográfica do Rio Taquari-Antas para fins de Licenciamento de
 463 Empreendimentos Hidrelétricos. *Fepam em Rev.* 11, 5–25.
- 464 Betts, J., Young, R.P., Hilton-Taylor, C., Hoffmann, M., Rodríguez, J.P., Stuart, S.N.,
 465 Milner-Gulland, E.J., 2020. A framework for evaluating the impact of the IUCN
 466 Red List of threatened species. *Conserv. Biol.* 34, 632–643.
 467 <https://doi.org/10.1111/cobi.13454>
- 468 Bishop, P.J., Angulo, A., Lewis, J.P., Moore, R.D., Rabb, G.B., Garcia Moreno, J.,
 469 2012. The Amphibian Extinction Crisis - what will it take to put the action into the
 470 Amphibian Conservation Action Plan? *S.A.P.I.EN.S*[Online]. 5, 97-111.
 471 <http://sapiens.revues.org/1406>
- 472 CITES (Conventional on International Trade in Endangered Species of Wild Fauna and
 473 Flora), 2020. CITES Trade Database v. 2020.1. Available at: <https://trade.cites.org/>
 474 (accessed April 2021).
- 475 CMP (Conservation Measures Partnership), 2016. CMP Direct Threats Classification v
 476 2.0. Available in: [https://conservationstandards.org/library-item/direct-threats-](https://conservationstandards.org/library-item/direct-threats-classification-v2-0/)
 477 [classification-v2-0/](https://conservationstandards.org/library-item/direct-threats-classification-v2-0/) (accessed April 2021).

478 CMP (Conservation Measures Partnership), 2020. Open Standards for the Practice of
479 Conservation. Version 4.0. CMP, Washington, D.C.

480 Crump, M.L., Scott Jr., N.J., 1994. Standard techniques for inventory and monitoring:
481 Visual Encounter Surveys., in: Heyer, W.R., Donnelly, M.A., McDiarmid, R.W.,
482 Hayek, L.C., Foster, M.S. (Eds.), Measuring and Monitoring Biological Diversity.
483 Standard Methods for Amphibians. Smithsonian Institution Press, Washington DC,
484 pp. 84–92.

485 da Silva, P.R., Borges-Martins, M., Oliveira, G.T., 2021. *Melanophryniscus admirabilis*
486 tadpoles' responses to sulfentrazone and glyphosate-based herbicides: an approach
487 on metabolism and antioxidant defenses. Environ. Sci. Pollut. Res. 28, 4156–4172.
488 <https://doi.org/10.1007/s11356-020-10654-x>

489 Di-Bernardo, M., Maneyro, R., Grillo, H., 2006. New Species of *Melanophryniscus*
490 (Anura: Bufonidae) from Rio Grande do Sul, Southern Brazil. J. Herpetol. 40, 261–
491 266. <http://dx.doi.org/10.1670/05-008.1>

492 Fonte, L.F.M. da, Abadie, M., Mendes, T., Zank, C., Borges-Martins, M., 2014. The
493 Times they are a-Changing: How a Multi-Institutional Effort Stopped the
494 Construction of a Hydroelectric Power Plant that Threatened a Critically
495 Endangered Red-Belly Toad in Southern Brazil. FrogLog 22, 18–21.

496 Fundación Vida Silvestre Argentina and WWF, 2017. State of the Atlantic Forest:
497 Three Countries, 148 Million People, One of the Richest Forests on Earth. Puerto
498 Iguazú, Argentina.

499 Gascon, C., Collins, J.P., Moore, R.D., Church, D.R., McKay, J.E., Mendelson, J.R.,
500 2007. Amphibian Conservation Action Plan. IUCN/SSC Amphibian Specialist
501 Group, Gland, Switzerland and Cambridge, UK.

502 Instrução Normativa MMA no 21, de 18 de dezembro de 2018, 2018. Planos de Ação
503 Nacional para Conservação de Espécies Ameaçadas de Extinção. Ministério do
504 Meio Ambiente, Brasil.

505 IUCN, 2012. IUCN Red List Categories and Criteria: Version 3.1. Second edition.
506 Gland, Switzerland and Cambridge, UK: IUCN. iv + 32pp.

507 IUCN SSC Amphibian Specialist Group, 2013. *Melanophryniscus admirabilis* [WWW
508 Document]. IUCN Red List Threat. Species 2013 e.T135993A44846478. URL
509 <https://dx.doi.org/10.2305/IUCN.UK.2013-2.RLTS.T135993A44846478.en>.
510 (accessed 25 March 2021).

511 IUCN Standards and Petitions Committee, 2019. Guidelines for Using the IUCN Red
512 List Categories and Criteria 14.
513 <https://doi.org/http://www.iucnredlist.org/documents/RedListGuidelines.pdf>.

514 Jani, A.J., Briggs, C.J., 2018. Host and Aquatic Environment Shape the Amphibian
515 Skin Microbiome but Effects on Downstream Resistance to the Pathogen
516 *Batrachochytrium dendrobatidis* Are Variable. *Front. Microbiol.* 9, 487.
517 <https://doi.org/10.3389/fmicb.2018.00487>

518 Key Biodiversity Areas Partnership, 2020. Key Biodiversity Areas factsheet: Rio
519 Forqueta. Extracted from the World Database of Key Biodiversity Areas.
520 Developed by the Key Biodiversity Areas Partnership: BirdLife International,
521 IUCN, American Bird Conservancy, Amphibian Survival Alliance, Conservation
522 International, Critical Ecosystem Partnership Fund, Global Environment Facility,
523 Global Wildlife Conservation, NatureServe, Rainforest Trust, Royal Society for the
524 Protection of Birds, World Wildlife Fund and Wildlife Conservation Society.
525 Downloaded from <http://www.keybiodiversityareas.org/site/factsheet/47065>,
526 (accessed 22 April 2021).

527 Krawczyk, N., Meyer, A., Fonseca, M., Lima, J., 2014. Suicide mortality among
528 agricultural workers in a region with intensive tobacco farming and use of
529 pesticides in Brazil. *J. Occup. Environ. Med.* 56, 993–1000.
530 <https://doi.org/10.1097/JOM.0000000000000214>

531 Lopes, C.M., Baêta, D., Valentini, A., Lyra, M.L., Sabbag, A.F., Gasparini, J.L.,
532 Dejean, T., Haddad, C.F.B., Zamudio, K.R., 2020. Lost and found: Frogs in a
533 biodiversity hotspot rediscovered with environmental DNA, in: *Molecular Ecology*.
534 Blackwell Publishing Ltd. <https://doi.org/10.1111/mec.15594>

535 Mace, G.M., Collar, N.J., Gaston, K.J., Hilton-Taylor, C., Akçakaya, H.R., Leader-
536 Williams, N., Milner-Gulland, E.J., Stuart, S.N., 2008. Quantification of extinction
537 risk: IUCN’s system for classifying threatened species. *Conserv. Biol.* 22, 1424–
538 1442. <https://doi.org/10.1111/j.1523-1739.2008.01044.x>

539 Macedo, A., 2021. O licenciamento ambiental evita impactos de hidrelétricas?
540 Unpublished master’s thesis. Federal University of Rio Grande do Sul, Porto
541 Alegre, Brazil

542 Maneyro, R., 2008. *Melanophryniscus admirabilis*. The IUCN Red List of Threatened
543 Species 2008: e.T135993A4223546. (accessed 15 April 2021).

544 Mann, M.B., Prichula, J., de Castro, Í.M.S., Severo, J.M., Abadie, M., Lima, T.M.D.F.,
545 Caorsi, V., Borges-Martins, M., Frazzon, J., Frazzon, A.P.G., 2021. The oral
546 bacterial community in *Melanophryniscus admirabilis* (Admirable red-belly toads):
547 Implications for conservation. *Microorganisms* 9, 1–11.
548 <https://doi.org/10.3390/microorganisms9020220>

549 McCoy, K.A., Peralta, A.L., 2018. Pesticides Could Alter Amphibian Skin
550 Microbiomes and the Effects of *Batrachochytrium dendrobatidis*. *Front. Microbiol.*
551 9, 748. <https://doi.org/10.3389/fmicb.2018.00748>

552 Pimentel, D., Acquay, H., Biltonen, M., Rice, P., Silva, M., Nelson, J., Lipner, V.,
553 Giordano, S., Horowitz, A., D'Amore, M., 1992. Environmental and Economic
554 Costs of Pesticide Use. *Bioscience* 42, 750–760. <https://doi.org/10.2307/1311994>

555 Pollock, K.H., 1982. A Capture-Recapture Design Robust to Unequal Probability of
556 Capture. *J. Wildl. Manage.* 46, 752. <https://doi.org/10.2307/3808568>

557 Portaria MMA N° 350, de 23 de julho de 2019, 2019. 2º Ciclo do Plano de Ação
558 Nacional para Conservação de Répteis e Anfíbios Ameaçados da Região Sul do
559 Brasil – PAN Herpetofauna do Sul. Instituto Chico Mendes de Conservação da
560 Biodiversidade, Ministério do Meio Ambiente, Brasil.

561 Portaria MMA No 444, de 17 de dezembro de 2014, 2014. Lista Nacional Oficial de
562 Espécies da Fauna Ameaçadas de Extinção. Ministério do Meio Ambiente, Brasil.

563 Resolução CONSEMA N° 388, de 08 de novembro de 2018, 2018. Critérios e Diretrizes
564 para o Licenciamento Ambiental de Pequenas Centrais Hidrelétricas – PCHs, e
565 Centrais Geradoras Hidrelétricas – CGHs. Conselho Estadual de Meio Ambiente,
566 Estado do Rio Grande do Sul, Brasil.

567 Rio Grande do Sul, G. do, 2014. Espécies da Fauna Silvestre Ameaçadas de Extinção
568 no Estado do Rio Grande do Sul. Decreto 51.797, de 08 de setembro de 2014.
569 Estado do Rio Grande do Sul, Brazil.

570 Salafsky, N., Margoluis, R., 1999. Threat reduction assessment: A practical and cost-
571 effective approach to evaluating conservation and development projects. *Conserv.*
572 *Biol.* 13, 830–841. <https://doi.org/10.1046/j.1523-1739.1999.98183.x>

573 Salafsky, N., Salzer, D., Stattersfield, A.J., Hilton-Taylor, C., Neugarten, R., Butchart,
574 S.H.M., Collen, B., Cox, N., Master, L.L., O'Connor, S., Wilkie, D., 2008. A
575 standard lexicon for biodiversity conservation: unified classifications of threats and
576 actions. *Conserv. Biol.* 22, 897–911. <https://doi.org/10.1111/j.1523-1739.2008.00937.x>
577 <https://doi.org/10.1111/j.1523-1739.2008.00937.x>

578 Scott Jr., N.J., Woodward, B.D., 1994. Standard techniques for inventory and
579 monitoring: surveys at breeding sites. In: Heyer, W.R., Donnelly, M.A.,
580 McDiarmid, R.W., Hayek, L.C., Foster, M.S. (Eds.), *Measuring and Monitoring*
581 *Biological Diversity, Standard Methods for Amphibians*. Smithsonian Institution
582 Press, Washington DC, pp. 92–96.

583 Stuart, S.N., Chanson, J.S., Cox, N.A., Young, B.E., Rodrigues, A.S.L., Fischman,
584 D.L., Waller, R.W., 2004. Status and trends of amphibian declines and extinctions
585 worldwide. *Science*. 306, 1783–1786. <https://doi.org/10.1126/science.1103538>

586 TNC (The Nature Conservancy), 2007. *Conservation Action Planning Handbook*. The
587 Nature Conservancy, Arlington, VA, USA.

588 Wren, S., Angulo, A., Meredith, H., Kielgast, J., Dos Santos, M., Bishop, P., 2015.
589 *Amphibian Conservation Action Plan [WWW Document]*. IUCN SSC Amphib.
590 Spec. Group. URL <https://www.iucn-amphibians.org/resources/acap/>

591 Young, B.E., Lips, K.R., Reaser, J.K., Ibáñez, R., Salas, A.W., Rogelio Cedeño, J.,
592 Coloma, L.A., Ron, S., Marca, E. La, Meyer, J.R., Muñoz, A., Bolaños, F., Chaves,
593 G., Romo, D., 2001. Population declines and priorities for amphibian conservation
594 in Latin America. *Conserv. Biol.* <https://doi.org/10.1046/j.1523-1739.2001.00218.x>

Considerações Finais

Nesta tese, apresento informações inéditas sobre ecologia e conservação do sapinho-admirável-de-barriga-vermelha *Melanophryniscus admirabilis*, uma espécie microendêmica e ameaçada. No primeiro capítulo, nós demonstramos que a espécie apresenta reprodução sazonal e explosiva e que parece ter desenvolvido adaptações para lidar com o ambiente barulhento e arriscado em que vive. Uma dessas possíveis adaptações é a modulação de frequência do canto A da espécie (GOUTTE *et al.*, 2018), primeira vez registrada para o gênero *Melanophryniscus*. Nós registramos que os ovos são depositados em pequenas porções (média de 15,4 ovos por desova; n= 17), sendo cada porção em uma poça temporária diferente. Juntamente com o rápido desenvolvimento dos girinos (em torno de 21 dias desde a oviposição até a metamorfose completa), essas podem ser adaptações ao ambiente efêmero das pequenas poças em beira de rio, que rapidamente seca (devido à exposição ao sol) e frequentemente alaga (tanto pelas cheias do rio, como diretamente pela água da chuva que contribui para que as poças transbordem). A energia depositada em uma rápida metamorfose pode contribuir para que o girino de *M. admirabilis* seja um dos menores já descritos para o gênero, apesar dos adultos serem um dos maiores. Nós também registramos a maior longevidade diretamente observada em *Melanophryniscus*: 7 anos (25 indivíduos) e 9 anos (3 indivíduos). Esperamos que as espécies do gênero sejam anfíbios de vida longa, uma vez que (1) o registro de nove anos de longevidade foi ocasional (não analisamos todo o banco de dados), que (2) os indivíduos recapturados sete anos depois já eram adultos na primeira captura, que (3) nossa janela temporal de amostragem sistemática também tem sete anos, e que (4) Jeckel et al. (2015) registraram por osteocronologia uma variação de 1 a 5 anos para uma fêmea de *M. moreirae* se tornar adulta. Discutimos que as defesas químicas

(GRANT *et al.*, 2012) e a suposta coloração aposemática (BONANSEA; VAIRA, 2012; mas veja BORDIGNON *et al.*, 2018), observada no gênero possam desempenhar um papel importante na determinação dessa inesperada longevidade para um anuro pequeno, subtropical e diurno (STARK; MEIRI, 2018). Nós também registramos dimorfismo sexual de tamanho, sendo fêmeas maiores que machos, e alta fidelidade de sítio, sendo os jovens mais propensos a se deslocar ao longo do sítio reprodutivo. A partir do conhecimento produzido neste capítulo da tese, diversas outras perguntas ecológicas e evolutivas surgiram e fica evidente o quão importante e interessante são os estudos sobre a história natural das espécies.

No segundo capítulo, estimamos o tamanho populacional anual da única população conhecida da espécie e a probabilidade de sobrevivência aparente. Nossas estimativas de abundância variaram mais entre os métodos no mesmo ano do que entre anos. As estimativas de POPAN e do Desenho Robusto ficaram mais próximas entre si do que as estimadas pelo Modelo Fechado. Porém, a baixa recaptura de fêmeas influenciou diretamente na precisão das estimativas, principalmente no método POPAN. Consideramos o Desenho Robusto mais adequado para estimar abundância ao longo dos anos, já que ele considera toda a janela temporal em uma mesma análise. As estimativas para esse método variaram de 747 ± 49 , em 2014, a 1541 ± 87 indivíduos adultos, em 2017. Discutimos que essa importante flutuação do tamanho populacional no meio do período (2013 e 2014) pode estar relacionada à estiagem de 2012. Nesses intervalos, a sobrevivência dos indivíduos adultos também foi menor do que em outros anos. Com base nessas estimativas, levantamos a hipótese de que a estiagem pode ter contribuído principalmente por ter sido um ano de pouca atividade reprodutiva e, provavelmente, muita mortalidade de ovos e girinos por dessecação das poças. Consequentemente isso contribuiria para o menor recrutamento de indivíduos maduros nos anos imediatamente

posteriores, já que esperamos que os indivíduos levem ao menos um ano desde a metamorfose para se tornarem adultos reprodutivos (JECKEL; SAPORITO; GRANT, 2015). Por fim, concluímos que, considerando os outros pequenos sítios reprodutivos não amostrados neste trabalho, a população global da espécie não passaria de 2 mil indivíduos adultos reprodutivos nos anos das maiores estimativas.

No terceiro e último capítulo, nós usamos as informações sobre abundância, produzida no capítulo II, a distribuição geográfica e o levantamento das ameaças para reavaliar o estado de conservação de *M. admirabilis*. Usamos o conhecimento adquirido no primeiro capítulo da tese para identificar as ameaças e como elas se relacionam com a biologia da espécie. A partir dessa lista de ameaças, usamos uma metodologia objetiva para ranquear desde as de menor importância até as mais relevantes, usando três parâmetros: abrangência, intensidade e urgência da ameaça. Com base nesse método, identificamos que a presença do fungo quitrídio na população e os agrotóxicos utilizados nas plantações de fumo e soja são as ameaças mais preocupantes atualmente. A ameaça inferida causada pela construção da hidrelétrica foi unanimemente classificada como a que causaria danos mais severos e teria a maior abrangência. No entanto, atualmente é uma ameaça controlada, já que existe uma diretriz ambiental que proíbe a construção de barragens hidrelétricas nas cabeceiras do rio (Resolução Consema, Nº 388, 2018). Além disso, nesse capítulo, nós mostramos que mesmo após o novo registro de ocorrência da espécie, a área de ocupação (AOO) não passa dos 4 km². O risco de extinção da espécie pôde ser avaliado com base nos critérios B, C e D da IUCN (IUCN STANDARDS AND PETITIONS COMMITTEE, 2019), podendo ser categorizada como Criticamente em Perigo (CR), Em Perigo (EN) e Vulnerável (VU), respectivamente. No entanto, pelo princípio da precaução adotado pela IUCN, a espécie permaneceria categorizada como CR B1ab(iii) + 2ab(iii). A partir disso, discutimos que as ações prioritárias para

conservação seriam (1) a proteção legal da área e (2) a mudança no uso da terra, incentivando e subsidiando a substituição das monoculturas de fumo, soja e eucalipto, por agroflorestas de erva-mate nativa. Ainda assim, a distribuição da espécie é tão pequena que, embora essas ações citadas sejam fundamentais para a redução do risco de extinção, *M. admirabilis* somente poderia ser removido das listas de espécies ameaçadas caso uma nova população fosse encontrada. Então, sugerimos que é tão importante continuar buscando a espécie em outros locais, usando outras metodologias como DNA ambiental ou gravadores passivos, quanto é necessário que seja mantido o monitoramento na área, a fim de acompanhar as tendências populacionais e de qualidade do hábitat.

Gostaria de aproveitar o espaço, nas considerações finais desta tese, para expor minhas conclusões sobre os cinco anos de doutorado no Programa de Pós-Graduação em Ecologia da Universidade Federal do Rio Grande do Sul. O aprendizado ao longo desse período vai muito além do que está previsto nas seções deste documento. Além dos artigos aqui apresentados (e dos que estão por vir), considero que alcancei objetivos importantes para a minha formação como pesquisadora e conservacionista. O trabalho de conservação que venho desenvolvendo com o sapinho-admirável-de-barriga-vermelha foi reconhecido nacional e mundialmente. Em 2018, durante o I Simpósio Brasileiro de Conservação de Anfíbios – ANFoCO, recebi o Prêmio Jovem Conservacionista, conferido pelo Grupo de Especialistas em Anfíbios do Brasil (ASG Brasil - *Amphibian Specialist Group Brazil*) e Sociedade Brasileira de Herpetologia (SBH). Dois anos depois, recebi bolsa parcial para apresentar no Simpósio de Pesquisa em Conservação de Anfíbios (ACRS - *Amphibian Conservation Research Symposium*), promovido pela Aliança para a Sobrevivência dos Anfíbios (ASA - *Amphibian Survival Alliance*) durante o Congresso Mundial de Herpetologia (Nova Zelândia, 2020), o trabalho de conservação que desenvolvemos com a espécie: *‘Victory against all odds: the successful conservation*

story of the Admirable Red-Belly Toad in Brazil (“Vitória contra todas as probabilidades: a bem-sucedida história de conservação do sapinho-admirável-de-barriga-vermelha”). Lá também recebi o prêmio de Futura Líder em Conservação de Anfíbios (*Future Leader of Amphibian Conservation*), conferido pela ASA. Além disso, durante o doutorado, coorientei alunos de graduação (e.g. RIBEIRO, 2017), participei de outros estudos acadêmicos (BORDIGNON, 2019; BORDIGNON *et al.*, 2018; CAORSI *et al.*, *in prep*), dei palestras, fui banca examinadora de Trabalho de Conclusão de Curso, revisei artigos em periódicos, participei da organização de eventos, participei (participo) como articuladora de ação do PAN Herpetofauna do Sul, fui representante discente durante dois anos e, no último ano de doutorado, iniciei meu trabalho no Centro de Pesquisa e Conservação de Répteis e Anfíbios do Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio/RAN). Todas essas experiências extracurriculares, com certeza, fortalecem e diferenciam a minha formação.

Referências Bibliográficas

BISHOP, P. J. *et al.* The Amphibian Extinction Crisis - what will it take to put the action into the Amphibian Conservation Action Plan? **Sapiens**, [s. l.], v. 5, n. 2, p. 97–111, 2012.

BONANSEA, Maria Ines; VAIRA, Marcos. Geographic and intrapopulational variation in colour and patterns of an aposematic toad , *Melanophryniscus rubriventris*. [s. l.], v. 33, p. 11–24, 2012. Disponível em: <https://doi.org/10.1163/156853811X619754>

BORDIGNON, Debora Wolff *et al.* Are the unken reflex and the aposematic colouration of red-bellied toads efficient against bird predation? **PLoS ONE**, [s. l.], v. 13, n. 3, p. e0193551, 2018. Disponível em: <https://doi.org/10.1371/journal.pone.0193551>. Acesso em: 23 fev. 2021.

CARREIRA, Santiago; MANEYRO, Raúl. **Lista Roja de los Anfibios y Reptiles del Uruguay**. [S. l.: s. n.], 2015.

CONDE, Dalia A. *et al.* Data gaps and opportunities for comparative and conservation biology. **Proceedings of the National Academy of Sciences**, [s. l.], v. 116, n. 19, p. 9658–9664, 2019. Disponível em: <https://doi.org/10.1073/pnas.1816367116>

DI-BERNARDO, Marcos; MANEYRO, Raúl; GRILLO, Hamilton. New Species of *Melanophryniscus* (Anura : Bufonidae) from Rio Grande do Sul , Southern Brazil. **Journal of Herpetology**, [s. l.], v. 40, n. 2, p. 261–266, 2006.

FONTE, Luis Fernando Marin da *et al.* The Times they are a-Changing: How a MultiInstitutional Effort Stopped the Construction of a Hydroelectric Power Plant that Threatened a Critically Endangered Red-Belly Toad in Southern Brazil. **FrogLog**, [s. l.], v. 22, n. 4, p. 18–21, 2014.

FROST, Darrel R. **Amphibian Species of the Word: an Online Reference**. [S. l.], 2021. Disponível em: <https://doi.org/doi.org/10.5531/db.vz.0001>. Acesso em: 10 mar. 2021.

GARRAFFO, H Martin *et al.* Alkaloids from single skins of the Argentinian toad *Melanophryniscus rubriventris* (ANURA, BUFONIDAE): An unexpected variability in

alkaloid profiles and a profusion of new structures. **SpringerPlus**, [s. l.], v. 1, n. 1, p. 51, 2012. Disponível em: <https://doi.org/10.1186/2193-1801-1-51>

GASTON, Kevin J. **Rarity**. [S. l.: s. n.], 1994. ISSN 1098-6596. Disponível em: <https://doi.org/10.1007/978-94-011-0701-3>

GASTON, Kevin J.; BLACKBURN, Tim M. **Pattern and process in macroecology**. [S. l.]: wiley, 2007. Disponível em: <https://doi.org/10.1002/9780470999592>

GOUTTE, S. *et al.* How the environment shapes animal signals: a test of the acoustic adaptation hypothesis in frogs. **Journal of Evolutionary Biology**, [s. l.], v. 31, n. 1, p. 148–158, 2018. Disponível em: <https://doi.org/10.1111/jeb.13210>. Acesso em: 22 fev. 2021.

GRANT, Taran *et al.* The occurrence of defensive alkaloids in non-integumentary tissues of the Brazilian red-belly toad *Melanophryniscus simplex* (Bufonidae). **Chemoecology**, [s. l.], v. 22, n. 3, p. 169–178, 2012. Disponível em: <https://doi.org/10.1007/s00049-012-0107-9>

INCHAUSTI, Pablo; HALLEY, John. On the relation between temporal variability and persistence time in animal populations. **Journal of Animal Ecology**, [s. l.], v. 72, n. 6, p. 899–908, 2003. Disponível em: <https://doi.org/10.1046/j.1365-2656.2003.00767.x>

INSTRUÇÃO NORMATIVA MMA Nº 3, de 27 de maio de 2003. **Lista das Espécies da Fauna Brasileira Ameaçadas de Extinção** Brasil: Ministério do Meio Ambiente, 2003.

IUCN. **The IUCN Red List of Threatened Species**. [S. l.], 2021. Disponível em: <https://www.iucnredlist.org>.

IUCN STANDARDS AND PETITIONS COMMITTEE. Guidelines for Using the IUCN Red List Categories and Criteria. [s. l.], v. 14, n. August, 2019. Disponível em: <https://doi.org/http://www.iucnredlist.org/documents/RedListGuidelines.pdf>.

JECKEL, Adriana M.; SAPORITO, Ralph a.; GRANT, Taran. The relationship between poison frog chemical defenses and age, body size, and sex. **Frontiers in Zoology**, [s. l.], v. 12, n. 1, p. 27, 2015. Disponível em: <https://doi.org/10.1186/s12983-015-0120-2>

MARQUES, Ana Alice B. *et al.* **Lista de Referência da Fauna Ameaçada de Extinção no Rio Grande do Sul. Decreto no 41.672, de 11 junho de 2002.** Porto Alegre: FZB/MCT-PUCRS/PANGEA: Publicações Avulsas FZB, 11, 2002.

OTIS, David L *et al.* Statistical inference from capture data on closed animal populations. **Wildlife monographs**, [s. l.], n. 62, p. 3–135, 1978.

PIMM, S. L. *et al.* The biodiversity of species and their rates of extinction, distribution, and protection. **Science**, [s. l.], v. 344, n. 6187, 2014. Disponível em: <https://doi.org/10.1126/science.1246752>

POLLOCK, Kenneth H. A Capture-Recapture Design Robust to Unequal Probability of Capture. **The Journal of Wildlife Management**, [s. l.], v. 46, n. 3, p. 752, 1982. Disponível em: <https://doi.org/10.2307/3808568>

PORTARIA MMA Nº 444, de 17 de dezembro de 2014. **Lista Nacional Oficial de Espécies da Fauna Ameaçadas de Extinção** Brasil: Ministério do Meio Ambiente, 2014. Disponível em: https://www.icmbio.gov.br/cepsul/images/stories/legislacao/Portaria/2014/p_mma_444_2014_lista_especies_amecadas_extincao.pdf

RABINOWITZ, D. Seven forms of rarity. **The biological aspects of rare plant conservation**, [s. l.], p. 205–217, 1981. Disponível em: <https://doi.org/10.1177/004728702237415>

RIBEIRO, Talita Menger. **Fotoidentificação em Liolaemus arambarensis (SAURIA, LIOLAEMIDAE), lagarto ameaçado de extinção endêmico das restingas da Lagoa dos Patos, sul do Brasil.** 2017. - Universidade Federal do Rio Grande do Sul, UFRGS, [s. l.], 2017.

SCHWARZ, Carl James; ARNASON, A. Neil. A General Methodology for the Analysis of Capture-Recapture Experiments in Open Populations. **Biometrics**, [s. l.], v. 52, n. 3, p. 860, 1996. Disponível em: <https://doi.org/10.2307/2533048>

STARK, Gavin; MEIRI, Shai. Cold and dark captivity: Drivers of amphibian longevity. **Global Ecology and Biogeography**, [s. l.], v. 27, n. 11, p. 1384–1397, 2018. Disponível em: <https://doi.org/10.1111/geb.12804>. Acesso em: 1 abr. 2021.

STUART, Simon N. *et al.* Status and trends of amphibian declines and extinctions worldwide. **Science**, [s. l.], v. 306, n. 5702, p. 1783–1786, 2004. Disponível em: <https://doi.org/10.1126/science.1103538>. Acesso em: 21 abr. 2021.

TOLEDO, Luís Felipe *et al.* Rarity as an indicator of endangerment in neotropical frogs. **Biological Conservation**, [s. l.], v. 179, p. 54–62, 2014. Disponível em: <https://doi.org/10.1016/j.biocon.2014.08.012>

VAIRA, Marcos *et al.* Categorización del estado de conservación de los anfibios de la República Argentina. **Cuadernos de herpetología**, [s. l.], v. 26, n. 1, p. 131–159, 2012.

VILLALOBOS, Fabricio *et al.* Is Rich and Rare the Common Share? Describing Biodiversity Patterns to Inform Conservation Practices for South American Anurans. **PLoS ONE**, [s. l.], v. 8, n. 2, p. e56073, 2013. Disponível em: <https://doi.org/10.1371/journal.pone.0056073>

ZANK, Caroline *et al.* Climate change and the distribution of neotropical red-bellied toads (*Melanophryniscus*, Anura, Amphibia): How to prioritize species and populations? **PLoS ONE**, [s. l.], v. 9, n. 4, p. 1–11, 2014. Disponível em: <https://doi.org/10.1371/journal.pone.0094625>